We are IntechOpen, the world's leading publisher of Open Access books Built by scientists, for scientists

Open access books available 5,500

International authors and editors 136,000 170M

**Downloads** 



Our authors are among the

most cited scientists TOP 1%





**WEB OF SCIENCE** 

Selection of our books indexed in the Book Citation Index in Web of Science™ Core Collection (BKCI)

# Interested in publishing with us? Contact book.department@intechopen.com

Numbers displayed above are based on latest data collected. For more information visit www.intechopen.com



#### **Chapter**

# The Orange Peel: An Outstanding Source of Chemical Resources

*Gianfranco Fontana*

# **Abstract**

*Citrus sinensis* (L.) Osbeck is a very common cultivar belonging to the *Rutaceae* family. It is largely diffused in several areas of the world characterized by mild to warm climate conditions. Its abundant worldwide production (up to  $10^7$  Tons. per year) and consumption both as the edible part of the fruit and as several types of derivative products imply the production of a huge amount of waste, such as the fruit pomace. Several ways of recycling this material have been developed in recent years: employment as fertilizer, fodder ingredient, and even cloth material. However, the chemical added value of *Citrus sinensis* peel has been underestimated despite the diversified and significant content of useful chemicals, such as polyphenols, polymethoxylated phenols, glycosylated flavonoids, volatile and non-volatile terpenoids, pectins, enzymes, etc. This work aims to highlight the outstanding chemical potential of *Citrus sinensis* peel.

**Keywords:** biological activity, *Citrus sinensis*, essential oil, flavonoids, orange peels, polymethoxyphenols

### **1. Introduction**

*Citrus sinensis* (CS) (L.) Osbeck is a perennial species growing in warm climate areas of the world and largely employed as food in form of fresh fruit, with a global production of ca. 6.7X10 $^{7}$  tons. per year (TPY) in 2016 [1], or as a processed derivative (ca.  $1.85x10<sup>7</sup>$  TPY) such as juice, marmalade, flavor, fragrance and coloring additive, pectin.

CS is an evergreen tree, 3 to 9 mts. high with sparingly barbed branches, alternate leaves with toothed blades differently shaped, oval or elliptical, connected to the stem by winged-petioles. Axillary flowers are present singly or in whorls of 6 and possess 5 white petals and up to about 25 yellow colored stamens. The pericarp of CS has a spherical or oval shape of 6–10 cm diameter with the color changing from green to yellow-orange during the ripening; the endocarp containing juice sac glands is enclosed within a wrinkled epicarp or exocarp or flavedo containing a great number of essential oil glands protected by a waxy epidermis. Below the flavedo is the albedo, also called mesocarp, a white filamentary tissue composed of tubular-like cells.

The principal industrial application of CS is the production of frozen concentrated juice. The procedure of juice extraction eventually accompanied by the extraction of the essential oil, implies the generation of a major "by-product" constituted by a pomace, mainly containing peels, accounting for up to around 60% w/w of the original fruit mass processed [2]. This huge amount of biomass does pose serious environmental concerns because of its high level of total organic carbon (TOC) and biological oxygen demand (BOD) that make disposal procedures rather complex and demanding from both the legal and industrial points of view. This is because there is an increasing trend to modify the way of approaching this problem by reconsidering the post-production orange pomace more like a by-product rather than a waste. In the last years, many producers have subjected this material to processings involving partial acidic fermentation, drying, and packaging to biologically and chemically stabilize the biomass before its application as animal feed in zootechnics, soil conditioners in agriculture, or the manufacturing of compost and biogas [2].

Beyond the standard workup of the *Citrus sinensis* peel (CSP) waste, new perspectives have been being opened in the context of the high chemical added value of the CSP [3–5] also by the complete knowledge of the rich metabolomics profile of this species. The use of CS peel has been proposed for a variety of purposes that include the production of antioxidant-enriched dietary supplements in veterinary [6], the preparation of human dietary supplements, and nutraceuticals such as citric acid [7] and flavonoids [8, 9]. The extract of CS peel is the source of a huge variety of phytochemicals and has been investigated on several applications including its chemotherapeutic and chemopreventive potential for several relevant human pathologies, such as cancer [10, 11] and obesity [12]. The extraction procedures vary in function of the main components that have to be obtained: from the simple cold pressing of pomace and the extraction with water to obtain highly hydroxylated compounds to the employment of mixtures organic protic solvent/water and finally low polar organic solvents such as Chloroform and Ethyl acetate to obtain polymethoxylated phenols (PMF, see below). New extraction technologies such as ultrasounds and microwaves may help to obtain better extraction yields.

In the following sections, the chemical structures and the biological effects of these compounds will be discussed.

#### **2. The chemistry of** *Citrus sinensis* **peel**

#### **2.1 Essential oils**

The essential oil (EO) is mainly obtained from the CS peel as a major by-product of the juice production process by a cold-pressing method that can provide the intact blend of compounds without losing the lighter, more volatile, components of the complex mixture that can be lost in the standard EO extraction procedure that is the hydrodistillation. The last one is mainly used in small scale applications, for example in research laboratories.

The chemical composition of CSP EO [13–15] is reported in **Table 1**. As it can be seen, the major component is D-Limonene, accompanied by several minor components belonging to the classes of monoterpene alkenes, oxygenated monoterpenes including alcohol aldehydes and esters, sesquiterpenes as well as linear alkanes and aldehydes. This rather complex blend accounts for the numerous deal of biological activities reported for the CSP EO [14–16], which include anthelmintic, antiaflatoxigenic [17], antibacterial [18–20], anticarcinogenic, antifungal [21], antioxidant [17], anti-tumor [22], anxiolytic [23], food preservative [24], hepatocarcinogenesis suppressant, insecticidal and larvicidal [25], pain relief and relaxant [26]. It can be argued that the main effects are due to the presence of the major component Limonene that showed several bioactivities when tested as pure compound [27]. However, it is possible that synergistic effects due to the combination of Limonene with other minor components may be speculated and should have to be demonstrated.

Comp.	Comp. name	$\%$	Compound.	Comp. name	$\frac{0}{0}$
$\mathbf{1}$	Aromadendrene	0.01	21	$\beta$ -Linalool	$0.4 - 5.6$
$\overline{2}$	δ-Amorphene	0.05	22	$\beta$ -Myrcene	$1.3 - 3.3$
3	D-Cadinene	$0.01 - 0.03$	23	Neral	$0.1 - 1.3$
$\overline{\mathbf{4}}$	δ-3-Carene	0.18	24	Neryl acetate	0.02
5	$\beta$ -Citral	$0.12 - 0.15$	25	Nonanal	$0 - 0.1$
6	$L-(+)$ -Citronellal	$0.01 - 0.1$	26	Nootkatone	0.01
$\mathcal{L}$	Citronellyl acetate	0.01	27	$cis$ - $\beta$ -Ocimene	$0.03 - 0.26$
8	$\alpha$ -Copaene	0.04	28	Octanal	$0.02 - 0.8$
9	$\alpha$ -Cubebene	$0.02 - 0.26$	29	Perillaldehyde	0.03
10	$\beta$ -Cubebene	0.03	30	$\alpha$ -Phellandrene	$0.02 - 0.07$
11	Decanal	$0.04 - 0.4$	31	$\alpha$ -Pinene	$0.49 - 0.59$
12	n-Dodecanal	0.06	32	$(+)$ -Sabinene	$0.2 - 1.0$
13	$\beta$ -Elemene	$0.01 - 0.02$	33	$\gamma$ -Terpinene	$0 - 1.21$
14	Geranial	$0 - 1.8$	34	$\gamma$ -Terpineol	$0.04 - 008$
15	Germacrene-D	$0.02 - 0.08$	35	$\alpha$ -Terpineol	$0.07 - 0.42$
16	$\beta$ -Gurjurene	0.01	36	Terpinolene	$0 - 0.08$
17	Hexadecanol	0.04	37	$\alpha$ -Thujene	0.04
18	D-Limonene	Ca. 95			
19	L-Limonene	0.02			
20	<i>trans-Limonene oxide</i>	0.01			

*The Orange Peel: An Outstanding Source of Chemical Resources DOI: http://dx.doi.org/10.5772/intechopen.96298*

#### **Table 1.**

*Composition of* C. sinensis *essential oil obtained from peels.*

### **2.2 Polyphenols**

#### *2.2.1 Flavanoids*

Polyphenols extracted from the CS peel belongs to the general structural categories of flavanones (**Figure 1a**), flavones (**Figure 1b**), flavonols (**Figure 1b**), with and without sugar moieties attached to one or more of the hydroxyl groups [28]. It is worthy of particular mention the rarely occurring class of C-glycolflavones (**Figure 1b**, compounds **63**–**65**, **85**, **86**).

These compounds are produced *in vivo* from the biogenetic mixed pathway of the Acetate and Shikimate that implies the enantiospecific formation of the basic aromatic bicyclic framework of the flavanone, from which a huge number of flavonoids originate employing selective enzymatic hydroxylations, methylations, and glycosylation steps. As can be seen from the structures shown in **Figure 1**, most of the chemical entities found in the peel extract contain several methoxy fragments that decorate the carbon skeleton. This characteristic makes those molecules to get a rather apolar character that explains their presence in the hydrophobic environment of the waxy peel. On the contrary, compounds containing a major number of hydroxyl groups are less present in the peel and are instead more significantly concentrated in the juice of the pericarp. However, some glycosylated compounds are present in the peel. In these molecules, the aglicone bears a monosaccharide unit (mainly glucose) or a disaccharide, in most of the cases being

Rutinose (**91**) – Rhamnosyl (α1 ! 6) glucose – or Neohesperidose (**92**)- Rhamnosyl  $(\alpha_1 \rightarrow 2)$  glucose (**Figure 2**).

The composition of the peel extracts described in the literature may slightly vary depending on the cultivar and the region of harvesting but some general points are





# *The Orange Peel: An Outstanding Source of Chemical Resources*

Glu: Glucose, Neohesp: Neohesperidose, Rut: Rutinose.



#### **Figure 1.** *Chemical structures of flavonoids from* C. sinensis *peels.*

common, that is the presence of the high amount of bioactive polymethoxyflavonoids [29, 30](PMF) some of which are rather ubiquitous, e.g. Nobiletin **53**, Sinensetin 51, 3',4',3,5,6,7,8-Heptamethoxyflavone 55; some other compounds



containing one to six methoxy groups in place of the hydroxyl groups are present at variable amounts. The presence of one or more residual hydroxy groups in the molecule may result in a higher bioavailability and in other general differences in their mechanism of biological and therapeutic actions [30, 31].

The biological role of these secondary metabolites in the plant is still matter of debate. It has been proposed their involvement in the mechanism of defense of the fruits exposed to the attack of phytopathogens, such as *Phytophthora citrophthora* [32].

Further, the composition of the PMF blend can be employed for the chemiotaxonomic characterization of the *Citrus* genus [33].

However, it needs to be stressed that in many cases the reported compounds were recognized by mass spectrometry and electronic spectroscopy. It is not always a matter of simplicity to discern the exact structure of a given PMF and to discriminate between different regioisomers, normally quite similar in terms of mass and electronic spectra, if an isolation procedure is not conducted and followed by a complete bi-dimensional NMR characterization. Significant differences in the extract composition do arise also in consequence of the extraction method; nonpolar solvents such as Methanol, Chloroform Ethyl acetate let to obtain PMFs-rich extracts while, on the other hand, hydroalcoholic and aqueous extracts do contain a low concentration of PMFs and a higher concentration of un-methylated polyphenols as well as glycosylated compounds.

The biological activities disclosed for the flavonoids extracted from CSP are variegated. They include antioxidant [9, 34–39], anti-inflammatory [40, 41], antimicrobial [39, 42–44], antimalarial [45], antitrypanosomal [46], cardioprotective [47], anti-osteoporosis [48], anti-ulcer [49], vascular protective [50], anti-diabetes [51, 52], hepatoprotective [53, 54], neurotrophic [55], antiadipogenesis and anti-obesity [56–58], anti-hypertensive [59], cataract prevention [60], sun protection [61], metabolic syndrome control [62]. Further, it has been demonstrated [63] that while both flavonoid set **40**, **42**, **43** and the PMFs **51**–**53** were able to inhibit the anion transportin polypeptide OATP2B1 in HEK293 cells, only the PMF group displayed this inhibitory activity also for the OATP1B1 and OATP1B3 carriers.

The most abundant PMF occurring in CSP, Nobiletin **53**, was proven to possess sevral bioactivities, such as antioxidant, anti-inflammatory, cancer preventive [64] and also a significant protective effect *in vivo* against the endotoxic shock [65] and ethanol-induced acute gastric lesions [66] in mice. Further, compound **53** demonstrated the capacity to induce autophagy in human keratinocyte HaCaT cells [67], vasodilatator effect in the rat aorta [68] and to protect the intestinal barrier from the demages induced by dextran sulfate sodium [69].

PMFs can be considered as especially promising lead compounds for cancer therapy as asignificant cytotoxic activity has been demonstrated toward a number of cancer cells [70, 71] with several mechanisms of action [72, 73]; the cytotypes investigated include MCF-7 [73–76], Hs578T triple-negative breast cancer [73, 77]; colon cancer cells CaCo-2 [19], LoVo [78], HTC-116 [79, 80] and HT-29 [79, 81]; lung cancer cells A549 [80, 82], H460 [82, 83], H1299 [82, 83]; gastric cancer cell lines AGS, BGC-823, and SGC-7901 [84]; leukemia cells HL-60 [85]. However, data regarding a possible antitumor activity *in vivo* are still rather uncommon. An interesting example is the case of the significant reduction of the intestinal tumor mass in ApcMin/+ mice treated with a CSP extract containing various PMF [86]. Further, CSP extract and pure Naringin **47** were tested for their efficacy against a YM1 esophageal cancer in an animal model [87].

Given the development of pharmacological applications of CSP extract components, further investigations are needed to better understand the bioavailability, safety, and efficacy of these compounds in humans. Most of the data reported concern *in vitro* experimentations or animal model tests. For example, the toxicity of Hesperidin **40** was evaluated [88] in Sprague Dawley rats showing a 50% lethal dose (LD50) of about 5 g/Kg body weight (BW) and a lowest-observed-adverseeffect level (LOAEL) of ca. 1 g/Kg BW.

In general, it should be emphasized as the body of evidence concerning the actual efficacy of sweet orange-derived compounds in human health is still far to be exhaustive. For example, while this work is under typewriting, a severe acute respiratory syndrome pandemic due to a COVID-19 virus is in act and a big deal of research has been being directed toward antiviral remedies and therapies. Research on nutraceuticals is not an exception and in particular some authors have shown by computational and molecular docking methods how Hesperidin **40**, the most abundant polyphenol obtained from *C. sinensis*, would be able to bind the spyke protein of this virus thus inhibiting its activity [89]. Despite their undoubted interest, these results need to be further investigated with different experimental approaches.

The pharmacological potential of pure Hesperidin **40** was also investigated for several relevant human morbidity, such as cancer, hypertension, and ulcer [90].

#### *2.2.2 Hydroxy-acids*

Several hydroxylated carboxylic acids belonging to several structural sub-classes are present foremostly in the extract obtained with mixed hydro-organic solvents, such as MeOH/water and EtOH / water [37, 38, 51, 78]; these include the aliphatic Ascorbic, Citric, Kojic, Lactic, and L-Malic acids; the aromatic 4-Hydroxybenzoic, Protocatechulic, and Gallic acids. Further, the cinnamyl compounds (**Figure 3**) Cinnamic (**93**), p-Cumaric (**94**), Caffeic (**95**), Ferulic (**96**), Sinapinic (**97**) acids, and Artepillin (**98**) were identified in some CSP extracts that showed relevant biological activities, such as antioxidant [34, 37, 38] and antidiabetes [51].

These organic acids are mainly found in free form but in some cases, they are esterified with a variety of alcoholic compounds, such as Ethanol in Ethyl gallate **99** [51], 2-Phenylethanol in Phenylethyl ester of Caffeic acid 100 [51] and (-)-Quinic acid in Chlorogenic acid **101** [51]. An interesting ester derivative (**102**) in which the anomeric hydroxyl of Glucose is esterified with a O-Caffeylsinapoyl acid unit was found in the methanolic extract of a Greek cultivar of *C. sinensis* [34].

It was shown [38] that the antioxidant properties of a CSP extract better correlated with the total phenols content (TPC) of the sample rather than with its total flavonoid content (TFC), as it can be expected from the known relevant antioxidant character of hydroxycynamic derivatives.



**Figure 3.** *Chemical structures of cinnamic acids extracted from* C. sinensis *peels.*

#### *2.2.3 Coumarins*

Coumarins are aromatic compounds biogenetically related to the o-hydroxysubstituted cynamic acids from which originate by the intramolecular condensation between the carboxylic and the o-hydroxy groups. These compounds are most commonly encountered in other species of *Citrus* taxa [91], such as *C. aurantium* (bitter orange), *C. limon*, (lemon), *C. limetta* (lime), *C. paradisi* (grapefruit) and only a few molecules of this class were Isolated from extracts of CSP endowed with activity against osteoporosis [48] and antioxidant [92]; these compounds are shown in **Figure 4**. As coumarins are relatively less common in *C. sinensis* cultivars compared to other species of the *Citrus* taxa, their rarity can be considered as a chemotaxonomic marker for *C. sinensis*.

#### *2.2.4 Catechins*

The NADPH dependent bioreduction of flavanols is the biogenetic origin of this class of compounds, present as minor constituents in CSP extract possessing significant antioxidant activity [38]; they are the two enantiomeric forms Catechin **113** and Epicatechin **114**, together with Epigallocatechin **115** (**Figure 5**).









#### **Figure 4.**

*Chemical structure of coumarins extracted from* C. sinensis *peels.*

#### **2.3 Pectins**

Pectins [93] are chemically definable as complex mixtures of polyglyconic acids in which a linear polymeric backbone is structured by a series of  $\alpha$  (1  $\rightarrow$  4) linkages (**Figure 6**). The main sugar monomer is always Galacturonic acid with the presence





**Figure 5.** *Chemical structure of catechins from* C. sinensis *peels.*



**Figure 6.** *Minimal representation of a Homopolygalacturonic acid domain of the linear primary pectin structure with a 1/3 Mol. /Mol. Esterification degree.*

of possible heterogeneous domains of other sugars such as Xylogalacturonan and Rhamnogalacturonan-I. A variable amount of the free carboxy functions may be esterified with methyl groups, while the hydroxy groups at C-2 and C-3 positions of the sugar monomers may be acetylated. Even though the primary structure of the main chain is linear, a possible degree of ramification, depending on the pectin source, may also be found. The differences in the pectins composition and structures, depending on their natural source, do confer them different physio-chemical properties, such as water solubility, sol–gel concentrations, etc. On the ground of the degree of methylation of the acid moieties, pectins are classified as "low methoxyl" (LMP, -COOMe/-COOH <50% mol.) or as "high methoxyl" (> 50% mol). A simplified representation of pectin structure is given in **Figure 6**.

Pectins find many applications in the food and drug industry as a thickening and gelling agents, excipients, and colloidal stabilizers [93].

As it has been already mentioned, the extraction method does affect the structure and the properties of the final product; the traditional acidic water extraction implies a certain degree of hydrolytic deterioration, so that new extraction technologies have been being investigated to improve the quality of the final pectins, that is microwave-assisted extraction (MAE) [94] and ultrasounds assisted extraction (USAE) [35, 95].

#### **2.4 Enzymes**

As it can be easily argued, the CSP cellular system, whose genomic profile has been fully characterized [96], is the site of a complex network of enzymatic activity. Some of the enzymes of CSP have been characterized and employed in many applications.

The acetylesterase (international enzymatic classification: EC 3.1.1.6) from CSP is known since 1947 [97] and was isolated and characterized [98]. The acetylesterase activity of the partially purified enzyme was used for the removal of the acetyl group at the 3 positions of β-lactamic antibiotics **116** [98] (**Figure 7a**). Further, the whole CSP, as well as pomace from the industrial waste of the orange juice production, was successfully employed to catalyze several relevant biotransformations [99] such as the conversion of Geranyl acetate **118** to Geraniol **119** (**Figure 7b**) and the di-acetoxynaphtalene derivative **120** to the vitamin k1 precursor **121** (**Figure 7c**).



**Figure 7.** *Chemical reactions biocatalysed by enzymes from* C. sinensis *peels.*

Recently, partial purification and functional characterization of a Uronic acid oxidase from CSP was accomplished [100]; this enzyme promotes the oxidation by O<sup>2</sup> of Galacturonic acid **122** to Galactaric acid **123** (**Figure 7d**).

#### **2.5 Miscellaneous**

#### *2.5.1 Highly lipophilic compounds*

The waxy environment of flavedo in CSP does contain several long-chain saturated and unsaturated compounds: alkanes, fatty acids, waxes, higher terpenoids.

Tetracosane, Tetratriacontanoic acid, and Ethyl pentacosanoate were identified in CSP of a Pineapple variety [101]. Further, some carotenoids were identified in the CSP extract obtained with a solvent mixture composed of Ethanol, Ethyl acetate, Petroleum ether 1: 1:1 [102]. This complex blend of carotenoids includes  $\alpha$ - and β-Carotene, Phytoene, Phytofluene, (all-E)- and (9Z)-Violoxanthin, (all-E)- Neoxanthin, (13Z)-, (13Z')- and (all-E)-Lutein, (9Z)-Zeaxanthin, (all-E)-Zeaxanthin; the mono and di-esters of violaxanthin, antheroxanthins, Xanthophyll, β-Citraurin with various fatty acids, including Lauraic, Myristic, Oleic, Palmitic, Stearic. The composition of the blend has been correlated with the maturity stage of the fruit.



#### **Figure 8.**

*Primary structure of cyclic peptide isolated from the* C. sinensis *peels.*

#### *2.5.2 Peptides*

Three cyclic peptides have been isolated from the hot water extract of CSP and were structurally characterized by FAB-MS and 2D-NMR techniques [103]. Their amino-acidic sequences, including a mostly lipophlic heptapeptide **124**, a dihydroxylated heptapeptide **125,** and a Glutamate-rich octapeptide **126**, are reported in **Figure 8**.

#### **3. Conclusions**

The chemical richness of the primary and secondary metabolome of *C. sinesnis* species is undoubtedly impressive. Thousands of different compounds belonging to dozens of structural classes have been isolated and described. The most deeply investigated are sure, on one hand, the mixtures of volatile compounds composing the blend of the essential oil and, on the other hand, polyphenols, especially flavonoids.

The chemical composition of the extract from the exocarp of *C. sinensis* does differ from the composition of juice, or leaf extracts for some aspects [104]: the presence of a higher amount of more lipophilic compounds such as polymethoxyflavonoids, r carotenoids, higher alkanes; a lesser extent of lighter terpenoids, a lower content of glycosylated compounds, the absence of cyanidins and sterols.

It is also a matter of fact that several interesting bioactivities were disclosed in the last years for the *C. sinensis* extracts that have been variously associated with the well-recognized beneficial effects that regular sweet oranges consumption may have on human health. However, a great deal of research work is still needed to clarify the molecular basis and the mechanism of these chemopreventive effects and to relate them with precise chemical entities that can be recognized as valuable nutraceuticals, as it is already the case for the well-established antioxidants Ascorbic acid, Hesperidin, Hesperetin, Quercetin, etc.

#### **Conflict of interest**

The author declares no conflict of interest.

### **Author details**

Gianfranco Fontana Department of Biological, Chemical and Pharmaceutical Sciences and Technologies (STEBICEF), University of Palermo, Palermo, Italy

\*Address all correspondence to: gianfranco.fontana@unipa.it

## **IntechOpen**

© 2021 The Author(s). Licensee IntechOpen. This chapter is distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/ by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.  $\left[\text{ce}\right]$  BY

## **References**

[1] Food and Agriculture Organization of the United nations: Citrus fruit fresh and processed - Statistical Bulletin 2016 [Internet]. 2017. Available from: http:// www. http://www.fao.org/3/a-i8092e. pdf [Accessed: 2020-12-16].

[2] Tamburino V, Zama DA. I sottoprodotti dell'industria di trasformazione: il pastazzo di agrumi. In: Vacante V, editor. Citrus - Trattato di agrumicoltura. Il Sole 24 Ore-Edagricole; 2009. p. 459–470.

[3] Putnik P, Kovačević DB, Jambrak AR, Barba FJ, Cravotto G, Binello A, Lorenzo JM, Shpigelman A. Innovative "Green" and Novel Strategies for the Extraction of Bioactive Added Value Compounds from CitrusWastes—A Review. Molecules. 2017:22:680–704. DOI: 10.3390/ molecules22050680.

[4] Senit JJ, Velasco D, Gomez Manrique A, Sanchez-Barba M, Toledo LM, Santos VE, Garcia-Ochoa F, Yustos P, Ladero M. Orange peel waste upstream integrated processing to terpenes, phenolics, pectin and monosaccharides: Optimization approaches. Ind. Crops & Prod. 2019:134:370–381. DOI: 10.1016/j. indcrop.2019.03.060.

[5] Lamine M, Gargouri M, Rahali FZ, Mliki A. Recovering and Characterizing Phenolic Compounds From Citrus By-Product: A Way Towards Agriculture of Subsistence and Sustainable Bioeconomy. Waste Biomass Valor. 2020. DOI: 10.1007/s1264 9-020-01306 -9.

[6] Williams CA. Specialized dietary supplements. In: Geor RJ, Harris PA, Coenen M, editors. Equine Applied and Clinical Nutrition. Saunders; 2013. p. 351–366. DOI: 10.1016/C2009-0- 39370-8.

[7] Patel S, Shukla S. Fermentation of Food Wastes for Generation of

Nutraceuticals and Supplements. In: Frias J, Martinez-Villaluenga C, Peñas E, editors. Fermented Foods in Health and Disease Prevention. Academic Press; 2016. p. 707–734. DOI: 10.1016/ C2014-0-01734-0.

[8] Khan N, Mongas M, Urpi-sarda M, Llorach R, Andres-La Cueva C. Contribution of Bioactive Foods and Their Emerging Role in Immunomodulation, Inflammation, and Arthritis. In: Watson RR. Preedy VR, editors. Bioactive Food as Dietary Interventions for Arthritis and Related Inflammatory Diseases. Academic Press; 2013. p. 43–65. DOI: 10.1016/ C2011-0-07467-7.

[9] Rehman Z. Citrus peel extract – A natural source of antioxidant. Food Chem. 2006;99:450–454. DOI: 10.1016/ j.foodchem.2005.07.054.

[10] Abe S, Fan K, Ho CT, Ghai G, Yang K. Chemopreventive Effects of Orange Peel Extract (OPE) II. OPE Inhibits Atypical Hyperplastic Lesions in Rodent Mammary Gland. J. Med. Food 2007;10:18–24; DOI: 10.1089/ jmf.2006.0215.

[11] Lai CS, Li S, Miyauchi Y, Suzawa M, Hob CT, Pan MH. Potent anti-cancer effects of citrus peel flavonoids in human prostate xenograft tumors. Food Funct. 2013;4:944–949; DOI: 10.1039/ c3fo60037h.

[12] Tung YC, Chang WT, Li S, Wu JC, Badmeav V, Ho CT, Pan MH. Citrus peel extracts attenuated obesity and modulated gut 1 microbiota in a high-fat diet-induced obesity mice. Food Funct. 2018;9:3363–3373; 10.1039/ C7FO02066J.

[13] Qiao Y, Jun Xie B, Zhang Y, Zhang Y, Fan G, Yao XL, Pan SY. Characterization of Aroma Active Compounds in Fruit Juice and Peel Oil

of Jinchen Sweet Orange Fruit (*Citrus sinensis* (L.) Osbeck) by GC-MS and GC-O. Molecules. 2008;13:1333–1344; DOI: 10.3390/molecules13061333.

[14] Dosoky NS, Setzer WN. Biological Activities and Safety of Citrus spp. Essential Oils. Int. J. Mol. Sci. 2018;19: 1966; DOI: 10.3390/ijms19071966.

[15] Ettoumi KY, Zouambia Y, Moulai-Mostefa N. Chemical composition, antimicrobial and antioxidant activities of Algerian *Citrus sinensis* essential oil extracted by hydrodistillation assisted by electromagnetic induction heating. J Food Sci Technol DOI: 10.1007/ s13197-020-04808-5

[16] Leherbauer I, Stappen I. Selected essential oils and their mechanisms for therapeutic use against public health disorders. An overview. Z. Naturforsch. 2020;75: 205–223.

[17] Singh P, Shukla R, Prakash B, Kumar A, Singh S, Mishra PK, Dubey NK. Chemical profile, antifungal, antiaflatoxigenic and antioxidant activity of *Citrus maxima* Burm. and *Citrus sinensis* (L.) Osbeck essential oils and their cyclic monoterpene, DLlimonene. Food Chem. Toxicol. 2010;48: 1734–1740; DOI:10.1016/j. fct.2010.04.001.

[18] O'Bryan CA, Crandall PG, Chalova VI, Ricke SC. Orange Essential Oils Antimicrobial Activities against Salmonella spp. J. Food Sc. 2008;73: M264-M267; DOI: 10.1111/ j.1750-3841.2008.00790.x.

[19] Fisher K, Phillips CA. The effect of lemon, orange and bergamot essential oils and their components on the survival of Campylobacter jejuni, *Escherichia coli* O157, *Listeria monocytogenes*, *Bacillus cereus* and *Staphylococcus aureus* in vitro and in food systems. J. Appl. Microbiol. 2006; 101:1232–1240; DOI:10.1111/ j.1365-2672.2006.03035.x.

[20] Muthaiyan A, Martin EM, Natesan S, Crandall PG, Wilkinson BJ, Ricke SC. Antimicrobial effect and mode of action of terpeneless coldpressed Valencia orange essential oil on methicillin-resistant *Staphylococcus aureus*. J. Appl. Microbiol. 2012;112: 1020–1033; DOI:10.1111/ j.1365-2672.2012.05270.x.

[21] Hung PV, Chi PTL, Phi NTL. Comparison of antifungal activities of Vietnamese citrus essential oils. Nat. Prod. Res. 2013;27:506–508; DOI: 10.1080/14786419.2012.706293.

[22] Najar B, Shortrede JE, Pistelli L, Buhagiar J. Chemical Composition and in Vitro Cytotoxic Screening of Sixteen Commercial Essential Oils on Five Cancer Cell Lines. Chem. Biodiversity 2020;17:e1900478; DOI: 10.1002/ cbdv.201900478.

[23] Faturi CB, Leite JR, Alves PB, Canton AC, Teixeira-Silva F. Anxiolytic-like effect of sweet orange aroma in Wistar rats. Prog. Neuro-Psychoph. 2010;34:605–609; DOI: 10.1016/j.pnpbp.2010.02.020.

[24] Uçar Y. Antioxidant Effect of Nanoemulsions Based on Citrus Peel Essential Oils: Prevention of Lipid Oxidation in Trout. Eur. J. Lipid Sci. Technol. 2020;122:1900405–1900419; DOI: 10.1002/ejlt.201900405..

[25] Ciriminna R, Meneguzzo F, Pagliaro M. Orange Oil. In: Nollet LML, Rathore HS editors. Green Pesticides Handbook – Essential Oils for Pest Control. CRC Press; 2017. p. 215–265.

[26] Lehrner J, Eckersberger C, Walla P, Pötsch G, Deecke L. Ambient odor of orange in a dental office reduces anxiety and improves mood in female patients. Physiology & Behavior 2000; 71:83–86.

[27] Anandakumar P, Kamaraj S, Vanitha MK. D-limonene: A

multifunctional compound with potent therapeutic effects. J. Food Biochem. 2020;45:e13566; DOI: 10.1111/ jfbc.13566.

[28] Li S, Lo CY, Ho CT. Hydroxylated Polymethoxyflavones and Methylated Flavonoids in Sweet Orange (*Citrus sinensis*) Peel. J. Agric. Food Chem. 2006;54:4176–4185. DOI: 0.1021/ jf060234n.

[29] Gao Z, Gao W, Zeng SL, Li P, Liu EH. Chemical structures, bioactivities and molecular mechanisms of citrus polymethoxyflavones. J. Funct. Foods 2018;40:498–509; DOI: 10.1016/j. jff.2017.11.036.

[30] Owis AL. Citrus Polymethoxyflavones: Biofunctional Molecules of Therapeutic Interest. Studies in Nat. Prod. Chem. 2019;59: 509–530. DOI: 10.1016/B978-0- 444-64179-3.00015-3.

[31] Lai CS, Wu JC, Ho CT, Pan MH. Disease chemopreventive effects and molecular mechanisms of hydroxylated polymethoxyflavones. Biofactors 2015; 41:301–313; DOI 10.1002/biof.1236.

[32] Del Río JA, Gómez P, Báidez AG, Arcas MC, Botía JM, Ortuño A. Changes in the Levels of Polymethoxyflavones and Flavanones as Part of the Defense Mechanism of *Citrus sinensis* (Cv. Valencia Late) Fruits against Phytophthora citrophthora. J. Agric. Food Chem. 2004;52:1913–1917; DOI: 10.1021/jf035038k.

[33] Mizuno M, Iinuma M, Ohara M, Tanaka T, Iwamasa M. Chemotaxonomy of the Genus Citrus Based on Polymethoxyflavones. Chem. Pharm. Bull. 1991;39:945–949; DOI: 10.1248/ cpb.39.942.

[34] Kanaze FI, Termentzi A, Gabrieli C, Niopas I, Georgarakis M, Kokkaloua E. The phytochemical analysis and

antioxidant activity assessment of orange peel (*Citrus sinensis*) cultivated in Greece–Crete indicates a new commercial source of hesperidin. Biomed. Chromatogr. 2009;23:239–249. DOI: 10.1002/bmc.1090.

[35] Montero-Calderon A, Cortes C, Zulueta A, Frigola A, Esteve MJ. Green solvents and Ultrasound- Assisted Extraction of bioactive orange (*Citrus sinensis*) peel compounds. Scientific Reports. 2019;9:16120–16128. DOI: 10.1038/s41598-019-52717-1.

[36] Manthey JA. Fractionation of Orange Peel Phenols in Ultrafiltered Molasses and Mass Balance Studies of Their Antioxidant Levels. J. Agric. Food Chem. 2004;52:7586–7592. DOI: 0.1021/ jf049083j.

[37] Anagnostopoulou MA, Kefalas P, Kokkalou E,. Assimopoulou AN, Papageorgiou VP. Analysis of antioxidant compounds in sweet orange peel by HPLC–diode array detection– electrospray ionization mass spectrometry. Biomed. Chromatogr. 2005;19:138–148. DOI: 10.1002/ bmc.430.

[38] Liew SS, Ho WY, Yeap SK, Bin Sharifudin SA. Phytochemical composition and in vitro antioxidant activities of *Citrus sinensis* peel extracts. PeerJ. 2018;6:e5331. DOI: 10.7717/ peerj.5331.

[39] Guo C, Shan Y, Yang Z, Zhang L, Ling W, Liang Y, Ouyang Z, Zhong B, Zhang J. Chemical composition, antioxidant, antibacterial, and tyrosinase inhibition activity of extracts from Newhall navel orange (*Citrus sinensis* Osbeck cv. Newhall) peel. J Sci Food Agric 2020;100:2664–2674. DOI: 10.1002/jsfa.10297.

[40] Gosslau A, Chen KY, Ho CT, Li S. Anti-inflammatory effects of characterized orange peel extracts enrichedwith bioactive

polymethoxyflavones. Food Sci. Human Well. 2014;3:26–35; DOI:10.1016/j. fshw.2014.02.002.

[41] Hagenlocher Y, Feilhauer K, Schäffer M, Bischoff SC, Lorentz A. Citrus peel polymethoxyflavones nobiletin and tangeretin suppress LPSand IgE-mediated activation of human intestinal mast cells. Eur J Nutr 2017;56: 1609–1620; DOI 10.1007/s00394-016- 1207-z.

[42] Liu L, Xu X, Cheng D, Yao X, Pan S. Structure-Activity Relationship of Citrus Polymethoxylated Flavones and Their Inhibitory Effects on Aspergillus niger. J. Agric. Food Chem. 2012;60: 4336–4341. DOI: 10.1021/jf3012163.

[43] Shetty SB, Mahin-Syed-Ismail P, Varghese S, Thomas-George B, Kandathil-Thajuraj P, Baby O, Haleem S, Sreedhar S, Devang-Divakar D. Antimicrobial effects of *Citrus sinensis* peel extracts against dental caries bacteria: An in vitro study. J Clin Exp Dent. 2016;8:e70–7. DOI:10.4317/ jced.52493.

[44] Ortuño A, Báidez A, Gómez P, Arcas MC, Porras I, García-Lidón A, Del Río JA. Citrus paradisi and *Citrus sinensis* flavonoids: Their influence in the defence mechanism against Penicillium digitatum. Food Chem. 2006;98:351– 358; DOI:10.1016/j. foodchem.2005.06.017.

[45] Bagavan A, Rahuman AA, Kamaraj C, Kaushik NK, Mohanakrishnan D, Sahal D. Antiplasmodial activity of botanical extracts against *Plasmodium falciparum*. Parasitol Res 2011;108:1099–1109. DOI: 10.1007/s00436-010-2151-0.

[46] Nakanishi M, Hino M, Yoshimura M, Amakura Y, Nomoto H. Identification of sinensetin and nobiletin as major antitrypanosomal factors in a citrus cultivar. Exp.

Parasitol. 2019;200:24–29; DOI: 10.1016/j.exppara.2019.03.008.

[47] Lara Testai L, Calderone V. Nutraceutical Value of Citrus Flavanones and Their Implications in Cardiovascular Disease. Nutrients. 2017; 9:502–515. DOI:10.3390/nu9050502.

[48] Shalaby NMM, Abd-Alla HI, Ahmed HH, Basoudan N. Protective effect of *Citrus sinensis* and *Citrus aurantifolia* against osteoporosis and their phytochemical constituents. J. Med. Plant. Res. 2011;5:579–588.

[49] Selmi S, Rtibi K, Grami D, Sebai H, Marzouki L. Protective effects of orange (*Citrus sinensis* L.) peel aqueous extract and hesperidin on oxidative stress and peptic ulcer induced by alcohol in rat. Lipids in Health and Disease. 2017;16:152– 164. DOI: 10.1186/s12944-017-0546-y.

[50] Chen PY, Li S, Koh YC, Wu JC, Yang MJ, Ho CT, Pan MH. Oolong Tea Extract and Citrus Peel Polymethoxyflavones Reduce Transformation of L-Carnitine to Trimethylamine-N-Oxide and Decrease Vascular Inflammation in L-Carnitine Feeding Mice. J. Agric. Food Chem. 2019; 67:7869–7879. DOI: 10.1021/acs. jafc.9b03092.

[51] Sathiyabama RG, Gandhia GR, Denadaib M, Sridharanc G, Jothic G, Sasikumard P, Siqueira Quintanse JdS, Narain N, Cuevas LE, Melo Coutinho HD, Barbosa Ramose AG, Quintans-Júniore LJ, Queiroz Gurgela R. Evidence of insulin-dependent signalling mechanisms produced by *Citrus sinensis* (L.) Osbeck fruit peel in an insulin resistant diabetic animal model. Food Chem. Toxicol. 2018;116: 86–99. DOI: 10.1016/j.fct.2018.03.050.

[52] Ahmeda OM, Hassanb MA, Abdel-Twabc SM, Abdel Azeem MN. Navel orange peel hydroethanolic extract, naringin and naringenin have anti-diabetic potentials in type 2 diabetic rats. Biomed.

& Pharmacotherapy 2017;94:197–205. DOI: 10.1016/j.biopha.2017.07.094.

[53] Ahmed OM, Fahim H, Ahmed HY, Al-Muzafar HM, Ahmed RR , Amin KA, El-Nahass ES, Abdelazeem WH. The Preventive Effects and the Mechanisms of Action of Navel Orange Peel Hydroethanolic Extract, Naringin, and Naringenin in N-Acetyl-p-aminophenol-Induced Liver Injury in Wistar Rats. Oxidative Medicine and Cellular Longevity. 2019; DOI: 10.1155/2019/ 2745352.

[54] Kim TW, Lee DR, Choi BK, Kang HK, Jung JY, Lim SW, Hwan Yang S, Suh JW. Hepatoprotective effects of polymethoxyflavones against acute and chronic carbon tetrachloride intoxication. Food Chem. Toxicol. 2016; 91: 91–99; DOI: 10.1016/j.fct.2016.03.004.

[55] Chiu SP, Wu MJ, Chen PY, Ho YR, Tai MH, Ho CT, Yen JH. Neurotrophic action of 5-hydroxylated polymethoxyflavones: 5 demethylnobiletin and gardenin A stimulate neuritogenesis in PC12 cells. J. Agric. Food Chem. 2013;61:9453–9463; DOI: 10.1021/jf4024678.

[56] Lai CS, Ho MH, Tsai ML, Li S, Badmaev V, Ho CT, Pan MH. Suppression of Adipogenesis and Obesity in High-Fat Induced Mouse Model by Hydroxylated Polymethoxyflavones. J. Agric. Food Chem. 2013;61:10320–10328; DOI: 10.1021/jf402257t.

[57] Sergeev IN, Li S, Ho CT, Rawson NE, Dushenkov S. Polymethoxyflavones Activate Ca2+- Dependent Apoptotic Targets in Adipocytes. J. Agric. Food Chem. 2009; 57:5771–5776; DOI:10.1021/jf901071k.

[58] Yu Wang Y, Lee PS, Chen YF, Ho CT, Pan MH. Suppression of Adipogenesis by 5-Hydroxy-3,6,7,8,30,40-Hexamethoxyflavone from Orange Peel in 3T3-L1 Cells. J. Med. Food 2016;19:830–835; DOI: 10.1089/jmf.2016.0060.

[59] Li GJ, Wang J, Cheng YJ, Tan X, Zhai YL, Wang Q, Gao FJ, Liu GL, Xin Zhao X, Hua Wang H. Prophylactic Effects of Polymethoxyflavone-Rich Orange Peel Oil on N<sup>o</sup>-Nitro-L-Arginine-Induced Hypertensive Rats. Appl. Sci. 2018;8:752–768; DOI:10.3390/ app8050752.

[60] Miyata Y, Oshitari T, Okuyama Y, Shimada A, Takahashi H, Natsugari H, Kosano H. Polymethoxyflavones as agents that prevent formation of cataract: Nobiletin congeners show potent growth inhibitory effects in human lens epithelial cells. Bioorg. Med. Chem. Lett. 2013;23:183–187; DOI: 10.1016/j.bmcl.2012.10.133.

[61] Li G, Tan F, Zhang Q, Tan A, Cheng Y, Zhou Q, Liu M, Tan X, Huang L, Rouseff R, Wu H, Zhao X, Liang G, Zhao X. Protective effects of polymethoxyflavone-rich cold-pressed orange peel oil against ultraviolet Binduced photoaging on mouse skin. J. Funct. Foods 2020; 67:103834– 103844; DOI: 10.1016/j.jff.2020. 103834.

[62] Zeng SL, Li SZ, Xiao PT, Cai YY, Chu C, Chen BZ, Li P, Li J, Liu EH. Citrus polymethoxyflavones attenuate metabolic syndrome by regulating gut microbiome and amino acid metabolism. Sci. Adv. 2020;6: eaax6208; DOI: 10.1126/sciadv.aax6208.

[63] Bajraktari-Sylejmani G, Weiss J. Potential Risk of Food-Drug Interactions: Citrus Polymethoxyflavones and Flavanones as Inhibitors of the Organic Anion Transporting Polypeptides (OATP) 1B1, 1B3, and 2B1. Eur. J. Drug Metab. Pharm. 2020;45:809–815; DOI: 10.1007/ s13318-020-00634-4.

[64] Li S, Wang H, Guo L, Zhao H, Ho CT. Chemistry and bioactivity of nobiletin and its metabolites. J. Funct. Food 2014;6;2–10. DOI: 10.1016/j. jff.2013.12.011.

[65] Li W, Wang X, Niu X, Zhang H, He Z, Wang Y,1 Zhi W, Liu F. Protective Effects of Nobiletin Against Endotoxic Shock in Mice Through Inhibiting TNF- $\alpha$ , IL-6, and HMGB1 and Regulating NF-κB Pathway. Inflammation 2016;39:786–797. DOI: 10.1007/s10753-016-0307-5.

[66] Li W, Wang X, Zhi W, Zhang H, He Z, Wang Y, Liu F, Niu X, Zhang X. The gastroprotective effect of nobiletin against ethanol-induced acute gastric lesions in mice: impact on oxidative stress and inflammation. Immunopharm. Immunot. 2017;39:354– 363; DOI: /10.1080/ 08923973.2017.1379088.

[67] Abe S, Hirose S, Nishitani M, Yoshida I, Tsukayama M, Tsuji A, Yuasa K. Citrus peel polymethoxyflavones, sudachitin and nobiletin, induce distinct cellular responses in human keratinocyte HaCaT cells. Biosci. Biotechnol. Biochem. 2018; 82:2064–2071; DOI: 10.1080/ 09168451.2018.1514246.

[68] Kaneda H, Otomo R, Sasaki N, Omi T, Sato T, Kaneda T. Endotheliumindependent vasodilator effects of nobiletin in rat aorta. J. Pharmacol. Sci. 2019;140:48–53; DOI: 10.1016/j. jphs.2019.04.004.

[69] Wen X, Zhao H, Wang L, Wang L, Du G, Guan W, Liu J, Cao X, Jiang X, Tian J, Wang M, Ho CT, Li S. Nobiletin Attenuates DSS-Induced Intestinal Barrier Damage through the HNF4α-Claudin-7 Signaling Pathway. J. Agric. Food Chem. 2020;68:4641–4649; DOI: 10.1021/acs.jafc.0c01217.

[70] Koolaji N, Shammugasamy B, Schindeler A, Dong Q, Dehghani F, Valtchev *P. citrus* Peel Flavonoids as Potential Cancer Prevention Agents. Current Developments In Nutrition. 2020;4:ID nzaa025;DOI: 10.1093/cdn/ nzaa025.

[71] Tung, YC, Chou YC, Hung WL, Cheng AC, Yu RC, Ho CT, Pan MH. Polymethoxyflavones: Chemistry and Molecular Mechanisms for Cancer Prevention and Treatment. Curr. Pharmacol. Rep. 2019;5:98–113; DOI: 10.1007/s40495-019-00170-z.

[72] Wang L, Wang J, Fang L, Zheng Z, Zhi D, Wang S, Li S, Ho CT, Zhao H. Anticancer Activities of Citrus Peel Polymethoxyflavones Related to Angiogenesis and Others. Biomed. Res. Int. 2014; Article ID 453972; DOI: 10.1155/2014/453972.

[73] Chan EWC, Soo OYM, Tan YH, Wong SK, Chan HT. Nobiletin and tangeretin (citrus polymethoxyflavones): an overview on their chemistry, pharmacology and cytotoxic activities against breast cancer. J. Chin. Pharm. Sci. 2020;29: 443–454; DOI: 10.5246/ jcps.2020.07.042.

[74] Sergeev IN, Ho CT, Li S, Colby J, Dushenkov S. Apoptosis-inducing activity of hydroxylated polymethoxyflavones and polymethoxyflavones from orange peel in human breast cancer cells. Mol. Nutr. Food Res. 2007;51:1478–1484. DOI: 10.1002/mnfr.200700136.

[75] Rahideh ST, Keramatipour M, Nourbakhsh M, Koohdani F, Hoseini M, Talebi S, Shidfar F. Comparison of the effects of Nobiletin and Letrozole on the activity and expression of aromatase in MCF-7 breast cancer cell line. Biochem. Cell. Biol. 2017;95:468–473;DOI: 10.1139/bcb-2016-0206.

[76] Sergeev IN, Li S, Colby J, Ho CT, Dushenkov S. Polymethoxylated

flavones induce Ca2+-mediated apoptosis in breast cancer cells. Life Sci. 2006;80:245–253; DOI: 10.1016/j. lfs.2006.09.006.

[77] Borah N, Gunawardana S, Torres H, McDonnell S, Van slambrouck S. 5,6,7,3',4',5'-Hexamethoxyflavone inhibits growth of triple-negative breast cancer cells via suppression of MAPK and Akt signaling pathways and arresting cell cycle. Int. J. Oncol. 2017; 51:1685–1693; DOI: 10.3892/ ijo.2017.4157.

[78] Ademosun AO, Oboh G, Passamonti S, Tramer F, Ziberna L, Augusti Boligon A, Athayde ML. Inhibition of metalloproteinase and proteasome activities in colon cancer cells by citrus peel extracts. J Basic Clin Physiol Pharmacol 2015;26: 471–477. DOI: 10.1515/jbcpp-2013-0127.

[79] Kaur J, Kaur G. An insight into the role of citrus bioactives in modulation of colon cancer. J. Funct. Foods. 2015;13: 239–261;DOI: 10.1016/j.jff.2014.12.043.

[80] Qiu P, Cui Y, Xiao H, Han Z, Ma H, Tang Y, Xu H, Zhang L. 5-Hydroxy polymethoxyflavones inhibit glycosaminoglycan biosynthesis in lung and colon cancer cells. J. Funct. Foods. 2017;30:39–47. DOI: 10.1016/j. jff.2017.01.008.

[81] Silva I, Estrada MF, Pereira CV, Bento da Silva A, Bronze MR, Alves PM,. Duarte CMM, Brito C, Serra AT. Polymethoxylated Flavones from Orange Peels Inhibit Cell Proliferation in a 3D Cell Model of Human Colorectal Cancer. Nutrition and Cancer. 2018;70:257–266. DOI: 10.1080/01635581.2018.1412473.

[82] Charoensinphon N, Qiu P, Dong P, Zheng J, Ngauv P, Cao Y, Li S, Ho CT, Xiao H. 5-Demethyltangeretin inhibits human nonsmall cell lung cancer cell growth by inducing G2/M cell cycle

arrest and apoptosis. Mol. Nutr. Food Res. 2013;57:2103–2111; DOI: 10.1002/ mnfr.201300136.

[83] Xiao H, Yang CS, Li S, Jin H, Ho CT, Patel T. Monodemethylated polymethoxyflavones from sweet orange (*Citrus sinensis*) peel inhibit growth of human lung cancer cells by apoptosis. Mol. Nutr. Food Res. 2009;53: 398–406; DOI 10.1002/ mnfr.200800057.

[84] Wang Y, Chen Y, Zhang H, Chen J, Cao J, Chen Q, Li X, Sun C. Polymethoxyflavones from citrus inhibited gastric cancer cell proliferation through inducing apoptosis by upregulating RARβ, both in vitro and in vivo. Food Chem. Toxicol. 2020;146: 111811–111821; DOI: 10.1016/j. fct.2020.111811.

[85] Li S, Pan MH, Lai CS, Lo CY, Dushenkovc S, Ho CT. Isolation and syntheses of polymethoxyflavones and hydroxylated polymethoxyflavones as inhibitors of HL-60 cell lines. Bioorg. Med. Chem. 2007;15:3381–338. DOI: 10.1016/j.bmc.2007.03.021.

[86] Fan K, Kurihara N, Abe S, Ho CT, Ghai G, Yang K. Chemopreventive Effects of Orange Peel Extract (OPE) I. OPE Inhibits Intestinal Tumor Growth in ApcMin+ Mice. Med. Food 2007;10:11–17. DOI: 10.1089/ jmf.2006.0214.

[87] Tajaldinia M, Samadia F, Khosravib A, Ghasemnejadd A, Asadie J. Polymethoxylated Protective and anticancer effects of orange peel extract and naringin in doxorubicin treated esophageal cancer stem cell xenograft tumor mouse model. Biomed. Pharm. 2020;121:e109594. DOI: 10.1016/j. biopha.2019.109594.

[88] Lia Y, Kandhare AD, Mukherjee AA, Bodhankar SL. Acute

and sub-chronic oral toxicity studies of hesperidin isolated from orange peel extract in Sprague Dawley rats. Regul. Toxicol Pharm. 2019;105:77–85. DOI: 10.1016/j.yrtph.2019.04.001.

[89] Bellavite P, Donzelli A. Hesperidin and SARS-CoV-2: New Light on the Healthy Function of Citrus Fruits. Antioxidants. 2020;9:742–760. DOI: 10.3390/antiox9080742.

[90] Ganeshpurkar A, Saluja A. The pharmacological potential of hesperidin. Indian J. Biochem. Biophys. 2019;56: 287–300.

[91] Murray RDH. Naturally Occurring Plant Cumarins. In: Herz W, Grisebach H, Kirby GW, editors. Progress in the Chemistry of Organic Natural Products. Springer-Verlag. New York; 1978. p. 199–400.

[92] Mohamed TK. Chemical constituents and antioxidant activity of *Citrus paradisi* (star-ruby red grapefruit) and *Citrus sinensis* (blood sweet orange) Egyptian cultivars. Asian J. Chem. 2004:887815.

[93] Cunha AG, Gandini A. Turning polysaccharides into hydrophobic materials: a critical review. Part 2. Hemicelluloses, chitin/chitosan, starch, pectin and alginates. Cellulose. 2010;17: 1045–1065. DOI: 10.1007/s10570-010- 9435-5.

[94] Sua DL, Li PJ, Quekc SY, Huang ZQ, Yuan YJ, Li GY, Shan Y. Efficient extraction and characterization of pectin from orange peel by a combined surfactant and microwave assisted process. Food Chem. 2019;286:1–7. DOI: 10.1016/j. foodchem.2019.01.200.

[95] Hosseini SS, Khodaiyan F, Kazemi M, Najari Z. Optimization and characterization of pectin extracted from sour orange peel by ultrasound assisted method. Int. J. Biol. Macromol. 2019;125:621–629. DOI: 10.1016/j. ijbiomac.2018.12.096.

[96] Jiao WB, Huang D, Xing F, Hu Y, Deng XX, Xu Q, Chen LL. Genomewide characterization and expression analysis of genetic variants in sweet oranged. The Plant Journal 2013;75:954– 964. DOI: 10.1111/tpj.12254.

[97] Jansen EF, Jang R, Mac Donnel LR. Citrus acetylesterase. Arch Biochem. 1947;15:415–431.

[98] Pasta P, Verga R, Zambianchi F, Daminati M. Acetyl Esterase from Mediterranean Oranges: Partial Purification, Immobilisation and Biotransformations. Biocatal. & Biotrasf. 2004;22:221–224. DOI: 10.1080/10242420410001697089.

[99] Fontana G, Bruno M, Maggio A, Rosselli S. Functional investigation and applications of the acetylesterase activity of the *Citrus sinensis* (L.) Osbeck peel. Nat. Prod. Res. 2020; DOI: 10.1080/14786419.2020.1737055.

[100] Wei Y, Tan YL, Ang FL, Zhao H. Identification and Characterization of Citrus Peel Uronic Acid Oxidase. ChemBioChem 2020;21:797–800. DOI: 10.1002/cbic.201900546.

[101] Rani G, Yadav L, Kalidhar SB. Chemical Examination of *Citrus sinensis* Flavedo Variety Pineapple. Indian J. Pharm. Sci. 2009;71:677–679.

[102] Lux PE, Carle R, Zacarías L, Rodrigo MJ, Schweiggert RM, Steingass CB. Genuine Carotenoid Profiles in Sweet Orange [*Citrus sinensis* (L.) Osbeck cv. Navel] Peel and Pulp at Different Maturity Stages. J. Agric. Food Chem. 2019; 67:13164–13175. DOI: 10.1021/acs.jafc.9b06098.

[103] Matsubara Y, Yusa T, Sawabe A, Itzuka Y, Takekuma S, Yoshida Y. Structures of New Cyclic Peptides in

*Citrus - Research, Development and Biotechnology*

Young Unshiu (*Citrus unshiu* Marcov.), Orange (*Citrus sinensis* Osbeck.) and Amanatzu (*Citrus natzudaidai*) Peelings. Agric. Biol. Chem. 1991;55:2923–2929.

[104] Favela-Hernández JMJ, González-Santiago O, Ramírez-Cabrera MA. Esquivel-Ferriño PC, Camacho-Corona MdR. Chemistry and Pharmacology of *Citrus sinensis*. Molecules. 2016;21, 247– 271. DOI: 10.3390/molecules21020247.

