



## A redescription of the Mediterranean endemic cladoceran *Daphnia chevreuxi* Richard, 1896 (Cladocera: Daphniidae)

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### Abstract

The Mediterranean area is a biodiversity and endemism hotspot. “Circum-Mediterranean” taxa are known among different hydrobionts, including the water fleas. Some Mediterranean endemic cladoceran taxa have been described or redescribed according to modern taxonomical standards, but accurate drawings are missing for others. Here we redescribe the Mediterranean endemic *Daphnia chevreuxi* Richard, 1896 (Crustacea: Cladocera) and briefly review available data on its distribution and ecology. The species is confirmed to be a typical inhabitant of the temporary ponds of the central Mediterranean area, whereas its populations from the eastern Balkans and the Middle East should be studied in order to check for their actual identity. We conclude that the Mediterranean area is an example of a well-studied region as Cladocera are concerned, but the study of other regions is necessary in order to understand better the cladoceran diversity and distribution patterns in Eurasia.

**Key words:** Cladocera, taxonomy, morphology, description, Mediterranean area

### Introduction

Several parts of the Palearctic biogeographical region are biodiversity hotspots, and the Mediterranean area is among these (Myers *et al.* 2000; Ramos *et al.* 2001; Cuttelod *et al.* 2009). It is known that the biodiversity hotspots are not always congruent with the endemism hotspots (Orme *et al.* 2005), but the Mediterranean area is an exception from this rule, and it is also a well-known endemism zone for terrestrial and inland water species (Ramos *et al.* 2001; Reyjol *et al.* 2007; Cheikh Albassatneh *et al.* 2021); this is possibly due to its role as a refuge for European terrestrial and freshwater fauna during the coldest phases of the Pleistocene (Hewitt 2001; Stewart & Lister 2001). In fact, in spite of the semi-arid climate currently characterizing most of the area, which could be suggestive of a poorly differentiated inland water fauna, circum-Mediterranean endemic taxa (*sensu* Bănărescu 1990) are frequent among different hydrobionts (Bănărescu 1990; Blondel *et al.* 2010; Hermoso & Clavero 2011; Tierno de Figueroa *et al.* 2013; Marrone *et al.* 2014; Marrone *et al.* 2017).

Cladocera (Crustacea: Branchiopoda) biodiversity is relatively well-studied there as its investigation has been carried out since the 19th century (Lucas 1849; Richard 1887; Guerne & Richard 1986; Gurney 1909; Gauthier 1928; Parenzan 1932; Parenzan 1933). In several countries, the cladoceran fauna has also been intensively studied in the last decades; this is the case of Spain (Alonso 1985; Alonso 1990; Alonso 1991; Alonso 1996; Alonso 1998), mainland Italy (Margaritora 1983; Margaritora 1985; Margaritora 2005) and large Tyrrhenian islands (Margaritora 1970; Margaritora & Ferrara 1974; Margaritora *et al.* 1975; Marrone *et al.*, 2005; Marrone *et al.* 2006), Turkey (Margaritora *et al.* 1977; Güher 2014; Bozkurt & Aktas 2016), Israel (Bromley 1993), Algeria and Tunisia (Dumont 1979; Dumont *et al.* 1979; Samraoui 2002; Ghaouaci *et al.* 2018); conversely, recent synoptical data are lacking for other countries and regions. Useful synopses of the available data, bibliography and distribution for the circum-Mediterranean countries are those provided by Mouelhi *et al.* (2000) and Błędzki & Rybak (2016).

Several endemic cladoceran taxa from the Mediterranean area (including the Balkans) are adequately described or redescribed according to the current standards of morphological taxonomy (Alonso 1996; Alonso & Pretus 1989; Brancelj 1990; Sinev *et al.* 2012; Alonso *et al.* 2021), but accurate morphological characterizations are missing for others.

The circum-Mediterranean species of the genus *Daphnia* O.F. Müller (Anomopoda: Daphniidae) were a subject of special interest for the cladocerologists at the end of the 20th century (Alonso 1985; Alonso 1996; Glagolev & Alonso 1990), but such a program of morphological studies apparently came to an end, while some molecular investigations were done recently in frames of wide-range phylogenetic (Petrušek *et al.* 2008; Petrušek *et al.* 2009; Adamowicz *et al.* 2009) and phylogeographic (De Gelas & Meester 2005; Crease *et al.* 2012) investigations, or to check the identity of local populations belonging to cryptic species complex or invasive taxa (Marková *et al.* 2015; Vecchioni *et al.* 2021).

Among the daphniids of the Mediterranean area, *Daphnia chevreuxi* Richard, 1896 is a remarkable species typical of poorly mineralized temporary water ponds (Benzie 2005; Błędzki & Rybak 2016). After the species description from Algeria (Richard 1896), it was found in the whole Maghreb (Gurney 1909; Gauthier 1928; Ramdani 1988), in continental Mediterranean Europe including the Balkans (Stephanides 1948; Petkovski 1970; Flössner 1980; Negrea 1983; Naidenow 1994), in different large and small islands in the Mediterranean Sea (Stephanides 1948; Margaritora 1983; Margaritora *et al.* 1975; Błędzki & Rybak 2016; Marrone *et al.* 2019a), and in Israel (Bromley 1993), while in the Iberian Peninsula it is replaced by the closely-related *Daphnia hispanica* Glagolev & Alonso, 1990 (Alonso 1991; Alonso 1998). However, to date, just a few drawings of *Daphnia chevreuxi* are available (Richard 1896; Stephanides 1948; Glagolev 1995; Margaritora 1985; Benzie 2005), and these do not fulfil the modern standards of cladoceran morphological taxonomy.

The aim of this paper is, therefore, to redescribe the morphology of *D. chevreuxi*, i.e. based on numerous samples of J. Richard kept at the Smithsonian National Museum of Natural History, USA (Kotov & Ferrari 2010) and novel samples from Italy and North Macedonia, and review data on its distribution and ecology.

## Material and methods

Most samples studied here were collected by FM by means of a 125- $\mu$ m mesh-sized hand net, paying attention to sample in each microhabitat present in each site, and fixed in situ in 95% ethanol; some additional samples were obtained from our colleagues. Samples were initially examined under a stereoscopic dissecting microscope Leica MZ7.5. For morphological analysis, samples were placed in small Petri dishes; specimens were picked from them by pipettes, placed on slides in drops of glycerol, covered by coverslips and examined under a high-power microscope Olympus CX41. Ten parthenogenetic females, five adult males and two juvenile males (if present) and females from each sample were dissected for analysis of appendages. Drawings were made using *camera lucida*. Some specimens were lyophilised, attached to aluminum stubs, coated with gold in a S150A Sputter Coater, and studied under a Tescan Vega TS5130MM scanning electron microscope.

**Abbreviations in illustrations and text:** ejh = ejector hooks on limb I; epp = epipodite; ext = exopodite; idl = inner distal lobe of limb I; odl = outer distal lobe of limb I; pep = preepipodite.

**Abbreviations for collections:** DGF = Collection of David G. Frey, Smithsonian National Museum of Natural History, Museum Support Center in Suitland, Maryland, U.S.A. GLAG = personal collection of S.M. Glagolev gifted to the Laboratory of aquatic ecology and biological invasions of A.N. Severtsov Institute of Ecology and Evolution of Russian Academy of Sciences (IEE), Moscow, Russia. AAK = personal collection of A.A. Kotov stored at IEE.

## Results

### Order Anomopoda Sars, 1865

### Family Daphniidae Straus, 1820

## Genus *Daphnia* O.F. Müller, 1776

### Subgenus *Daphnia* (*Ctenodaphnia*) Dybowski & Grochowski, 1895

#### *Daphnia* (*Ctenodaphnia*) *chevreuxi* Richard, 1896

(Figs. 1–14)

*Daphnia chevreuxi* Richard, 1896: P. 206–209, Pl. 20, Figs. 10–11; Pl. 21, Fig. 4; Pl. 23, Fig. 18; Pl. 24, Fig. 4. Gauthier, 1928: P. 44–45, Fig. 15A–H. Petkovski, 1970: P. 139–142, Figs. 1–7. Flössner, 1980: P. 65–67, Fig. 3. Negrea, 1983: P. 104. Margaritora, 1983: P. 58–62, Fig. 37. Margaritora, 1985: P. 116–119, Figs. 49A–H. Glagolev and Alonso, 1990: P. 159–162. Glagolev, 1995: P. 53, Pl. 41, Figs. 1–5. Benzie, 2005: P. 125–128, Figs. 343–352. Kotov *et al.* 2010: P. 201, Fig. 117: 5–9. Korovchinsky *et al.*, 2021: P. 138–140, Fig. 41: 10–14.  
*Daphnia psittacea* Baird, 1850 in Stephanides 1948: P. 7–8, Pl. 9, Fig. 9–13.  
? *Daphnia byzantina* Muckle, 1951: P. 373–374, Fig. 2a–g.

**Type locality.** «Algérie: Environs de Bône (Guerrah El M’Krada, bord du lac Fetzara, marais des Kharézas, et abreuvoirs des environs de Bône)» (Richard 1896).

**Type material. Syntypes.** Many parthenogenetic, ephippial females and males in samples DGF 730 and DGF 767, “Environs de Bône, Abreuvoir”; DGF 779, “Environs de Bône. Dans une abreuvoir”, DGF 783; “Environs de Bône”; DGF 761, “Guerrah el M’Krada. Eau legerement salee”; DGF 797 “Au bord du Guerrah el M’Krada”, all from Algeria.

**Other material studied here. Italy: Sicily.** Many males, ephippial and parthenogenetic females from a swamp (37.85847°N, 12.92082°E), Margio di Gallitello (Calatafimi), coll. by F. Marrone in 02.03.2014, AAK M-5314. Parthenogenetic females from the same locality, coll. by Marrone in 30.11.2018, AAK M-6906. Many males, ephippial and parthenogenetic females from a pond (37.87225°N, 14.67638°E), Stagno di C. da Buffali (Nebrodi, Cesarò), coll. by F. Marrone in 16.05.2018, AAK M-6932. Many males, ephippial and parthenogenetic females from the same locality, coll. by F. Marrone on 19.05.2021, AAK M-6941. Few juvenile females from Gorgo di Gaetano (37.88559°N, 13.36919°E), coll. by F. Marrone in 08.03.2009, AAK M-5317. Parthenogenetic females from a pond (37.97351°N, 13.4936°E), Pozze di Bosco Tumminia (Bolognetta), coll. by F. Marrone in 09.11.2018, AAK M-6938. Many males, ephippial and parthenogenetic females from a pond (38.02833°N, 13.32666°E), Gorgo di Rebuttone (Altofonte), coll. by F. Marrone in 19.02.2021, AAK M-6943. Parthenogenetic females from a pond (38.10313°N, 12.67736°E), Gorgo di Baglio Cofano (Monte Cofano), coll. by F. Marrone in 19.12.2019, AAK M-6928. Parthenogenetic females from the same locality, coll. by F. Marrone in 11.12.2019, AAK M-6946. **North Macedonia.** Many males, ephippial and parthenogenetic females from Slavej (41.3°N, 21.4°E) coll. by T. Petkovski in 25.05.1985, GLAG040.

**Diagnosis.** Adult parthenogenetic female with body high for the subgenus (body height/length without shell spine = 0.56–0.62). Head shield with projected, angled-rounded fornices, a median anterior projection of carapace especially short for the subgenus, it penetrates only to about 1/5–1/6 of length of the head shield. Postabdomen obviously tapering distally. Numerous small anal teeth of subequal size located on anal portion, this row continues more laterally on preanal portion where it is accompanied by groups of smaller spinules. The first (proximal) and second pectens on outer face of postabdominal claw consisting of relatively strong teeth (the longest ones approximately as long as claw diameter); the third pecten consisting of somewhat shorter spines. Antenna I as a minute conical tubercle with nine terminal aesthetascs; tips of aesthetascs not projected beyond tip of rostrum. Limb I with accessory seta; outer distal lobe bearing a long seta distally armed with short setules, and a short second seta; inner distal lobe with a single, long anterior seta 1 armed distally with short setules. Limb II with inner-distal lobe bearing a thin, stiff anterior seta with length 3/4 of soft seta length, armed with minute setules distally. Limb V with exopod supplied with two distal setae and a large lateral seta.

Ephippium dark brown, elongated, bean-like; two eggs with axes located at a very acute angle or almost parallel to the dorsal margin; anterior processes present, postero-dorsal portion of valves (with shell spine) initially incorporated into ephippium. Sculpture of ephippium as a network of small protuberances having smooth tips oriented somewhat posteriorly.

Adult male with head having anteriormost extremity completely occupied with a very large optic vesicle; a shallow post-ocular depression present. Abdomen with a shallow mound on basal segment, other segments

without projections. Postabdomen tapering distally, its distal portion bent, ventral margin convex; gonopore opens subdistally, any genital papilla absent. Few anal teeth present only in basal portion of anal margin. Antenna I long, somewhat bent; length of flagellum less than half body antenna I length; distal segment of flagellum covered with short setules. Limb I with inner distal lobe bearing a single long seta (1) and a rudimentary seta 1'. Limb II inner distal portion with seta 1 remarkably stronger as in female, slightly bent and asymmetrically setulated, with a blunt tip. Limb V as in female.

Size of parthenogenetic females 0.9–3.8 mm, adult males 1.1–2.4 mm.

**Redescription. Adult parthenogenetic female.** *General.* Body almost transparent, high for the subgenus (body height/length without shell spine = 0.56–0.62), subovoid in lateral view, with maximum height in middle of valves (Figs. 1A, 2A–B, 3A–B). Dorsal margin slightly convex. Postero-dorsal angle with a moderately developed to long caudal spine projected posteriorly and somewhat dorsally (Figs. 1A, 2A–B, 3A–B, 4A), ventral margin regularly convex.

*Head* with a short, rounded rostrum (Figs. 1B, 2C–D, 3C–D); posterior margin of head slightly concave, without a projection, pre-rostral fold not expressed; antero-ventral margin usually almost straight, but rarely slightly concave; maximum body anterior extremity lies somewhat dorsally to body longitudinal midline. Head without any pre-ocular and post-ocular depressions, a very shallow depression present in posterior head portion, but this is not a border between head and valves. Any helmet fully absent in the adults. Compound eye relatively small, ocellus minute. Head shield with projected, angled-rounded fornicies; the median anterior projection of carapace especially short for the subgenus since it penetrates only to about 1/5–1/6 of length of the head shield (Fig. 2E–F). Labrum as a fleshy lobe with a large distal plate (Fig. 5A).

*Carapace* in general semi-ovoid, the free edge uniformly convex. A group of relatively long setae in middle of its ventral margin (Fig. 4A–C); short setae at postero-ventral and posterior margin, with setules between them (Fig. 4D–F).

*Abdomen* with the first (proximalmost) abdominal segment with a relatively short (but longer than postabdominal claw) process, slightly bent anteriorly; the second segment with a long process (also longer than postabdominal claw) bent posteriorly; the third segment with a massive process; the fourth process small. All processes covered by rows of minute setules (Figs. 1C–D, 4G–I).

*Postabdomen* elongated, obviously tapering distally. Postanal margin straight, preanal and postanal angle smooth. Numerous small anal teeth of subequal size located on anal portion, this row continues more laterally on preanal portion where it is accompanied by groups of smaller spinules (Figs. 1C, E, 4G–H). Postabdominal seta as long as preanal margin, its distal segment somewhat shorter than proximal segment. Postabdominal claw regularly bent, with a pointed tip. On its outer side, along the dorsal margin there are three pectens: the first (proximal) and second pectens consisting of relatively strong teeth, the longest ones about as long as the diameter of the claw base; the third pecten consisting of somewhat shorter spines, not reaching tip of claw (Figs 1E, 4J).

*Antenna I* as a minute conical tubercle with nine terminal aesthetascs; tips of aesthetascs not projected beyond tip of rostrum; antennular sensory setae small, arising from base of mound of antenna I and projecting beyond the mound (Fig. 5A–B).

*Antenna II* relatively long (Fig. 5C–D); coxal portion with two short setae (Fig. 5E–F); basal segment distally with a short anterior spine (Fig. 5D: arrow) and a longer posterior seta (Fig. 5G: arrow). A spine on second exopod segment short (Fig. 5D: arrow). Antennal formula: setae 1–1–3/0–0–1–3. Length of apical setae approximately equal to the length of the branches.

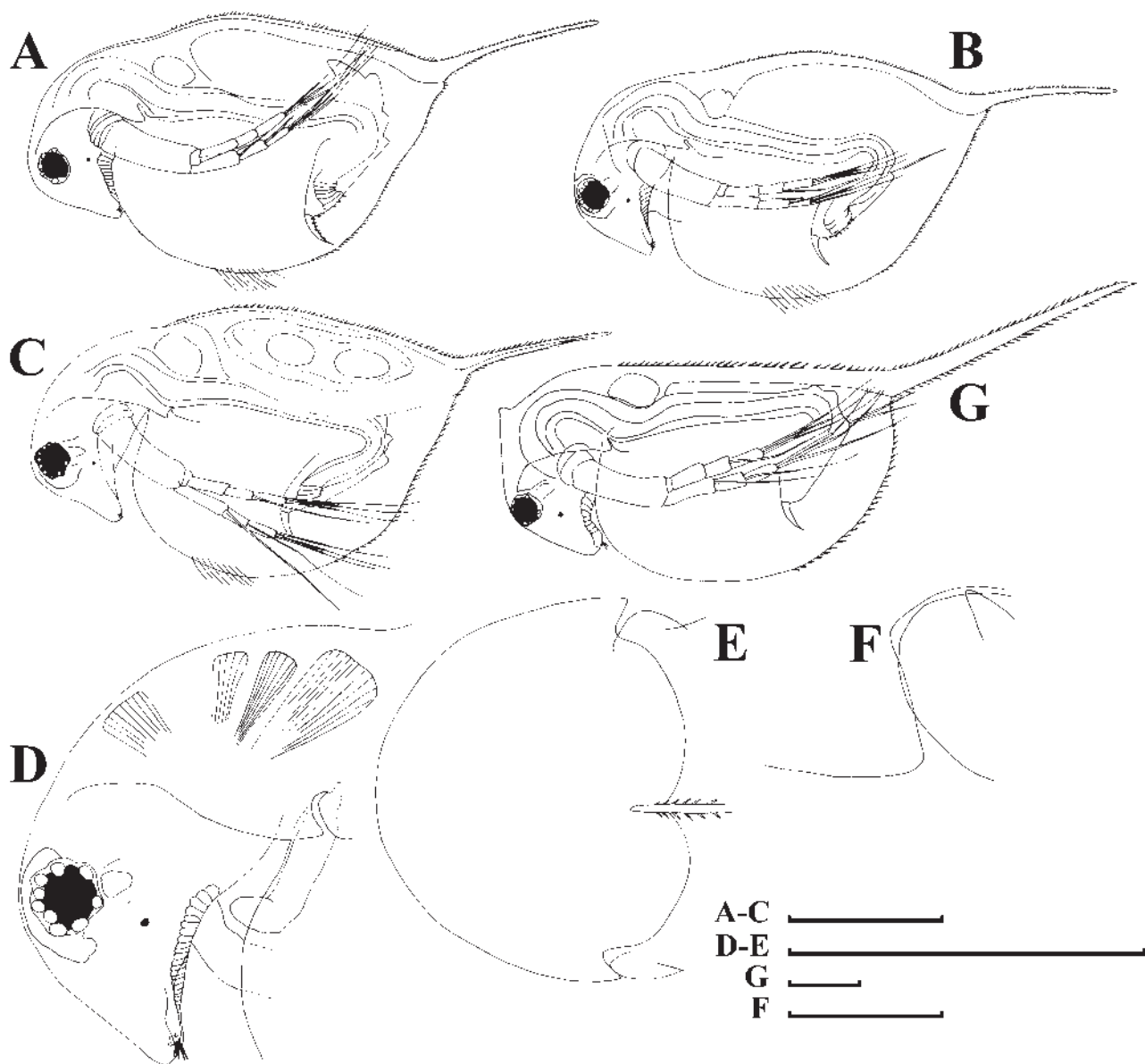
*Maxilla I* as a projection bearing three longer and a single shorter seta (Fig. 6A).

*Limb I* with accessory seta (Fig. 6C: acs). Outer distal lobe (Figs 1F, 6B–D: odl) cylindrical, with a long seta distally armed with short setules, and a short second seta. Endite 5 = inner distal lobe (Fig. 6C: idl) with a single, long anterior seta 1 armed distally with short setules. Endite 4 with a long anterior seta (Fig. 6C: 2) and two posterior setae (a–b). Endite 3 with a long anterior seta (Fig. 6C: 3) and two posterior setae (c–d). Endite 2 with a relatively short anterior seta (Fig. 6C: 4) and four posterior setae (e–h). Endite 1 = gnathobase fully absent. Two ejector hooks of different length (Fig. 6C: ejh).

*Limb II* with exopodite as an elongated lobe (Fig. 6E: ext) bearing a soft distal seta, and a large, soft, lateral seta of same size with the former. Inner-distal lobe bearing five setae: four posterior setae (Fig. 6E: a–d) and a thin, stiff anterior seta (1) with length 3/4 of soft seta length, armed with minute setulae distally. Gnathobase (Fig. 1G) with two rows of setae: four anterior setae (Fig. 6F: 1'–4'), and numerous posterior setae of gnathobasic filter plate.



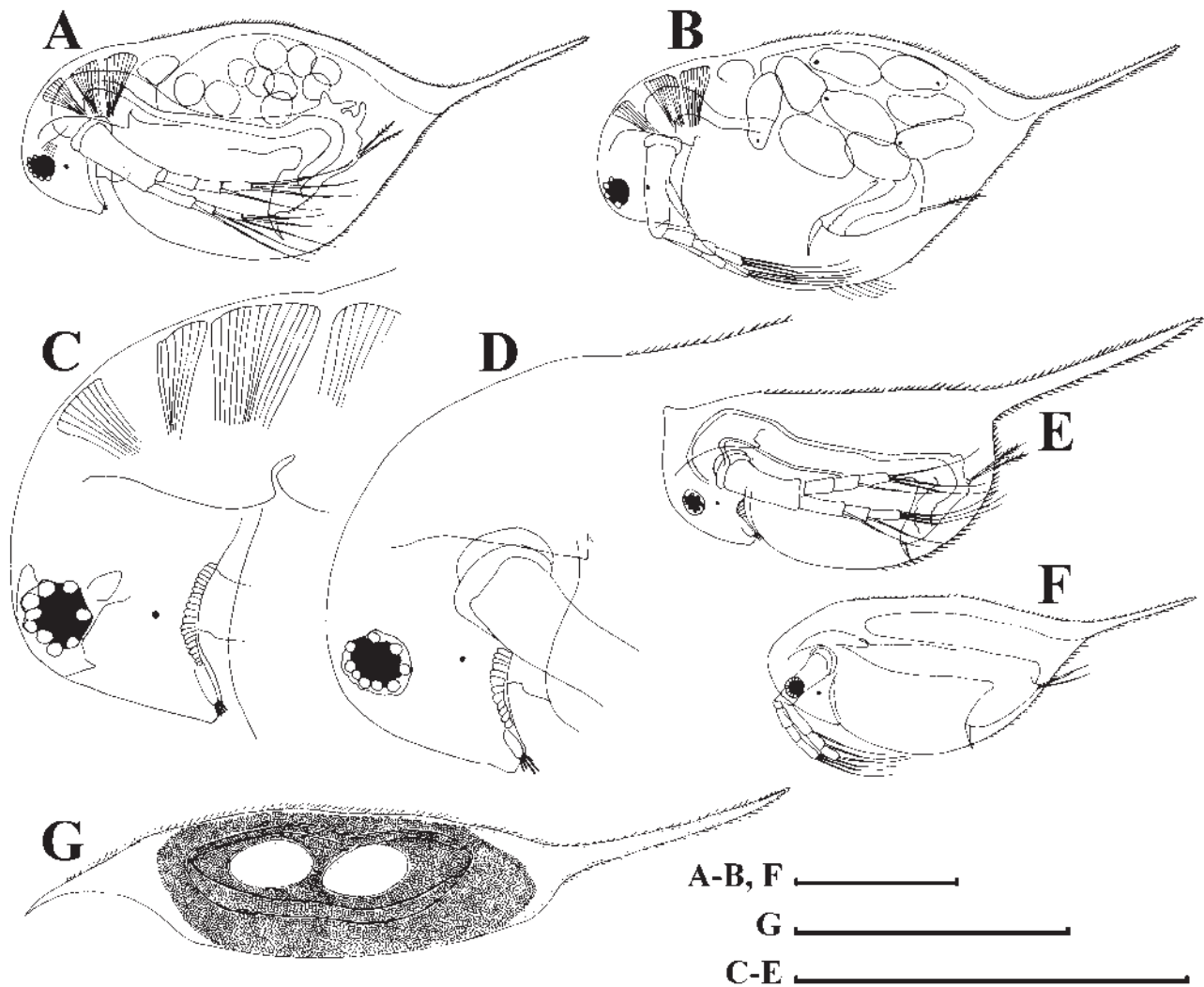
**FIGURE 1.** *Daphnia chevreuxi*, adult parthenogenetic female from “Environs de Bône”, Algeria, sample DGF 0783 (A–B) and Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy (C–G): A, large adult female. B, its head. C, postabdomen. D, abdomen. E, distal portion of postabdomen. F, limb I. G, gnathobase of limb II. H, limb V. Scale bars: 1 mm A; 0.1 mm for B–H.



**FIGURE 2.** *Daphnia chevreuxi*, female from Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy: A–B, adult parthenogenetic female. C, ephippial female. D, head, lateral view. E, head shield, dorsal view. F, fornix. G, juvenile female. Scale bars: 1 mm A–E; 0.1 mm for F–G.

*Limb III* with a large, setulated pre-epipodite (Fig. 7A: pep), ovoid epipodite (epp) and a flat exopodite (ext) bearing four distal setae (Fig. 7A: 1–4), among them seta 2 longest, distally with short setules (Fig. 7B), and two lateral setae (Fig. 7A: 5–6). Inner-distal portion of limb with endite 5 bearing a single, large anterior seta (Fig. 7C: 1), armed distally with short setulae and a large posterior seta, bearing long setulae (Fig. 7C: a); endite 4 with a single setulated anterior seta (2) and a single setulated posterior seta (b) somewhat shorter than anterior seta; endite 3 with a large anterior seta (Fig. 7C: 3) and two posterior setae; endite 2 with a large anterior seta (Fig. 7C: 4) and four posterior setae. The rest of the limb inner-distal portion as a singular large lobe, modified gnathobase, bearing numerous posterior soft setae, an anterior seta in its distal corner (Fig. 7C: 1') and two very short setae in middle of filter plate (Fig. 7D: 2' and 3').

*Limb IV* with a large, setulated pre-epipodite (pep), ovoid epipodite (epp) and a wide, flat exopodite (ext), bearing four distal (Fig. 7E: 1–4) and two lateral (Fig. 7E: 5–6) setae. Inner-distal portion of this limb with completely fused endites, distally with two setae of unclear homology (Fig. 7F: 1 and 2); the most part of limb inner margin is a gnathobase filter plate consisting of numerous posterior setae.



**FIGURE 3.** *Daphnia chevreuxi* from «Environs de Bône», sample DGF 0783 (A–F) and «Environs du Bône, Abreuvoir», sample DGF 0730 (D), Algeria: A–B, large adult parthenogenetic female. C–D, head, lateral view. E, juvenile female, instar I. F, juvenile female. G, ephippium. Scale bars 1 mm.

*Limb V* with a large, subovoid epipodite (epp), triangular exopodite (ext) supplied with two distal setae (Fig. 1H, 7G–H: 1–2), and a large, slightly curved lateral seta (3). Inner limb portion as an ovoid flat lobe, with setulated inner margin and a single, large seta.

**Juvenile female.** Body more elongated, dorsal margin almost straight, caudal spine longer (Figs 2G, 3E–F, 8A). Head relatively large as in adult female, with a rounded rostrum (Fig. 8B–C), sometimes with a small pointed helmet (Figs 2G, 3E). In instar I, head shield according to *Ctenodaphnia*-type: median anterior projection from carapace somewhat widened anteriorly, almost touching dorsal organ (Fig. 8D–E), instead of *Daphnia*-type with solely located dorsal organ and absent anterior projection of valves (Kotov & Boikova 2001).

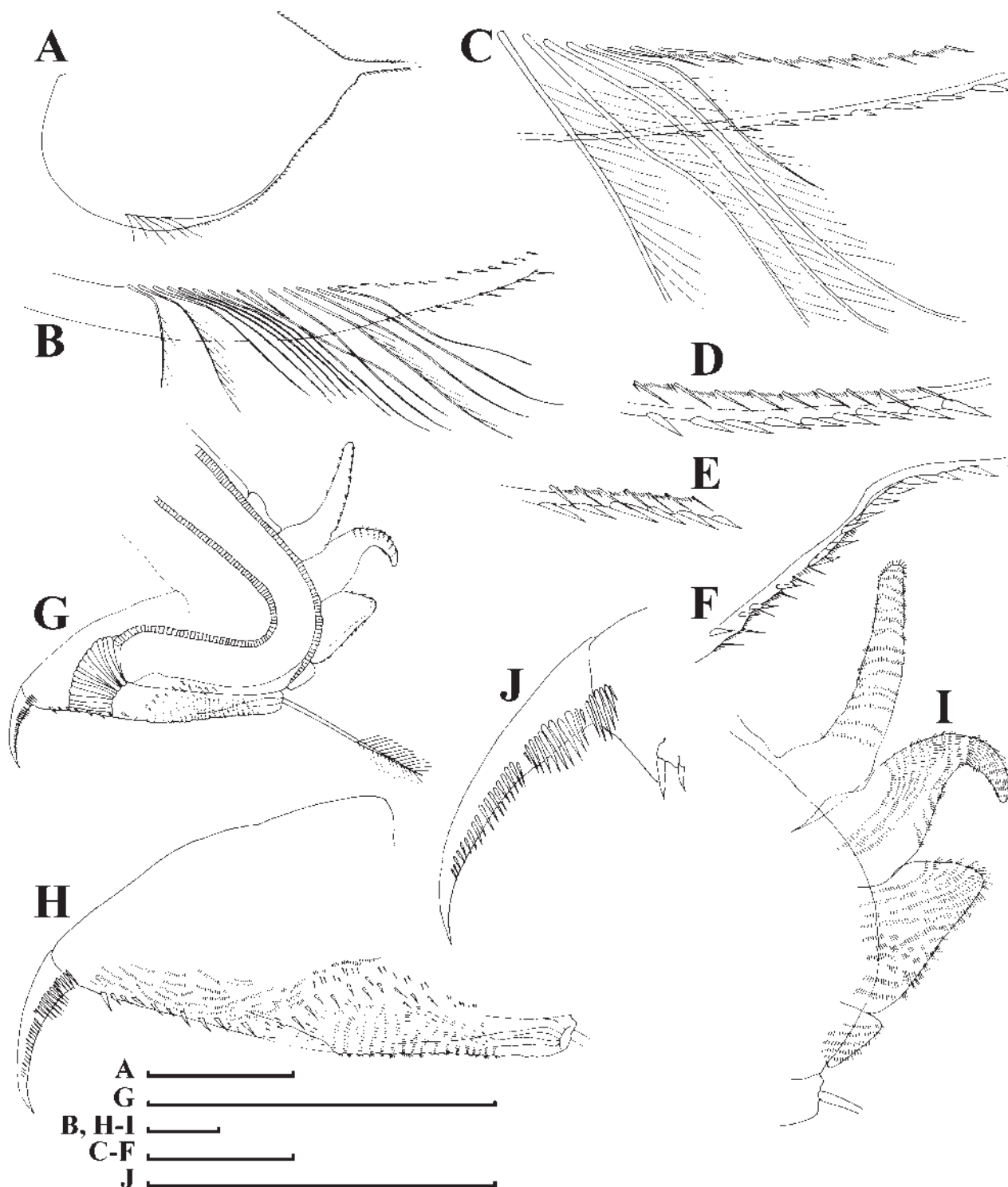
**Ephippial female.** Only dorsal carapace portion modified in ephippial female (Fig. 2C). Ephippium dark brown, elongated, bean-like (Figs 2C, 3G); two eggs with axes located at a very acute angle or almost parallel to dorsal margin; anterior processes present; postero-dorsal portion of valves (with shell spine) initially incorporated into ephippium. Dorsal margin of carapace specially re-enforced (Fig. 9A–C). Sculpture of ephippium as a network of small protuberances having smooth tips oriented somewhat posteriorly (Fig. 9B–D).

**Preephippial female.** A female transits from parthenogenesis to gamogenesis after two moults. Preephippial (after first such moult) female has an ephippium forming covers of its carapace. As a result, sculpture of future ephippium partly seen through its covers (Fig. 10A–F).

**Adult male. General.** Body elongated, body height/length (without shell spine) about 0.45–0.5; dorsal margin

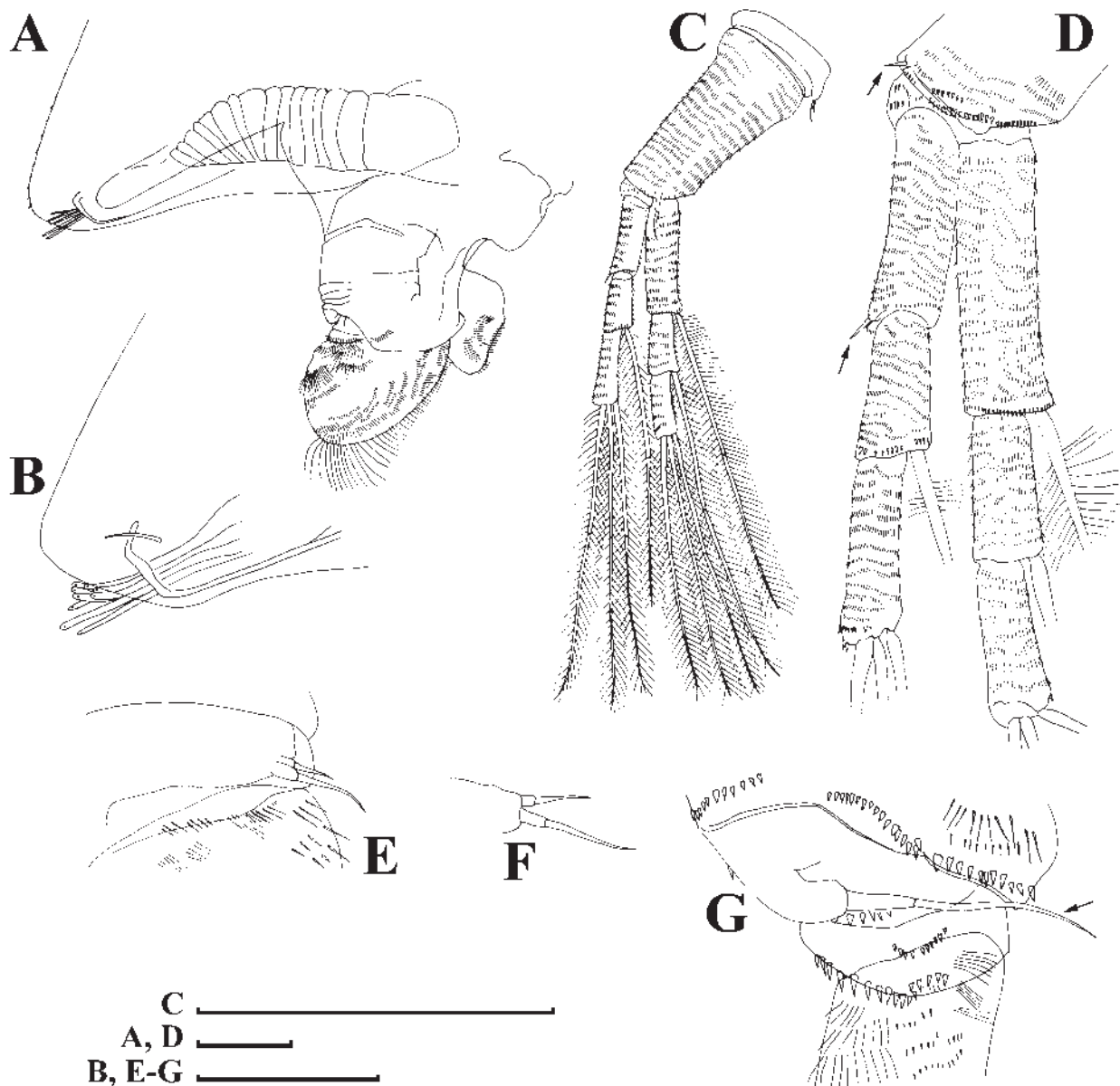
of valves almost straight, elevated above head; no distinct depression between head and valves; postero-dorsal angle distinct, with a relatively long caudal spine (Figs. 11A–C, 12A).

*Head* with a very short, rounded rostrum; its posterior margin straight; ventral margin straight or slightly concave, anteriormost extremity completely occupied with a very large optic vesicle (Figs 11D, 12B); a shallow post-ocular depression present. Compound eye especially large, ocellus minute. A short anterior projection from valves (Fig. 12D). Fornices well-developed, their tips smooth (Figs 11D, 12C–E).



**FIGURE 4.** *Daphnia chevreuxi*, adult parthenogenetic female from Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy; A, valve. B–C, ventral margin. D–F, postero-ventral margin. G, postabdomen. H, its distal portion. I, abdomen. J, postabdominal claw. Scale bars: 1 mm A, G; 0.1 mm for B–F, I–J.





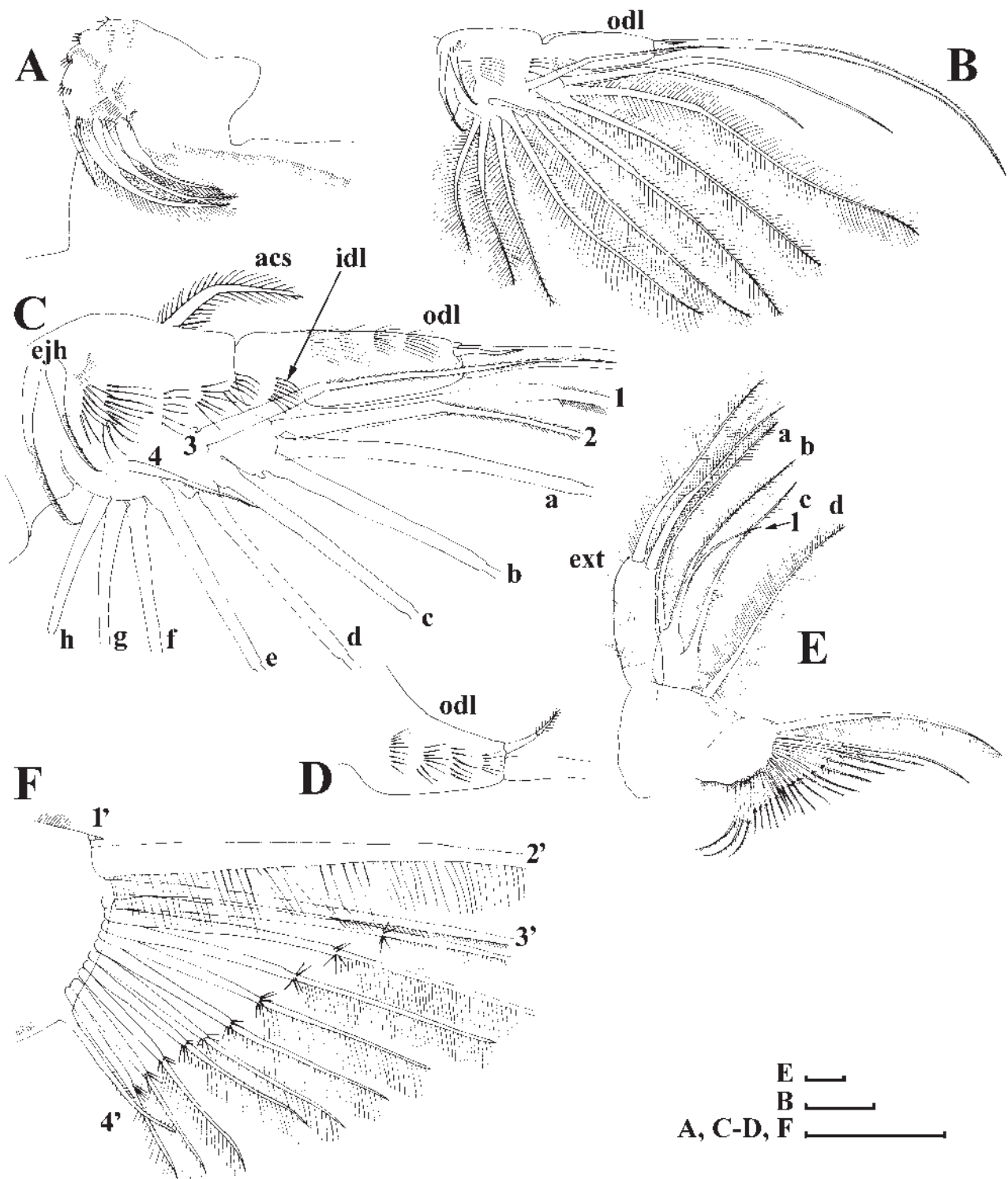
**FIGURE 5.** *Daphnia chevreuxi*, adult parthenogenetic female from Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy: A–B, rostrum and antenna I. C–D, antenna II. E–F, coxal portion. G, distal portion of basal segment. Scale bars: 1 mm C; 0.1 mm for A–B, D–G.

*Valve* with anterior margin slightly convex, supplied with exactly marginal, relatively short setae (Fig. 13A–B); antero-ventral angle prominent anteriorly, supplied with long setae; whole ventral margin with numerous setae located submarginally on inner face of valve. Postero-ventral portion of valve with marginal denticles and short setae located submarginally on inner face of valve; short setules between these setae (Fig. 13C–D).

*Abdomen* with a shallow mound on basal segment, other segments without projections (Figs 11E, 14A).

*Postabdomen* tapering distally, its distal portion bent (Figs 11E, 14A); ventral margin convex; preanal margin straight; anal margin depressed; gonopore opens subdistally (Fig. 11F, 14B); genital papilla absent. Few anal teeth present only in basal portion of anal margin (Figs. 11F, 14B). On outer side of postabdominal claw, the first and second (proximal) pectens consisting of relatively strong teeth; longest teeth shorter than the diameter of the claw base; third pecten consisting of numerous fine setulae not reaching the tip of claw.

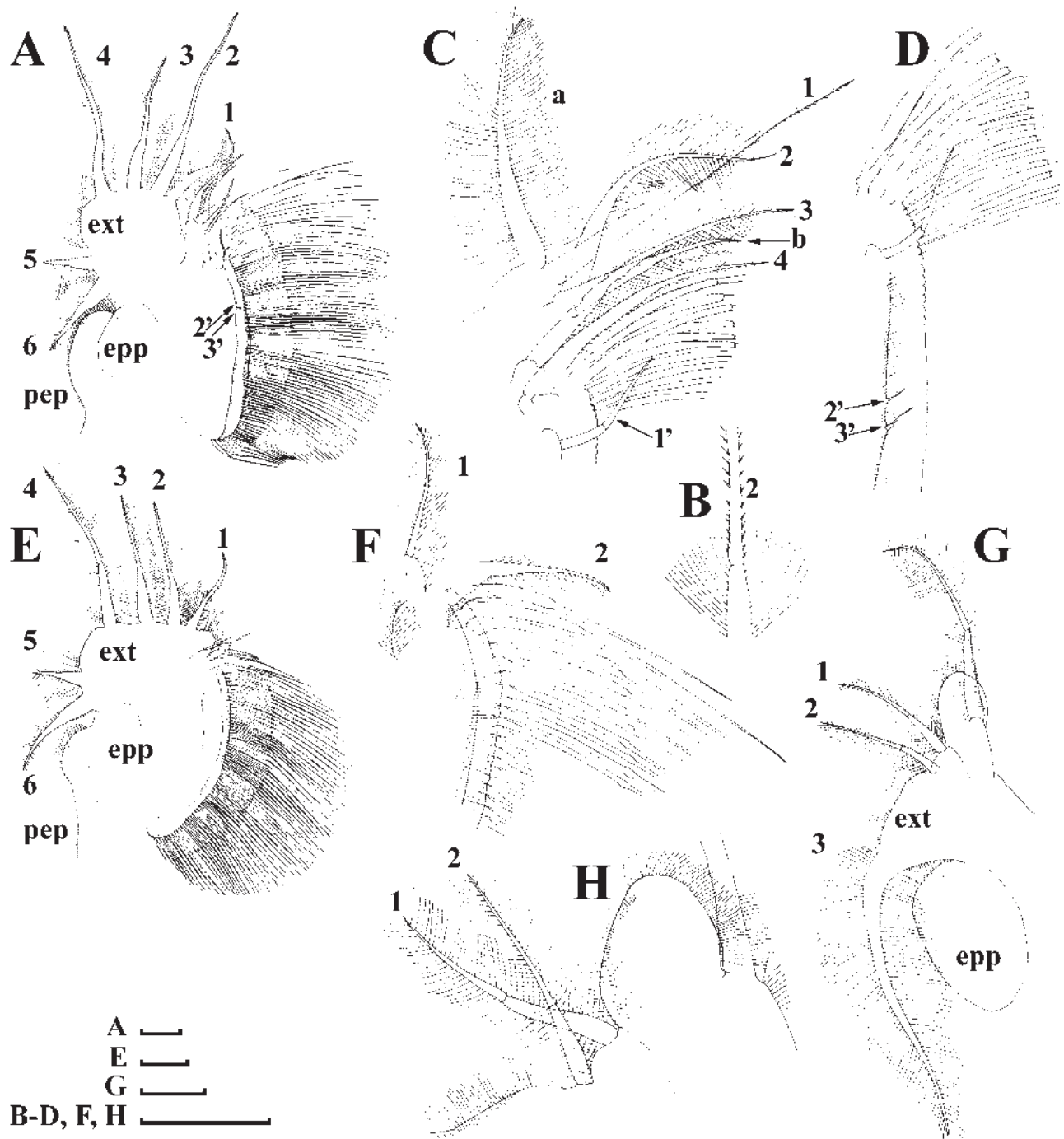
*Antenna I* long, regularly curved (Figs 11D, 12B, 13E). Nine short aesthetascs; antennular sensory seta very short, located distally. Length of flagellum less than half body length of the antenna I. The distal segment of flagellum covered with short setules (Figs. 11G, 13E).



**FIGURE 6.** *Daphnia chevreuxi*, head and thoracic limbs of adult parthenogenetic female from Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy: A, maxilla I. B–C, limb I. D, its outer distal lobe. E, limb II. F, its gnathobase. Scale bars 0.1 mm.

*Antenna II* (Figs 12A, 13F) relatively larger as compared to female.

*Limb I.* Outer distal lobe (Fig. 14C–E: odl) large, bearing a rudimentary seta, a small hillock and a very large seta. Inner distal lobe (idl) with a bent copulatory hook, a single long seta (Fig. 14 E: 1) and a rudimentary seta (Fig. 14 E: 1'). Additional seta on endite 4 (Fig. 14 D: 2') anterior seta shorter than in female and supplied with longer setules (Fig. 14 D: 3) while anterior seta longer than in female (Fig. 14 D: 4).

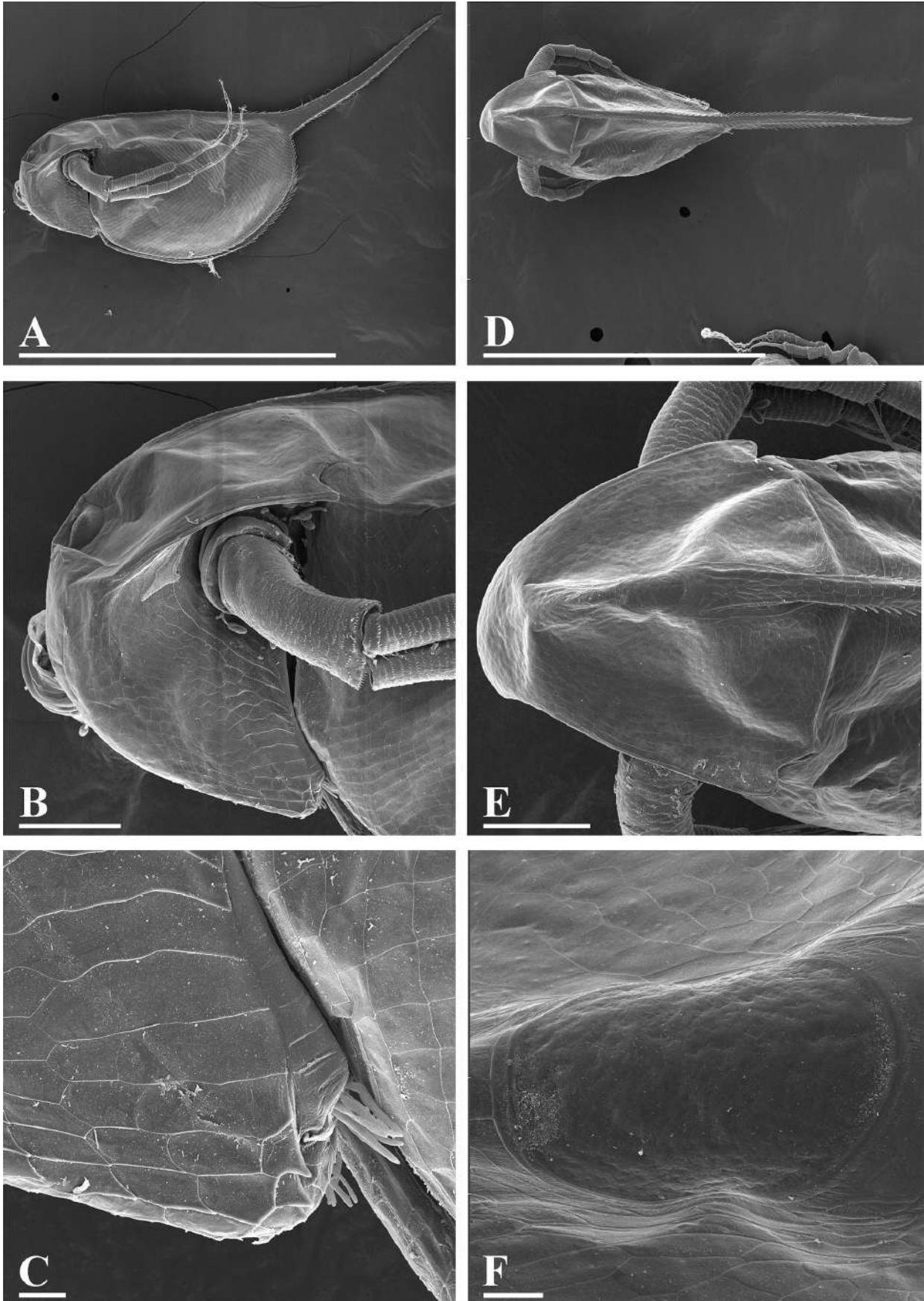


**FIGURE 7.** *Daphnia chevreuxi*, thoracic limbs of adult parthenogenetic female from Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy: A, limb III. B, seta 2 of its exopodite. C–D, its inner-distal portion. E, limb IV. F, its inner-distal portion. G, limb V. H, its distal portion. Scale bars 0.1 mm.

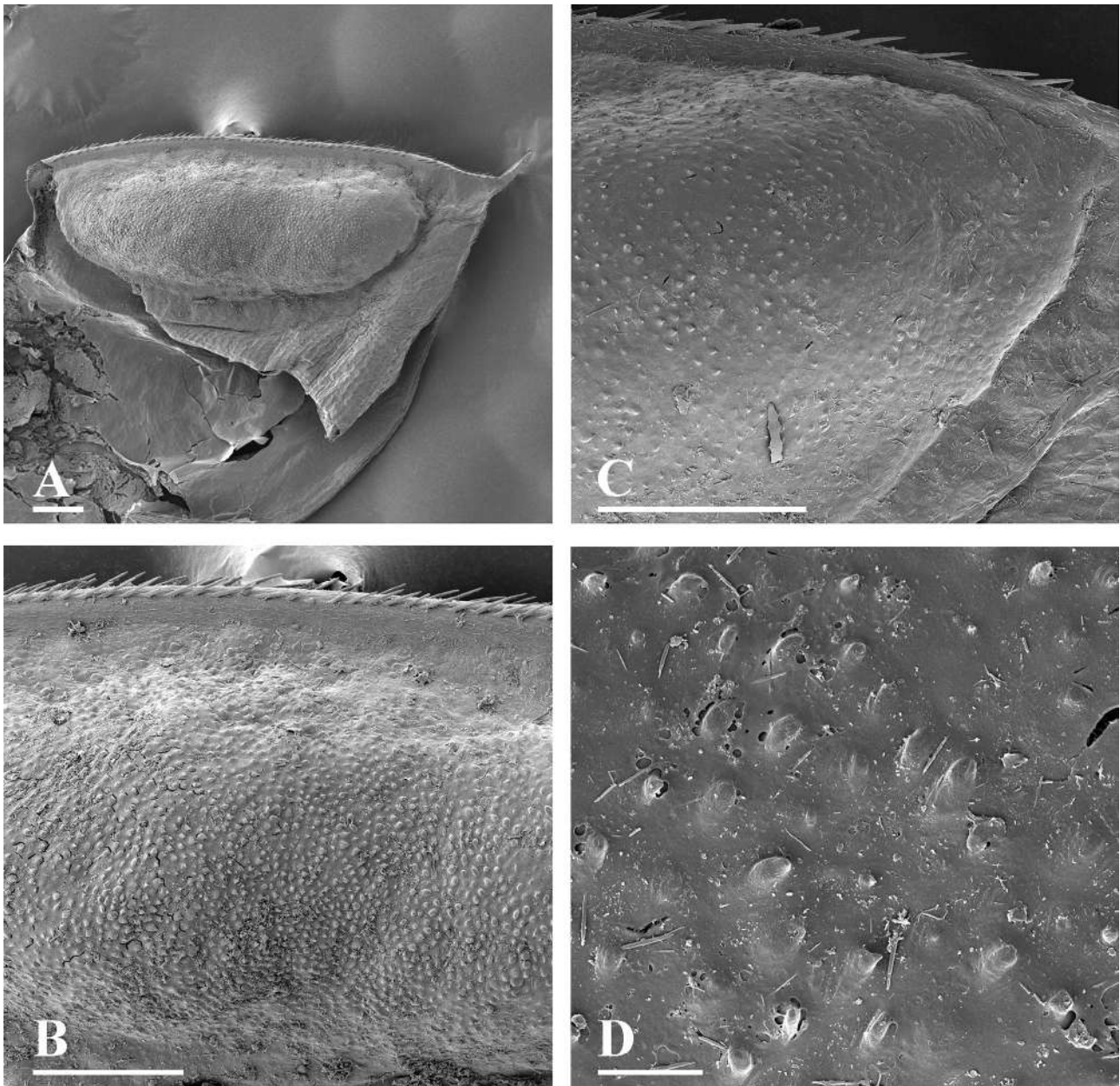
*Limb II.* Distalmost endite with seta 1 remarkably stronger than in female, slightly bent and asymmetrically setulated, with a blunt tip (Figs 11H: arrows, 14F: 1). Limb V as in female (Fig. 14G).

**Juvenile male II.** Body elongated, eye capsule less developed as compared to adult male, posterior incision more protected anteriorly, fornices small; no setules at antero-ventral valve portion, antenna I shorter than in adult male, with shorter flagellum (Fig. 12F–G).

**Size.** Parthenogenetic females 0.9–3.8 mm in our material (2.4–3.8 mm according to Benzie, 2005); adult males 1.1–2.4 mm in our material (1.3–1.6 mm according to Benzie, 2005).



**FIGURE 8.** *Daphnia chevreuxi*, juvenile female from Gorgo di Rebuttone, Sicily, Italy: A, lateral view. B, head. C, rostrum. D, dorsal view. E, head, dorsal view. F, dorsal organ. Scale bars: 1 mm for A, B; 0.1 mm for B, E; 0.01 mm for C, F.



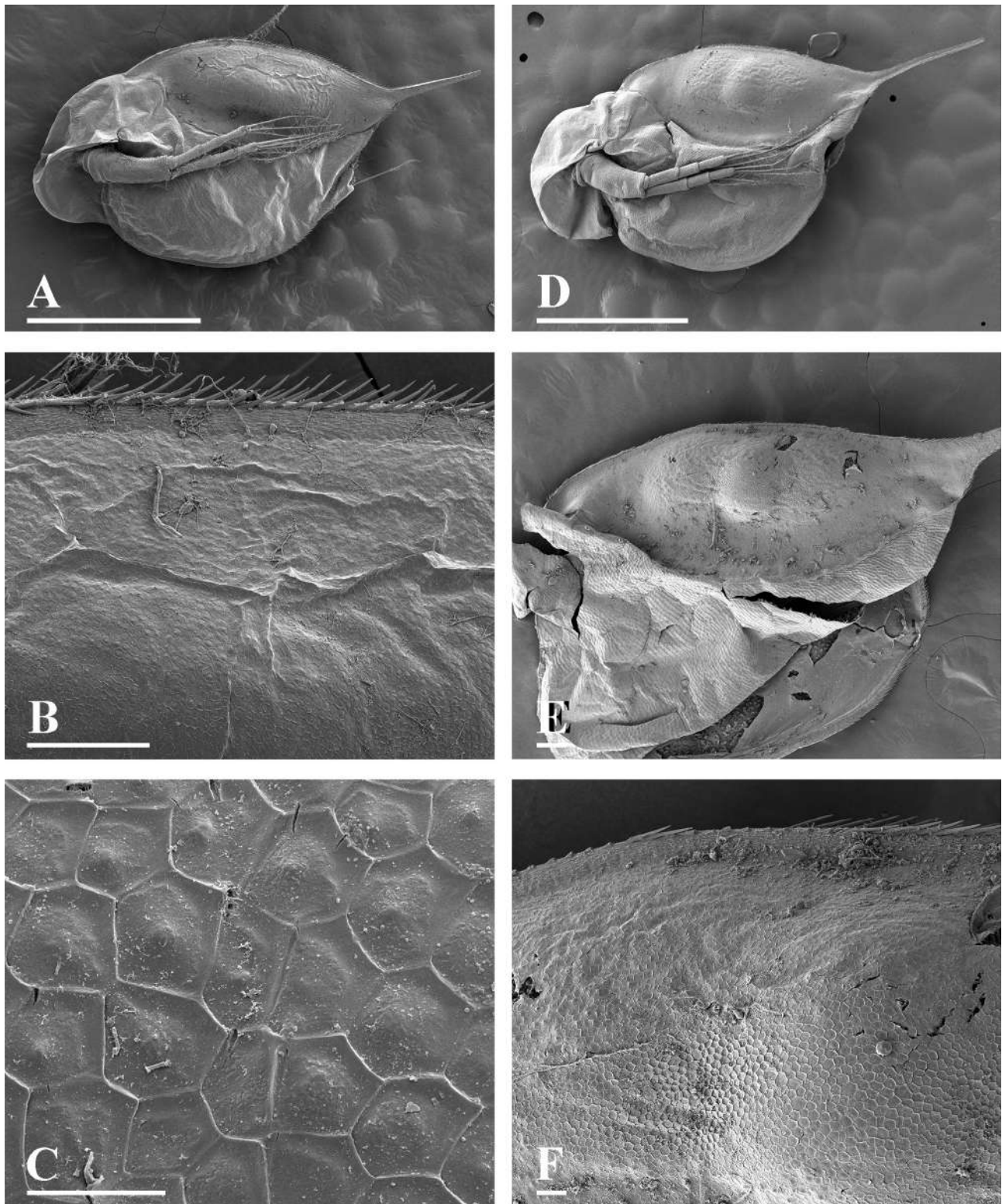
**FIGURE 9.** *Daphnia chevreuxi*, ephippial female from «Environs de Bône. Abreuvoir», sample DGF 0767, Algeria: A, carapace with ephippium. B–D, ephippium sculpture. Scale bars: 0.1 mm for A–C; 0.01 mm for D.

**Differential diagnosis.** Main characteristic traits of this taxon are (1) strongly reduced (almost undiscernible) body of antenna I and (2) a very short median anterior projection of the carapace (its length less than 0.2 of head shield length) (Glagolev & Alonso 1990; Glagolev 1995; Benzie 2005; Korovchinsky *et al.* 2021). Among Eurasian species of the subgenus, only *D. (C.) chevreuxi* and *D. (C.) hispanica* have an accessory seta on limb I (Glagolev & Alonso 1990). Moreover juveniles of *D. (C.) chevreuxi* sometimes bear a small triangular helmet not characteristic of other Eurasian ctenodaphniids.

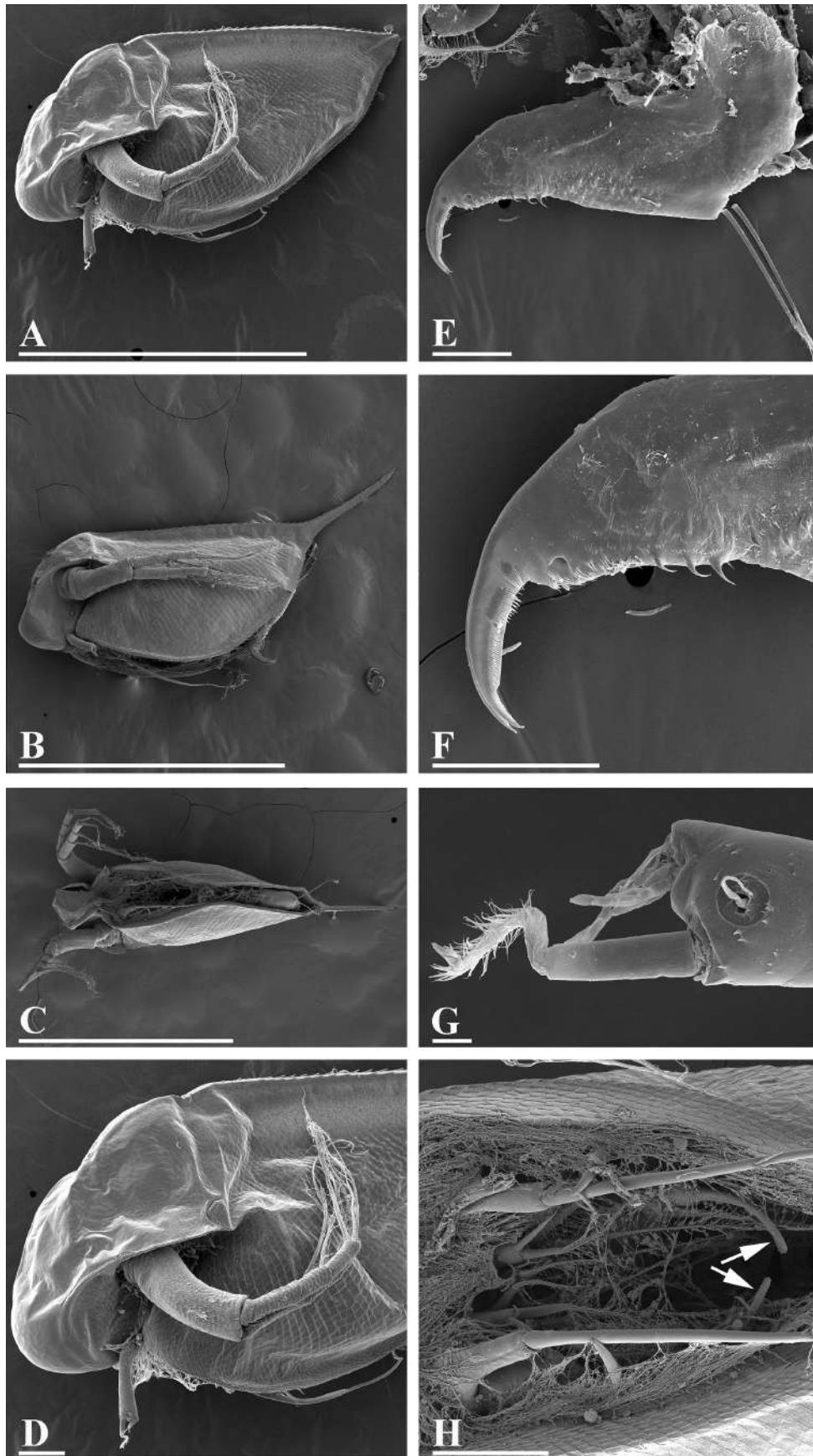
*D. chevreuxi* has no: lateral keels on the head shield as *D. (C.) magna*; sharp elongated fornices as *D. (C.) lumholtzi*; dorsal head plate as *D. (C.) atkinsoni*-group; sharp dorsal keel as *D. (C.) hispanica*. The most problematic is differentiation between *D. (C.) chevreuxi* and *D. (C.) similis*-group, first of all, the former and *D. (C.) similis* s.str. which is common in southern half of Europe. Female of *D. (C.) chevreuxi* has: (1) more curved dorsal margin; (2) more reduced (almost undiscernible) body of antenna I; (3) longer seta 1' on gnathobase III (length c.a. 0.5 length of seta 4); (4) two well-developed distal setae on exopodite V (while one of them reduced in size or completely absent

in in *D. similis* group). Male has: (1) strongly reduced postanal teeth, (2) regularly curved antenna I; (3) two well-developed distal setae on exopodite V.

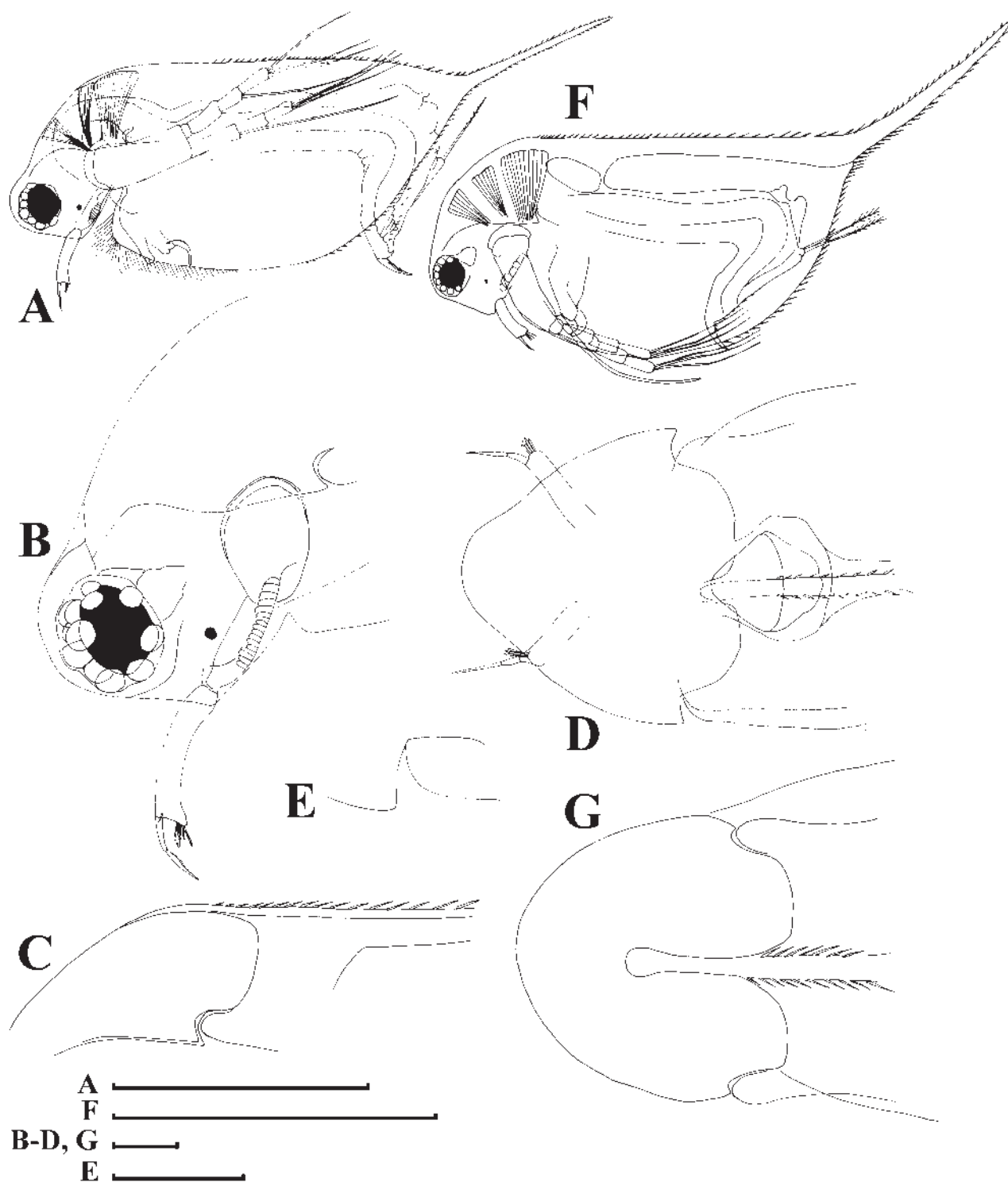
**Distribution and ecology.** We have analysed *D. (C.) chevreuxi* specimens from Algeria, Sicily and North Macedonia. In North Macedonia, the species was found in former clay pits (Petkovski 1970), but such populations could have been introduced due to human activities, and their native status is questionable.



**FIGURE 10.** *Daphnia chevreuxi*, pre-ephippial female from Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy (A–C) and Slavej, North Macedonia (D–F): A, lateral view. B–C, sculpture of carapace. D, lateral view. E, carapace. F, its sculpture. Scale bars: 1 mm for A, D; 0.1 mm for B, E; 0.01 mm for C, F.



**FIGURE 11.** *Daphnia chevreuxi*, adult male from Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy: A–B, lateral and latero-ventral view. C. ventral view. D. head. E, postabdomen. F, its distal portion. G, distal portion of antenna I. H, central portion of body, ventral view. Scale bars: 1 mm for A–C; 0.1 mm for D–F, H; 0.01 mm for G.

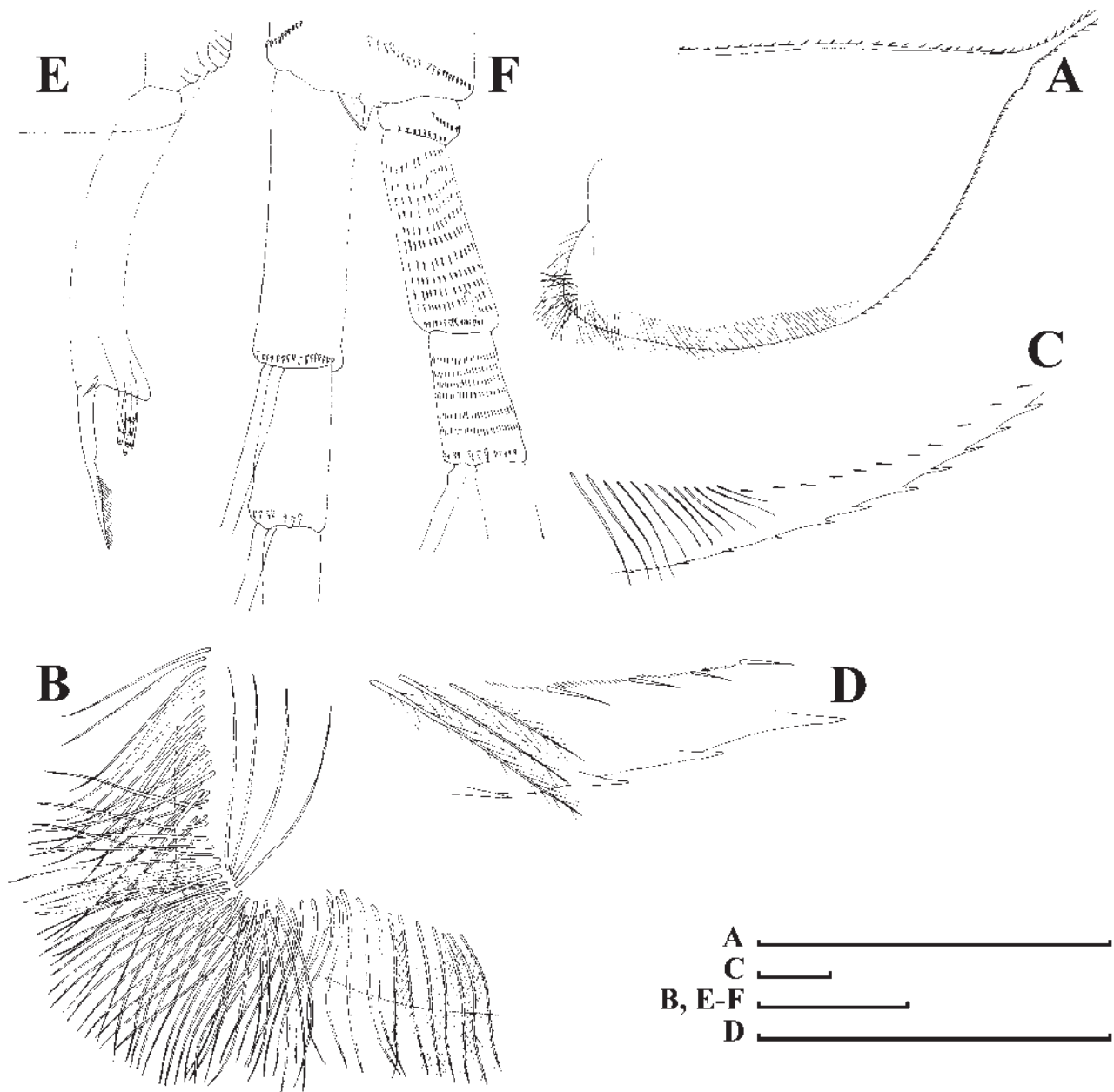


**FIGURE 12.** *Daphnia chevreuxi*, male from Stagno di C. da Buffali (Nebrodi, Cesarò), Sicily, Italy (A–B, D–E) and “Environs de Bône”, Algeria, sample DGF 0783 (C, F–G): A, adult male, lateral view. B, head. C, dorsal portion of head, lateral view. D, head, dorsal view. E, fornix. F, juvenile male of instar II. G, head, dorsal view. Scale bars: 1 mm for A, F; 0.1 mm for B–E, G.

The taxon was also recorded from mainland Italy (Margaritora 1983; Margaritora 1985), mainland Greece (Stephanides 1948; Marrone *et al.* 2019b), Romania (Negrea 1983), Bulgaria (Flössner 1980; Naidenow 1994), Corfu (Stephanides 1948), Corsica and Sardinia (Margaritora 1985; Margaritora *et al.* 1975), Crete (Marrone *et al.* 2019b), and the Maghreb (Dumont 1979; Mouelhi *et al.* 2000), but no samples from these regions have been analysed in the frame of present work. This species is also reported from Israel, although the actual conspecificity of



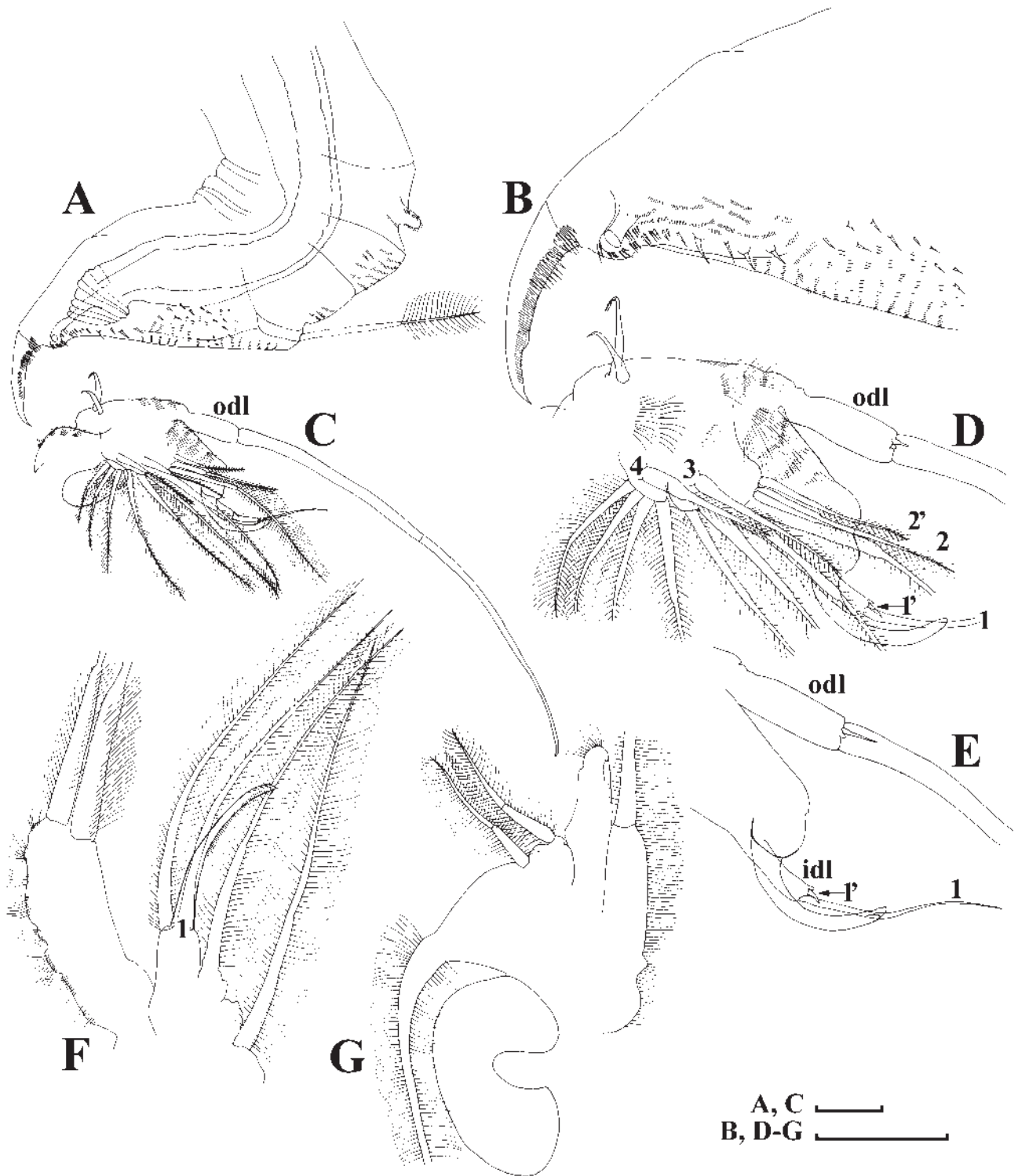
Israeli populations with *D. chevreuxi* s.str. should be investigated with molecular tools (Adamowicz *et al.* 2009). In reality, the taxon could be represented by a series of close species as it is demonstrated for some other Mediterranean endemics (e.g. Marrone *et al.* 2010).



**FIGURE 13.** *Daphnia chevreuxi*, adult male from Gorgo di Rebuttone (Altofonte), Sicily, Italy: A, valve. B, its anterior portion. C–D, its postero-ventral portion. E, antenna I. F, antenna II. Scale bars: 1 mm for A; 0.1 mm for B–F.

Actual occurrence of the species in Morocco (Mouelhi *et al.* 2000; Ramdani 1988) and Romania (Negrea 1983) needs to be verified. *D. chevreuxi* is considered a “circum-Mediterranean” taxon (Benzie 2005), but its verified distribution ranges from the Maghreb through Italian Peninsula and Tyrrhenian islands to the Balkans, whereas it is absent from Iberian Peninsula (Alonso 1996), France (Amoros 1984), Libya, Egypt (Dumont 1979) and Turkey (Güher 2014). Moreover *D. byzantina* Mucke, 1951 described from Turkey (Mucke 1951) is most probably a junior synonym of *D. chevreuxi*, or a member of the *chevreuxi* group possibly conspecific with the aforementioned taxon from Israel.

The species is linked with long-lasting and poorly mineralized temporary ponds, located from the sea level up to 1500 m.a.s.l. (Gauthier 1928; Stephanides 1948; Margaritora 1985; Ghaouaci *et al.* 2018; Marrone & Vecchioni 2021), in areas characterized by a typical Mediterranean climate (Peel *et al.* 2007).



**FIGURE 14.** *Daphnia chevreuxi*, adult male from a pond, Gorgo di Rebuttone (Altofonte), Sicily, Italy: A, postabdomen. B, its distal portion. C–D, limb I. E, its inner distal lobe. F, distal portion of limb II. G, limb V. Scale bars 0.1 mm.

## Discussion

The genus *Daphnia* is relatively well characterized in the Mediterranean area: *D. (Ctenodaphnia) chevreuxi* was the last endemic species of the subgenus *D. (Ctenodaphnia)* which was inadequately described to date. In addition to *D. (C.) chevreuxi*, two other Mediterranean endemic species of this subgenus are well-described, namely *D. (C.)*

*mediterranea* Alonso, 1985 and *D. (C.) hispanica* Glagolev & Alonso, 1990; the latter is the sister species of *D. chevreuxi* (Adamowicz *et al.* 2009). Moreover, several cryptic lineages with presumably endemic Mediterranean distribution are known within *D. atkinsoni* group (Petrušek *et al.* 2009).

*Daphnia chevreuxi* is currently associated with areas which were only marginally influenced by the drastic climate changes, including strong aridification and cooling phases, which took place during coldest Pleistocene phases (Hewitt 2001; Stewart & Lister 2001). According to Korovchinsky (2006), the subtropics and adjacent regions with warm climate represent zones currently inhabited by pre-Quaternary relicts which could survive in situ during Plio-Pleistocene climatic fluctuations (Marrone *et al.* 2010; Reniers *et al.* 2013; Marrone *et al.* 2017; Kappas *et al.* 2017). In this frame, *Daphnia chevreuxi* represents both a biogeographical relictual taxon and a strongly diverging species within the subgenus *D. (Ctenodaphnia)* as it is distributed in regions known as important Pleistocene refugia, and it belongs to an earlier derived branch of this subgenus (Adamowicz *et al.* 2009).

To date, no Mediterranean endemic species are known for the subgenus *Daphnia (Daphnia)*. Such a pattern is possibly to be ascribed to the occurrence of a latitudinal diversity pattern differently affecting the two *Daphnia* subgenera in Eurasia (Benzie 2005). Within the genus *Daphnia*, the highest diversity of *Daphnia (Daphnia)* is located at temperate and higher latitudes of the Palaearctic region, and that of *D. (Ctenodaphnia)* is located at lower latitudes and in warmer climates. In this way, *D. (Ctenodaphnia)* could be considered the most typical *Daphnia* subgenus of the Mediterranean area, and its pattern of endemism seems consistent with it.

Moreover, representatives of the subgenus *D. (Ctenodaphnia)* are, with few exceptions, typical inhabitants of temporary and ephemeral waters, which are often regarded as “ecological refugia” (Davis *et al.* 2013), hosting an ancient and very peculiar crustacean biota (Sahuquillo & Miracle 2013); conversely, the vast majority of the species belonging to *Daphnia (Daphnia)* prefers larger and permanent water bodies, although they can be present in temporary waters as well (Benzie 2005). The paradox of temporary pools being “probably the most permanent of all freshwater habitats” (Fryer 1985) is well-known, and temporary pools are also inhabited by other “living fossils”, including large branchiopods (Dumont & Negrea 2002). Stability of such communities during millions of years are confirmed by fossil records (Gueriau *et al.* 2016; Zharov *et al.* 2020); unfortunately, temporary water bodies and their biota are nowadays threatened by human activities and human-induced climate change, which is predicted to have a significant impact on the inland waters of the Mediterranean area (Markovic *et al.* 2017). In this framework, the realization of a complete and precise inventory of the biological diversity of the inland waters of the Mediterranean area is urgent and necessary to understand, and possibly face, the risks it currently faces.

Temporary waters of Eurasia are also inhabited by the representatives of *Daphnia (Daphnia) pulex* and *D. (D.) obtusa* species complexes (Benzie 2005), which coexist with the ctenodaphniids. The former are awaiting detailed revisions and there are only preliminary ideas about their distribution in Eurasia (Crease *et al.* 2012). However there is a well-studied group of *D. (Daphnia)* inhabiting temporary waters with a contrast pattern to *D. (Ctenodaphnia)*, namely the *D. (D.) curvirostris* complex. In Southern Europe it is represented by a single taxon, *D. curvirostris* Eylmann, 1887, and only in the Tatras there are few relict populations of the closely related *D. hrbaceki* Juračka, Kořínek & Petrušek, 2010. In contrast, along the Pacific coast of Eurasia temporary water bodies are populated by many endemic taxa of the *D. (D.) curvirostris* species complex, which differentiated in early-middle Caenozoic (Kotov *et al.* 2021), whereas temporary water bodies are inhabited only by a single ctenodaphniid species, *D. (C.) sinensis* Gu, Xu, Li, Dumont & Han, 2013 (Xiang *et al.* 2015; Garibian *et al.* 2020). Therefore, we observe contrasting patterns at the two ends of Eurasia: several species of *D. (Ctenodaphnia)* and only one species of the *D. (D.) curvirostris* complex in the West, and a single *D. (Ctenodaphnia)* with several *curvirostris*-like species in the East.

Subtropics and closest territories of the Pacific coast of Eurasia represent another relict endemism zone predicted by Korovchinsky (2006). Many endemic species and locally distributed clades have been found there recently based on morphology, molecular methods and their combination (Kotov *et al.* 2021; Korovchinsky 2013; Maruoka *et al.* 2018; Neretina *et al.* 2021) similarly to the Mediterranean region. To date, the two ends of Eurasia are well-studied, while the subtropics and adjacent territories in the central portion of continent are significantly less studied, and studies of the Pontocaspian region and Middle Asia are missing.

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## References

- Adamowicz, S.J., Petrusek, A., Colbourne, J.K., Hebert, P.D.N. & Witt, J.D.S. (2009) The scale of divergence: a phylogenetic appraisal of intercontinental allopatric speciation in a passively dispersed freshwater zooplankton genus. *Molecular phylogenetics and evolution*, 50, 423–436.  
<https://doi.org/10.1016/j.ympev.2008.11.026>
- Alonso, M. (1985) *Daphnia (Ctenodaphnia) mediterranea*: A new species of hyperhaline waters, long confused with *D. (C.) dolichocephala* Sars, 1895. *Hydrobiologia*, 128, 217–228.  
<https://doi.org/10.1007/BF00006817>
- Alonso, M. (1990) Anostraca, Cladocera and Copepoda of Spanish saline lakes. In: Comín, F.A. & Northcote, T.G. (Eds.), *Saline Lakes*. Springer Netherlands, Dordrecht, pp. 221–231.
- Alonso, M. (1991) Review of Iberian Cladocera with remarks on ecology and biogeography. *Hydrobiologia*, 225, 37–43.  
<https://doi.org/10.1007/BF00028383>
- Alonso, M. (1996) Crustacea Branchiopoda. In: Ramos, M. (Ed.), *Fauna Ibérica. Vol. 7*. Museo Nacional de Ciencias Naturales. Consejo Superior de Investigaciones Científicas, Madrid, pp. 1–486.
- Alonso, M. (1998) Las lagunas de la España peninsular. *Limnetica*, 15, 1–176.
- Alonso, M., Neretina, A.N. & Ventura, M. (2021) *Ceriodaphnia smirnovi* (Crustacea: Cladocera), a new species from the Mediterranean Region, and a phylogenetic analysis of the commonest species. *Zootaxa*, 4974 (1), 146.  
<https://doi.org/10.11646/zootaxa.4974.1.1>
- Alonso, M. & Pretus, J.L. (1989) *Alona iberica*, new species: first evidence of noncosmopolitanism within the *A. karua* complex (Cladocera, Chydoridae). *Journal of Crustacean Biology*, 9, 459–476.  
<https://doi.org/10.1163/193724089X00449>
- Amoros, C. (1984) Crustacés cladocères. *Bulletin mensuel de la Société Linnéenne de Lyon*, 53, 1–63.
- Bănărescu, P. (1990) s.n. In: *Zoogeography of fresh waters. Vol. 2. distribution and dispersal of freshwater animals in North America and Eurasia*. Aula-Verlag, Wiesbaden, pp. 519–1091.
- Benzie, J.A.H. (2005) *The genus Daphnia (including Daphniopsis) (Anomopoda. Daphniidae)*. Kenobi Productions, Ghent, vii + 376 pp.
- Błędzki, L.A. & Rybak, J. (2016) *Freshwater crustacean zooplankton of Europe. Cladocera & Copepoda (Calanoida, Cyclopoida) key to species identification, with notes on ecology, distribution, methods and introduction to data analysis*. Springer, Cham, XV + 918 pp.
- Blondel, J., Aronson, J., Bodiou, J. & Boeuf, G. (2010) *The Mediterranean region. Biological diversity in space and time*. Oxford University Press, Oxford, xv + 376 pp.
- Bozkurt, A. & Aktas, M. (2016) Distribution of Cladocera species in different waters of Turkey. *Journal of Limnology and Freshwater Fisheries Research*, 137, 137–143.  
<https://doi.org/10.17216/limnofish.279722>
- Brancelj, A. (1990) *Alona hercegovinae* n. sp. (Cladocera: Chydoridae), a blind cave-inhabiting Cladoceran from Hercegovina (Yugoslavia). *Hydrobiologia*, 199, 7–16.  
<https://doi.org/10.1007/BF00007828>
- Bromley, H.J. (1993) A checklist of the Cladocera of Israel and Eastern Sinai. *Hydrobiologia*, 257, 21–28.  
<https://doi.org/10.1007/BF00013993>
- Cheikh Albassatneh, M., Escudero, M., Monnet, A.-C., Arroyo, J., Bacchetta, G., Bagnoli, F., Dimopoulos, P., Hampe, A., Leriche, A., Médail, F., Nikolic, T., Ponger, L., Vendramin, G.G. & Fady, B. (2021) Spatial patterns of genus-level phylogenetic endemism in the tree flora of Mediterranean Europe. *Diversity and Distributions*, 27, 913–928.  
<https://doi.org/10.1111/ddi.13241>
- Crease, T.J., Omilian, A.R., Costanzo, K.S. & Taylor, D.J. (2012) Transcontinental phylogeography of the *Daphnia pulex* species complex. *PloS One*, 7, e46620.  
<https://doi.org/10.1371/journal.pone.0046620>
- Cuttelod, A., García, N., Malak, D.A., Temple, H.J. & Katariya, V. (2009) The Mediterranean: a biodiversity hotspot under threat. *Wildlife in a Changing World—an analysis of the 2008 IUCN Red List of Threatened Species*, 89, 9.
- Davis, J., Pavlova, A., Thompson, R. & Sunnucks, P. (2013) Evolutionary refugia and ecological refuges: key concepts for conserving Australian arid zone freshwater biodiversity under climate change. *Global change biology*, 19, 1970–1984.  
<https://doi.org/10.1111/gcb.12203>

- De Gelas, K. & Meester, L. de (2005) Phylogeography of *Daphnia magna* in Europe. *Molecular ecology*, 14, 753–764. <https://doi.org/10.1111/j.1365-294X.2004.02434.x>.
- Dumont, H.J. (1979) *Limnologie van Sahara en Sahel*. Ph.D. Thesis. University of Ghent, Ghent, 557 pp.
- Dumont, H.J., Laureys, P. & Pensaert, J. (1979) Anostraca, Conchostraca, Cladocera and Copepoda from Tunisia. *Hydrobiologia*, 66, 259–274. <https://doi.org/10.1007/BF00020908>.
- Dumont, H.J. & Negrea, Ș. (2002) *Branchiopoda*. Backhuys, Leiden, 398 pp.
- Flössner, D. (1980) Über zwei bemerkenswerte Branchiopoda aus Bulgarien – *vornatscheri bulgaricus* subsp. n. (Crustacea, Anostraca) und *Daphnia chevreuxi* Richard (Crustacea, Cladocera). *Acta Zoologica Bulgarica*, 16, 63–68.
- Fryer, G. (1985) Structure and habits of living branchiopod crustaceans and their bearing on the interpretation of fossil forms. *Earth and Environmental Science Transactions of the Royal Society of Edinburgh*, 76, 103–113. <https://doi.org/10.1017/S0263593300010373>
- Garibian, P.G., Neretina, A.N., Korovchinsky, N.M., Sinev, A.Y., Tchabovsky, A.V., Kotov, A.A. & Smirnov, N.N. (2020) The Southern part of Russian Far East and Korean Peninsula as a transition zone between the boreal and tropical faunas of the waterfleas (Cladocera, Crustacea). *Zoologicheskii Zhurnal*, 99, 1094–1109. <https://doi.org/10.1134/S1062359021070128>
- Gauthier, H. (1928) *Recherches sur la faune des eaux continentales de l'Algérie et de la Tunisie*. Minerva, Alger, 419 pp.
- Ghaouaci, S., Amarouyache, M., Sinev, A.Y., Korovchinsky, N.M. & Kotov, A.A. (2018) An annotated checklist of the Algerian Cladocera (Crustacea: Branchiopoda). *Zootaxa*, 4377, 412–430. <https://doi.org/10.11646/zootaxa.4377.3.5>
- Glagolev, S.M. (1995) Genus *Daphnia*. In: Alekseev, V.R. (Ed.). *A key-book of the freshwater invertebrates of Russia and its vicinity*. Vol. 2. Crustacea. Zoological Institute of Russian Academy of Sciences, St. Petersburg, pp. 48–58.
- Glagolev, S.M. & Alonso, M. (1990) *Daphnia (Ctenodaphnia) hispanica* sp.nov., a new daphnid (Cladocera) from Spain. *Hydrobiologia*, 194, 149–162.
- Gueriau, P., Rabet, N., Clément, G., Lagebro, L., Vannier, J., Briggs, D.E.G., Charbonnier, S., Olive, S. & Béthoux, O. (2016) A 365-Million-Year-Old Freshwater Community Reveals Morphological and Ecological Stasis in Branchiopod Crustaceans. *Current biology: CB*, 26, 383–390. <https://doi.org/10.1016/j.cub.2015.12.039>
- Guerne, J. & Richard, J. (1986) Première list des Copépodes et Cladocères d'eau douce du Portugal. *Bulletin de la Société Zoologique de France*, 21, 157–159.
- Güher, H. (2014) A checklist for zooplankton (Rotifera, Copepoda, Cladocera) of European Turkey inland waters. *Ege Journal of Fisheries and Aquatic Sciences*, 31, 221–225. <https://doi.org/10.12714/egejfas.2014.31.4.08>
- Gurney, R. (1909) X. On the Fresh-water Crustacea of Algeria and Tunisia. *Journal of the Royal Microscopical Society*, 29, 273–305. <https://doi.org/10.1111/j.1365-2818.1909.tb01699.x>
- Hermoso, V. & Clavero, M. (2011) Threatening processes and conservation management of endemic freshwater fish in the Mediterranean basin: a review. *Marine and Freshwater Research*, 62, 244. <https://doi.org/10.1071/MF09300>
- Hewitt, G.M. (2001) Speciation, hybrid zones and phylogeography – or seeing genes in space and time. *Molecular ecology*, 10, 537–549. <https://doi.org/10.1046/j.1365-294x.2001.01202.x>
- Kappas, I., Mura, G., Synefiaridou, D., Marrone, F., Alfonso, G., Alonso, M. & Abatzopoulos, T.J. (2017) Molecular and morphological data suggest weak phylogeographic structure in the fairy shrimp *Streptocephalus torvicornis* (Branchiopoda, Anostraca). *Hydrobiologia*, 801, 21–32. <https://doi.org/10.1007/s10750-017-3203-6>
- Korovchinsky, N.M. (2006) The Cladocera (Crustacea: Branchiopoda) as a relict group. *Zoological Journal of the Linnean Society*, 147, 109–124. <https://doi.org/10.1111/j.1096-3642.2006.00217.x>
- Korovchinsky, N.M. (2013) A new species of the genus *Diaphanosoma* Fischer, 1850 (Crustacea: Cladocera: Sididae) from Japan. *Limnology*, 14, 13–18. <https://doi.org/10.1007/s10201-012-0377-5>
- Korovchinsky, N.M., Kotov, A.A., Sinev, A.Y., Neretina, A.N. & Garibian, P.G. (2021) *Water fleas (Crustacea: Cladocera) of North Eurasia*. Vol. 2. KMK Press, Moscow, 544 pp.
- Kotov, A.A. & Boikova, O.S. (2001) Study of the late embryogenesis of *Daphnia* (Anomopoda, 'Cladocera', Branchiopoda) and a comparison of development in Anomopoda and Ctenopoda. *Hydrobiologia*, 442, 127–143. <https://doi.org/10.1023/A:1017594402114>
- Kotov, A.A. & Ferrari, F.D. (2010) The taxonomic research of Jules Richard on Cladocera (Crustacea: Branchiopoda) and his collection at the National Museum of Natural History, USA. *Zootaxa*, 2551 (1), 37. <https://doi.org/10.11646/zootaxa.2551.1.2>
- Kotov, A.A., Garibian, P.G., Bekker, E.I., Taylor, D.J. & Karabanov, D.P. (2021) A new species group from the *Daphnia curvirostris*

- species complex (Cladocera: Anomopoda) from the eastern Palaearctic: taxonomy, phylogeny and phylogeography. *Zoological journal of the Linnean Society*, 191, 772–822.  
<https://doi.org/10.1093/zoolinnean/zlaa046>
- Kotov, A.A., Sinev, A.Y., Glagolev, S.M. & Smirnov N.N. (2010) Water fleas (Cladocera). In: Alexeev, V.R. & Tsalolokhin, S.Y. (Eds.), *Key book for zooplankton and zoobenthos of fresh waters of European Russia*. KMK Press, Moscow, pp. 151–276.
- Lucas, H. (1849) *Exploration scientifique de l'Algérie pendant les années 1840, 1841, 1842. Zoologie. Vol. 1. Crustacés, etc.* Imprimerie Royale, Paris, 81 pp.
- Margaritora, F.G. (1970) Contributo alla conoscenza dei Cladoceri delle acque temporanee della Sardegna I; Chydoridae. *Rivista di Idrobiologia*, 9, 1–31.
- Margaritora, F.G. (1983) *Cladoceri (Crustacea, Cladocera). Guide per il riconoscimento delle specie animali delle acque interne italiane. Vol. 22.* Consiglio Nazionale delle Ricerche, Verona, 169 pp.
- Margaritora, F.G. (1985) *Cladocera. Fauna d'Italia. Vol. 23.* Edizioni Calderini, Bologna, 399 pp.
- Margaritora, F.G. (2005) Crustacea Branchiopoda Cladocera. *Checklist e distribuzione della fauna italiana. Memorie del Museo Civico di Storia Naturale di Verona, Serie 2, Sezione di Scienze della Vita*, 16, 87–90.
- Margaritora, F.G., Champeau, A. & Ferrara, O. (1975) Contribution à l'étude de la faune des eaux stagnantes de Corse. Les Cladocères (Crustacés). *Revue de Biologie et d'Ecologie Méditerranéenne*, 2, 3–14.
- Margaritora, F.G. & Ferrara, O. (1974) Osservazioni sistematico-biologiche su Cladoceri del genere *Daphnia* (s. str.) nelle acque astatiche della Sardegna. *Rendiconti Istituto Lombardo Accademia Di Scienze e Lettere B*, 108, 3–17.
- Margaritora, F.G., Stella, E. & Mastrantuono, L. (1977) Contributo allo studio della fauna ad Entomostraci delle acque temporanee della Turchia Asiatica. *Rivista di Idrobiologia*, 16, 151–172.
- Marková, S., Maurone, C., Racchetti, E., Bartoli, M. & Rossi, V. (2015) *Daphnia* diversity in water bodies of the Po River Basin. *Journal of Limnology*, 76 (No. 2). [published online]  
<https://doi.org/10.4081/jlimnol.2016.1531>
- Markovic, D., Carrizo, S.F., Kärcher, O., Walz, A. & David, J.N.W. (2017) Vulnerability of European freshwater catchments to climate change. *Global change biology*, 23, 3567–3580.  
<https://doi.org/10.1111/gcb.13657>
- Marrone, F., Alfonso, G., Naselli-Flores, L. & Stoch, F. (2017) Diversity patterns and biogeography of Diaptomidae (Copepoda, Calanoida) in the Western Palearctic. *Hydrobiologia*, 800, 45–60.  
<https://doi.org/10.1007/s10750-017-3216-1>
- Marrone, F., Arculeo, M., Georgiadis, C. & Stoch, F. (2019a) On the non-malacostracan crustaceans (Crustacea: Branchiopoda, Copepoda, Ostracoda) from the inland waters of Fthiotida (Greece). *Biogeographia—The Journal of Integrative Biogeography*, 34, 87–99.  
<https://doi.org/10.21426/B634043868>
- Marrone, F., Barone, R. & Naselli Flores, L. (2006) Ecological characterization and cladocerans, calanoid copepods and large branchiopods of temporary ponds in a Mediterranean island (Sicily, southern Italy). *Chemistry and Ecology*, 22, S181–S190.  
<https://doi.org/10.1080/02757540600557827>
- Marrone, F., Giuseppe, A., Stoch, F., Pieri, V., Alonso, M., Dretakis, M. & Naselli-Flores, L. (2019b) An account on the non-malacostracan crustacean fauna from the inland waters of Crete, Greece, with the synonymization of *Arctodiaptomus piliger* Brehm, 1955 with *Arctodiaptomus alpinus* (Imhof, 1885) (Copepoda: Calanoida). *Limnetica*, 38, 167–187.  
<https://doi.org/10.23818/limn.38.01>
- Marrone, F., Lo Brutto, S. & Arculeo, M. (2010) Molecular evidence for the presence of cryptic evolutionary lineages in the freshwater copepod genus *Hemidiaptomus* G.O. Sars, 1903 (Calanoida, Diaptomidae). *Hydrobiologia*, 644, 115–125.  
<https://doi.org/10.1007/s10750-010-0101-6>
- Marrone, F., Petrussek, A., Alfonso, G. & Arculeo, M. (2014) The diaptomid fauna of Israel (Copepoda, Calanoida, Diaptomidae), with notes on the systematics of *Arctodiaptomus similis* (Baird, 1859) and *Arctodiaptomus irregularis* Dimentman & Por, 1985 stat. rev. *Zoological Studies*, 53, 74.  
<https://doi.org/10.1186/s40555-014-0074-7>
- Marrone, F. & Vecchioni, L. (2021) The genus *Daphnia* in Sicily and Malta (Crustacea, Branchiopoda, Anomopoda). In: La Mantia, T., Badalamenti, E., Carapezza, A., Lo Cascio, P. & Troia, A. (Eds.), *Life on islands. 1. Biodiversity in Sicily and surrounding islands. Studies dedicated to Bruno Massa*. Edizioni Danaus, Palermo, pp. 105–123.
- Maruoka, N., Ohtsuki, H., Makino, W. & Urabe, J. (2018) Rediscovery after Almost 120 Years: Morphological and genetic evidence supporting the validity of *Daphnia mitsukuri* (Crustacea: Cladocera). *Zoological science*, 35, 468–475.  
<https://doi.org/10.2108/zs170081>
- Mouelhi, S., Balvay, G. & Kraiem, M. (2000) Branchiopodes (Cténopodes et Anomopodes) et Copépodes des eaux continentales d'Afrique du Nord: inventaire et biodiversité. *Zoosystema*, 22, 731–748.
- Muckle, R. (1951) Cladoceren aus türkische Binnegwässern I. *Review of the Faculty of Science, University of Istanbul*, Series B, 16, 367–387.
- Myers, N., Mittermeier, R.A., Mittermeier, C.G., Da Fonseca, G.A. & Kent, J. (2000) Biodiversity hotspots for conservation priorities. *Nature*, 403, 853–858.  
<https://doi.org/10.1038/35002501>

- Naidenow, W. (1994) Wandel in der Zusammensetzung der Cladocera-und Copepoda-Fauna des Süßwassers in Bulgarien im letzten Jahrhundert. *Lauterbornia*, 19, 95–106.
- Marrone, F., Barone, R. & Naselli-Flores, L. (2005) Cladocera (Branchiopoda: Anomopoda, Ctenopoda, and Onychopoda) from Sicilian inland waters: an updated inventory. *Crustaceana*, 78, 1025–1039.  
<https://doi.org/10.1163/156854005775361043>
- Negrea, Ș. (1983) Cladocera. *Fauna Republicii Socialiste România, București. Crustacea*, 4 (12), 1–399.
- Neretina, A.N., Karabanov, D.P., Sacherova, V. & Kotov, A.A. (2021) Unexpected mitochondrial lineage diversity within the genus *Alonella* Sars, 1862 (Crustacea: Cladocera) across the Northern Hemisphere. *PeerJ*, 9, e10804.  
<https://doi.org/10.7717/peerj.10804>
- Orme, C.D.L., Davies, R.G., Burgess, M., Eigenbrod, F., Pickup, N., Olson, V.A., Webster, A.J., Ding, T.-S., Rasmussen, P.C., Ridgely, R.S., Stattersfield, A.J., Bennett, P.M., Blackburn, T.M., Gaston, K.J. & Owens, I.P.F. (2005) Global hotspots of species richness are not congruent with endemism or threat. *Nature*, 436, 1016–1019.  
<https://doi.org/10.1038/nature03850>
- Parenzan, P. (1932) *Cladocera. Sistematica e corologia dei Cladoceri limnicoli italiani ed appendice sui Cladoceri in generale*. Istituto Poligrafico dello Stato, Roma, 340 pp.
- Parenzan, P. (1933) Cladoceri d’Albania. *Atti Accademia Cientifico Veneto, Trentino, Istriana, Pádua*, 22, 33–48.
- Peel, M.C., Finlayson, B.L. & McMahon, T.A. (2007) Updated world map of the Köppen-Geiger climate classification. *Hydrology and Earth System Sciences*, 11, 1633–1644.  
<https://doi.org/10.5194/hess-11-1633-2007>
- Petkovski, T.K. (1970) Zwei seltene *Daphnia*-Arten aus Mazedonien: *D. chevreuxi* Richard und *D. ulomskyi* Behning (Crustacea —Cladocera). *Fragmenta Balcanica*, 7 (15/173), 137–147.
- Petrusek, A., Hobaek, A., Nilssen, J.P., Skage, M., Černý, M., Brede, N. & Schwenk, K. (2008) A taxonomic reappraisal of the European *Daphnia longispina* complex (Crustacea, Cladocera, Anomopoda). *Zoologica Scripta*, 37, 507–519.  
<https://doi.org/10.1111/j.1463-6409.2008.00336.x>
- Petrusek, A., Tollrian, R., Schwenk, K., Haas, A. & Laforsch, C. (2009) A “crown of thorns” is an inducible defense that protects *Daphnia* against an ancient predator. *Proceedings of the National Academy of Sciences of the United States of America*, 106, 2248–2252.  
<https://doi.org/10.1073/pnas.0808075106>
- Ramdani, M. (1988) Les eaux stagnantes du Maroc: Etudes biotypologique et biogéographique du zooplancton. *Travaux Institut Scientifique Rabat, Serie Zoologie*, 43, 1–40.
- Ramos, M.A., Lobo, J.M. & Esteban, M. (2001) Ten years inventorying the Iberian fauna: results and perspectives. *Biodiversity and Conservation*, 10, 19–28.  
<https://doi.org/10.1023/A:1016658804566>
- Reniers, J., Vanschoenwinkel, B., Rabet, N. & Brendonck, L. (2013) Mitochondrial gene trees support persistence of cold tolerant fairy shrimp throughout the Pleistocene glaciations in both southern and more northerly refugia. *Hydrobiologia*, 714, 155–167.  
<https://doi.org/10.1007/s10750-013-1533-6>
- Reyjol, Y., Hugueny, B., Pont, D., Bianco, P.G., Beier, U., Caiola, N., Casals, F., Cowx, I., Economou, A., Ferreira, T., Haidvogel, G., Noble, R., Sostoa, A. de, Vigneron, T. & Virbickas, T. (2007) Patterns in species richness and endemism of European freshwater fish. *Global Ecology and Biogeography*, 16, 65–75.  
<https://doi.org/10.1111/j.1466-8238.2006.00264.x>
- Richard, J. (1887) Liste des Cladocères at des Copépodes d’eau douce observés en France. *Bulletin de la Société Zoologique de France*, 12, 156–164.
- Richard, J. (1896) Révision des Cladocères. Deuxième Partie. Anomopoda. Famille III.—Daphnidae. *Annales des Sciences Naturelles, Zoologie, Serie 8*, 8, 187–363.
- Sahuquillo, M. & Miracle, M.R. (2013) The role of historic and climatic factors in the distribution of crustacean communities in Iberian Mediterranean ponds. *Freshwater Biology*, 58, 1251–1266.  
<https://doi.org/10.1111/fwb.12124>
- Samraoui, B. (2002) Branchiopoda (Ctenopoda and Anomopoda) and Copepoda from eastern Numidia, Algeria. *Hydrobiologia*, 470, 173–179.  
<https://doi.org/10.1023/A:1015640525662>
- Sinev, A.Y., Alonso, M., Miracle, M.R. & Sahuquillo, M. (2012) The West Mediterranean *Alona azorica* Frenzel & Alonso, 1988 (Cladocera: Anomopoda: Chydoridae) is composed of two species. *Zootaxa*, 3276 (1), 51.  
<https://doi.org/10.11646/Zootaxa.3276.1.3>
- Stephanides, T. (1948) A survey of the freshwater biology of Corfu and of certain other regions of Greece. *Practika of the Hellenic Hydrobiological Institute*, 2, 1–263.
- Stewart, J.R. & Lister, A.M. (2001) Cryptic northern refugia and the origins of the modern biota. *Trends in ecology and evolution*, 16, 608–613.  
[https://doi.org/10.1016/S0169-5347\(01\)02338-2](https://doi.org/10.1016/S0169-5347(01)02338-2)
- Tierno de Figueroa, J.M., López-Rodríguez, M.J., Fenoglio, S., Sánchez-Castillo, P. & Fochetti, R. (2013) Freshwater biodiversity in the rivers of the Mediterranean Basin. *Hydrobiologia*, 719, 137–186.

<https://doi.org/10.1007/s10750-012-1281-z>

- Vecchioni, L., Arculeo, M. & Marrone, F. (2021) Molecular data attest to the occurrence of autochthonous *Daphnia pulex* (Crustacea, Branchiopoda) populations in Sicily, Italy. *Advances in Oceanography and Limnology*, 12. [published online] <https://doi.org/10.4081/aiol.2021.9947>
- Xiang, X.-F., Ji, G.-H., Chen, S.-Z., Yu, G.-L., Xu, L., Han, B.-P., Kotov, A.A. & Dumont, H.J. (2015) Annotated Checklist of Chinese Cladocera (Crustacea: Branchiopoda). Part I. Haplopoda, Ctenopoda, Onychopoda and Anomopoda (families Daphniidae, Moinidae, Bosminidae, Ilyocryptidae). *Zootaxa*, 3904 (1), 1–27. <https://doi.org/10.11646/zootaxa.3904.1.1>
- Zharov, A.A., Neretina, A.N., Rogers, D.C., Reshetova, S.A., Sinitsa, S.M. & Kotov, A.A. (2020) Pleistocene Branchiopods (Cladocera, Anostraca) from Transbaikalian Siberia demonstrate morphological and ecological stasis. *Water*, 12, 3063. <https://doi.org/10.3390/w12113063>