

Volatile Constituents of *Calamintha organifolia* Boiss. Growing Wild in Lebanon

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The essential oil of aerial parts of *Calamintha organifolia* Boiss. (Lamiaceae), growing wild in Lebanon, was obtained by hydrodistillation and was analysed by GC and GC-MS. 49 compounds, representing 92.2% of the oil, were identified. The major components, belonging to the class of oxygenated monoterpenes, were pulegone (22.5%), isomenthone (12.2%) and piperitenone (9.6%). The oil showed a slight antimicrobial activity against three bacterial strains.

Keywords: *Calamintha organifolia*, essential oil, GC-MS, oxygenated monoterpenes, pulegone, isomenthone, piperitenone.

Calamintha (syn. *Cyclotrichium*) is a genus of about thirty species that belongs to the tribe Menthae, subfamily Nepetoideae, family Lamiaceae. It is native to the northern temperate regions of Europe and Asia. According to Marin *et al.* [1], the genus *Calamintha* Miller is closely related to *Micromeria* Benth, *Satureja* L., *Clinopodium* L. and *Acinos* Miller, and for this reason the use of chemotaxonomic markers is essential to better differentiate these genera.

Many *Calamintha* species are used as spices in various culinary recipes because of their pleasant mint-like smell. Besides, they are known for different medicinal uses. Common calamint is used as diaphoretic, in syrups for coughs and colds and as an expectorant. The tea is used to help with gas and colic [2]. Externally, it is useful in poultices for bruises and as a strengthener and nerve soother. The essential oil shows different activities. The oil of *C. sylvatica* subsp. *ascendens* exerts significant sedating and antipyretic activities in the rat, due to the presence of the monoterpenes pulegone, menthone and eucalyptol [3]. Monoterpenes,

particularly pulegone and isopulegone, are also reported to be the responsible of the strong antibacterial and antifungal activities showed by essential oils from different *Calamintha* species [4]. Due to its good antimicrobial activity, *C. officinalis* essential oil has been proved to be effective as preservative in two current formulations (cream and shampoo) [5].

Calamintha organifolia Boiss. (syn. *Cyclotrichium organifolium* (Labill.) Manden & Scheng.) is a strongly aromatic, suffruticose, much branched species wild growing in the Horsh Ehden reserve that is located on the northern part of the Lebanese western mountain range, just below Cornet As Sawda, the highest mountain peak in Lebanon. The Reserve represents a mountainous ecosystem on the elevated slopes of the northern Mt. Lebanon chain. In this paper, as a continuation of our studies on the essential oils from Lamiaceae growing wild in Lebanon [6], we report on the chemical composition of the essential oil of *Calamintha organifolia* collected in the Lebanese Horsh Ehden reserve.

Table 1: Essential oil composition of *Calamintha origanifolia* Boiss.

| I ^a | I ^b | Component | Method ^c | % ^d |
|-------------------------|----------------|----------------------------------|---------------------|----------------|
| 798 | | Hexanal | I,MS | 0.1 |
| 930 | | α -Thujene | I,MS | t |
| 963 | 1543 | Benzaldehyde | I,MS,Co-GC | 0.3 |
| 973 | 1132 | Sabinene | I,MS | 0.1 |
| 980 | 1118 | β -Pinene | I,MS,Co-GC | 0.3 |
| 1025 | 1280 | <i>p</i> -Cymene | I,MS,Co-GC | 0.2 |
| 1030 | 1203 | Limonene | I,MS,Co-GC | 0.6 |
| 1034 | 1213 | 1,8-Cineole | I,MS,Co-GC | 0.6 |
| 1111 | | <i>p</i> -Mentha-1,3,8-triene | I,MS | 0.7 |
| 1117 | 1152 | <i>trans-p</i> -Menth-2-en-1-ol | I,MS | 0.9 |
| 1125 | 1540 | Chrysanthenone | I,MS | 0.9 |
| 1138 | 1475 | Menthone | I,MS,Co-GC | 7.7 |
| 1145 | 1663 | <i>cis</i> -Verbenol | I,MS | 1.9 |
| 1163 | 1502 | Isomenthone | I,MS | 12.2 |
| 1175 | 1582 | Isopulegone [#] | I,MS | 5.8 |
| 1177 | 1755 | Dihydrocarveol | I,MS | 0.1 |
| 1182 | 1652 | Menthol | I,MS,Co-GC | 0.7 |
| 1233 | 1662 | Pulegone | I,MS,Co-GC | 22.5 |
| 1244 | 1750 | Carvone | I,MS | 1.5 |
| 1293 | 2198 | Thymol | I,MS,Co-GC | 0.8 |
| 1299 | 2239 | Carvacrol | I,MS,Co-GC | 1.1 |
| 1329 | 1949 | Piperitenone | I,MS | 9.6 |
| 1343 | 1748 | Piperitone | I,MS | 6.9 |
| 1353 | 2186 | Eugenol | I,MS,Co-GC | 0.2 |
| 1363 | | Piperitenone oxide | I,MS | 0.7 |
| 1372 | 1493 | α -Ylangene | I,MS | 0.2 |
| 1377 | 1497 | α -Copaene | I,MS | 0.1 |
| 1382 | | β Cubebene | I,MS | t |
| 1385 | 1535 | β -Bourbonene | I,MS | 0.1 |
| 1387 | 1600 | β -Elemene | I,MS | 0.2 |
| 1415 | 1612 | Caryophyllene | I,MS,Co-GC | 1.9 |
| 1451 | 1868 | Geranyl acetone | I,MS | 0.3 |
| 1452 | 1673 | (<i>E</i>)- β -Farnesene | I,MS | t |
| 1455 | 1689 | α -Humulene | I,MS | 0.1 |
| 1477 | 1726 | Germaacrene D | I,MS | 0.1 |
| 1515 | 1776 | γ -Cadinene | I,MS | t |
| 1520 | 1839 | <i>cis</i> -Calamenene | I,MS | 0.2 |
| 1526 | | δ -Cadinene | I,MS | t |
| 1640 | 2187 | τ -Cadinol | I,MS | 4.0 |
| 1642 | 2209 | τ -Muurolol | I,MS | 0.7 |
| 1649 | 2255 | α -Cadinol | I,MS | 1.2 |
| 1835 | 2131 | Hexahydrofarnesylacetone | I,MS | 1.6 |
| 1957 | 2931 | Hexadecanoic acid | I,MS,Co-GC | 1.4 |
| 2500 | 2500 | Pentacosane | I,MS | 0.3 |
| 2600 | 2600 | Hexacosane | I,MS | 0.2 |
| 2700 | 2700 | Heptacosane | I,MS | 1.0 |
| 2800 | 2800 | Octacosane | I,MS | 0.1 |
| 2900 | 2900 | Nonacosane | I,MS | 1.3 |
| 3100 | 3100 | Hentriacontane | I,MS | 0.8 |
| Total identified | | | | 92.2 |

^a: HP-5 MS column; ^b: HP Innowax; ^c: *I* is the retention index, MS = mass spectrum, Co-GC = co-injection with authentic compound; ^d: t = trace, less than 0.05%; [#]: correct isomer not identified.

Great variations occur in the volatile compounds from *Calamintha* genus, but the major components in the oils generally belong to the C-3 oxygenated *p*-menthanes such as pulegone, isomenthone, menthone, piperitone and piperitenone with their oxides [4a,4c-4f,5,7-9]. According to Baldovini *et al.* [8], three types of oils can be distinguished: in the first pulegone is the major component, associated with different compounds such as menthone and/or isomenthone, menthol and its isomers, piperitenone, piperitone and piperitenone oxides. The second type is characterized by the predominance of piperitone

Table 2: Antimicrobial activity of *Calamintha origanifolia* oil (C).

| Strain | MIC (MBC) μ g/mL | |
|---|----------------------|------|
| | C | Ch |
| <i>Bacillus subtilis</i> ATCC 6633 | 50 (100) | 12.5 |
| <i>Staphylococcus aureus</i> ATCC 25923 | 100 (>100) | 25 |
| <i>Staphylococcus epidermidis</i> ATCC 12228 | 25 (50) | 3.12 |
| <i>Streptococcus faecalis</i> ATCC 29212 | 100 | 25 |
| <i>Escherichia coli</i> ATCC 25922 | 50 (100) | 12.5 |
| <i>Klebsiella pneumoniae</i> ATCC 10031 | 100 | 50 |
| <i>Proteus vulgaris</i> ATCC 13315 | 100 (>100) | 25 |
| <i>Pseudomonas aeruginosa</i> ATCC 27853 | >100 | 100 |

Ch: Chloramphenicol

oxide and/or piperitenone oxide. Last type is distinguished by the presence of carvone and 1,8-cineole as main components [8 and references cited therein].

The essential oil of *C. origanifolia* belongs to the first type, as pulegone (22.5%) is the most abundant component. In total, forty-nine constituents have been identified; representing 92.2% of the total oil; their retention indices and percentage composition are given in Table 1, where the components are listed in order of elution from a HP 5MS column. As reported in the literature for other *Calamintha* species [9], the oxygenated monoterpenes were the most abundant components of the oil, particularly those with *p*-menthane skeleton, and their content represented 59.7% of the oil. The most abundant compounds of this fraction were pulegone (22.5%), isomenthone (12.2%) and piperitenone (9.6%). The high content of isomenthone can be considered a characteristic of the present oil because this compound is reported in lower amounts in other *Calamintha* oils. Isomenthone was detected in a quite similar extent only in the oils of *C. grandiflora* (15.2%) [9b] and *C. sylvatica* ssp. *sylvatica* in the pre-blossom phase (13.4%) [9e]. The greatest amount of isomenthone was detected in the oil of *C. sylvatica* ssp. *ascendens* (36.8-43.3%) [9f]. Other ketones identified in the oil were chrysanthenone (0.9%), geranyl acetone (0.3%) and hexahydrofarnesyl acetone (1.6%). Also a few monoterpene hydrocarbons were present but they represented only 1.9% of the oil, ranging between 0.7% (*p*-mentha-1,3,8-triene) and traces (α -thujene). Twelve sesquiterpene hydrocarbons were detected. Caryophyllene represented the 1.9% of the oil whereas the other sesquiterpene hydrocarbons were present in low content, from traces to 0.2%. Three

oxygen-containing sesquiterpenes were present and τ -cadinol (4.0%) was the major component of this fraction. In the oil were also identified three phenols that amounted to the 2.1%. Carvacrol (1.1%) and thymol (0.8%) were the most abundant while eugenol represented the 0.2% of the oil. Data obtained allow us to ascribe the oil of *Calamintha organifolia* Boiss. growing wild in Lebanon to a type pulegone/isomenthone oil.

The MIC and MBC values of the essential oil against eight selected micro-organisms are reported in Table 2. The oil showed action mainly against *B. subtilis*, *S. epidermidis* and *E. coli*.

Experimental

Plant material: Aerial parts of *C. organifolia* Boiss were collected at the full flowering stage from plants growing wild on rocky soil at Oyoun Ouvghanch, 2200 m a.s.l., in June 2005. The required authorizations for the plant collection were given by the Lebanese authorities to Apostolides Arnold. A voucher specimen (leg. & det. N. Arnold s. n., confirm. Th. Raus) was deposited in the Herbarium of the Botanischer Garten, Berlin Universität.

Essential oil isolation: The oil from air-dried and ground aerial parts of plants was isolated by hydrodistillation for 3 h, using a Clevenger-type apparatus according to the method recommended in the *European Pharmacopoeia* [10]

The oil was dried over anhydrous sodium sulphate and stored under N₂ at +4°C in the dark until tested and analysed. The sample yielded 0.13% of yellow oil (w/w), with a pleasant smell of mint.

GC analysis: Analytical gas chromatography was carried out on a Perkin-Elmer Sigma 115 gas chromatograph fitted with a HP-5 MS capillary column (30 m x 0.25 mm i.d.; 0.25 μ m film thickness). Helium was the carrier gas (1 mL min⁻¹). Column temperature was initially kept at 40°C for 5 min, then gradually increased to 250°C at 2°C min⁻¹, held for 15 min and finally raised to 270°C at 10°C min⁻¹. Diluted samples (1/100 v/v, in *n*-hexane) of 1 μ L were injected manually at 250°C, and in the splitless mode. Flame ionization detection (FID) was performed at 280°C. Analysis was also run by using a

fused silica HP Innowax polyethylenglycol capillary column (50 m x 0.20 mm i.d.; 0.20 μ m film thickness).

GC-MS analysis: GC-MS analysis was performed on an Agilent 6850 Ser. II apparatus, fitted with a fused silica HP-1 capillary column (30 m x 0.25 mm i.d.; 0.33 μ m film thickness), coupled to an Agilent Mass Selective Detector MSD 5973; ionization voltage 70 eV; electron multiplier energy 2000 V. Gas chromatographic conditions were as reported above; transfer line temperature, 295°C.

Qualitative and quantitative analyses: Most constituents were identified by gas chromatography by comparison of their retention indices (*I*) with either those of the literature [11,12] or with those of authentic compounds available in our laboratories. The retention indices were determined in relation to a homologous series of *n*-alkanes (C₈-C₂₄) under the same operating conditions. Further identification was made by comparison of their mass spectra on both columns with either those stored in NIST 02 and Wiley 275 libraries or with mass spectra from the literature [11,13] and our home made library. Component relative concentrations were calculated based on GC peak areas without using correction factors.

Antimicrobial activity: The antibacterial activity was evaluated by determining the minimum inhibitory concentration (MIC) and the minimum bactericidal concentration (MBC) using the broth dilution method as previously described [6e]. Eight bacteria species, selected as representative of the class of Gram positive and Gram negative, were tested: *Bacillus subtilis* (ATCC 6633), *Staphylococcus aureus* (ATCC 25923), *Staphylococcus epidermidis* (ATCC 12228), *Streptococcus faecalis* (ATCC 29212), *Escherichia coli* (ATCC 25922), *Klebsiella pneumoniae* (ATCC 10031), *Proteus vulgaris* (ATCC 13315) and *Pseudomonas aeruginosa* (ATCC 27853).

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