

Behaviour of carbonyl groups in several clinical conditions: Analysis of our survey

Gregorio Caimi^{a,*}, Eugenia Hopps^a, Maria Montana^a, Caterina Carollo^a,
Vincenzo Calandrino^a, Eleonora Gallà^a, Baldassare Canino^a and Rosalia Lo Presti^b

^a*Department of Health Promotion, Mother and Child Care, Internal Medicine and Medical Specialties, Università degli Studi di Palermo, Palermo, Italy*

^b*Department of Psychology, Educational Science and Human Movement, Università degli Studi di Palermo, Palermo, Italy*

Abstract. Protein carbonylation is a marker of oxidative protein damage, that is likely involved in the pathogenesis of several diseases. The aim of this study was to evaluate the protein carbonyl (PC) groups in different clinical conditions. It included different groups of subjects: 81 trained subjects; 23 subjects with mild essential hypertension; 31 middle-aged subjects with metabolic syndrome (MS); 106 subjects with MS not selected for age (subdivided into two subgroups, with and without diabetes mellitus); 91 obese adults subdivided in two subgroups (BMI 30–35 Kg/m² and BMI > 35 kg/m²); 48 subjects with obstructive sleep apnea syndrome (OSAS) subdivided in accordance with the apnea/hypopnea index (AHI); 27 subjects with chronic kidney disease (CKD) on conservative therapy; 31 subjects with CKD on haemodialysis treatment; and 50 subjects with juvenile myocardial infarction. PC groups were reduced in trained subjects in comparison with sedentary controls, while no variation was observed in mild essential hypertension. PC groups were increased in MS subjects and in adult obese subjects. In MS subjects the PC groups were not influenced by the presence of diabetes mellitus and in adult obese subjects were not influenced by the obesity degree. In OSAS subjects only those with AHI > 30 showed an increase of PC groups. PC groups increased in CKD subjects undergoing conservative treatment and haemodialysis therapy. In dialyzed subjects, after a standard dialysis session, there was a marked increase in PC groups. In juvenile myocardial infarction PC groups were higher than in controls; there was no difference between STEMI and NSTEMI and their concentration was unaffected by the number of cardiovascular risk factors or stenosed coronary vessels.

Keywords: Oxidative stress, arterial hypertension, metabolic syndrome, juvenile myocardial infarction, chronic kidney failure

1. Introduction

An imbalance between the synthesis of reactive oxygen species (ROS) and reactive nitrogen species (RNS) and the endogenous antioxidant mechanisms induces the oxidative stress. The oxidative stress is evident in a large variety of clinical disorders and it is believed to act as a pathogenetic factor [1, 2]. While ROS include free radicals such as superoxide, hydroxyl, peroxy and hydroperoxy, RNS include nitric oxide and nitrogen dioxide.

Proteins are the principal target of ROS and RNS because they are present in high concentration in biological systems and dismiss 50–75% of generated ROS [3]. The advanced oxidation protein products (AOPPs) are produced as a consequence of the action of chlorinated compounds, leading to the production of dityrosine residues and to protein cross-linking [3]. AOPPs may maintain and amplify oxidative stress and inflammation activating neutrophils, monocytes and T lymphocytes [4]. *In vitro*, AOPPs inhibit inducible NO synthesis by macrophages [5] and induce ROS synthe-

*Corresponding author: Prof. Gregorio Caimi, Department of Health Promotion, Mother and Child Care, Internal Medicine and Medical Specialties, Università degli Studi di Palermo, Via del Vespro 129, 90100 Palermo, Italy. Tel.: +39 091 73294406; E-mail: gregorio.caimi@unipa.it.

36 sis in endothelial cells partially through NADPH oxidase activation [6]. ROS and RNS modify
37 protein conformation directly or indirectly [7]. ROS- and RNS-induced lipid peroxidation gener-
38 ates certain relatively stable products, such as malondialdehyde (MDA), hydroxynonenal (HNE),
39 oxynonenal (ONE) and isoprostanes [7]. Aminoacid modification induced by α/β unsaturated aldehy-
40 des often occurs at the nucleophilic residues of cysteine, histidine and lysine. The damaged proteins
41 cause the cleavage of the primary structure, cross-linking or variation of a single aminoacid chain
42 [7]. Oxidation of the thiol groups of cysteine residues by ROS and RNS may change the struc-
43 ture and function of proteins. Some of the ROS- and RNS-induced modifications of cysteine are
44 reversible, while other reactions with ROS and RNS develop irreversible products [8, 9]. Some
45 reversible protein changes may be protective against irreversible oxidation, but irreversible modifi-
46 cations of protein conformation may cause inhibition of enzymatic activity, increased susceptibility
47 to aggregation and altered proteolysis [10]. Oxidized proteins are almost always catabolized by
48 proteasomes and lysosomes, but some inactive proteins form aggregates inside the cells or in the
49 extracellular matrix.

50 Protein carbonylation is a marker of oxidative protein damage. Its employment as an indicator of
51 oxidative stress is useful because of the early synthesis and the stability of carbonylated proteins in com-
52 parison with other oxidation products. Carbonylation may occur through different mechanisms: direct
53 oxidation of lysine, arginine, proline and threonine residues, interaction with reactive carbonyl species
54 (RCS) produced from carbohydrate or lipid oxidation and non-oxidative reactions with dicarbonyl
55 compounds [10].

56 We examined plasma concentrations of protein carbonyl (PC) groups in several groups of subjects:
57 healthy sedentary or trained subjects, patients with mild essential hypertension, middle-aged subjects
58 with metabolic syndrome (MS), diabetic and non-diabetic subjects with MS, obese subjects, patients
59 with obstructive sleep apnea syndrome (OSAS), patients with chronic kidney disease (CKD) under-
60 going conservative treatment or haemodialysis, and a group of relatively young subjects with recent
61 acute myocardial infarction (AMI).

62 2. Materials and methods

63 2.1. We examined 9 groups of subjects.

64 The **first group** included 81 trained subjects (62 men and 19 women; mean age 31.5 ± 8.7 years;
65 BMI 23.8 ± 2.6 kg/m²). All subjects were free from cardiovascular diseases or other medical diseases.
66 None of them assumed nutritional supplements of antioxidant substances. In each subject the maximal
67 oxygen consumption (VO₂max) was determined by means of a cardiopulmonary incremental test with
68 a cycloergometer. This group was compared with a control group of sedentary 27 sedentary subjects
69 (20 men and 7 women; mean age 33.2 ± 5.6 years; BMI 24.7 ± 2.7 kg/m²). This one, as the other
70 control groups described in this study, was recruited amongst the hospital staff members and students.
71 In the whole group of trained subjects (TS) the value of VO₂max was 32.7 ± 11.2 ml/kg/min, while
72 in the group of sedentary controls it was 22.0 ± 5.6 ml/kg/min.

73 The **second group** included 23 subjects with mild essential hypertension (17 men and 6 women;
74 mean age 45.05 ± 6.14 years; range 31–53 years). The diagnosis of hypertension was based on blood
75 pressure (BP) measurements taken on two separate occasions with the patient in a seated position after
76 15 minutes of rest. The mean values of these measurements were 144/87 mmHg; only a minority of
77 the patients showed a simultaneous increase of systolic and diastolic BP above 140/90 mmHg, the
78 great majority having only either systolic or diastolic values within the range of hypertension. In this
79 group, the hypertension duration was 13.4 ± 11.9 months and the BMI was 27.04 ± 3.38 . This group

was compared with 26 healthy subjects (17 men and 9 women; mean age 43.54 ± 6.92 years; BMI 25.58 ± 4.45 kg/m²).

The **third group** included 31 middle-aged non-diabetic subjects with MS (18 men and 13 women; mean age 42.9 ± 6.5 years; BMI 31.72 ± 3.60 kg/m²; waist circumference 103 ± 8 cm; systolic BP 128.3 ± 11.3 mmHg and diastolic BP 80.8 ± 10.0 mmHg; fasting glucose level 90.0 ± 9.7 mg/dl; total cholesterol 228.1 ± 44.0 mg/dl; LDL-C 153.5 ± 42.4 mg/dl; HDL-C 37.6 ± 9.5 mg/dl; triglycerides 268.2 ± 172.6 mg/dl). This group was compared with 54 normal subjects matched by age (mean age 41.3 ± 7.4 years) and sex (35 men and 19 women).

The **fourth group** included 106 subjects with MS (mean age 53.5 ± 8.9 years; waist circumference 106.7 ± 11.2 cm, BMI 32.21 ± 4.53 kg/m²; SBP 132.1 ± 16.3 mmHg; DBP 81.2 ± 9.9 mmHg; fasting glucose level 114.3 ± 44.3 mg/dl; total cholesterol 213.9 ± 53.0 mg/dl; LDL-C 133.2 ± 46.5 mg/dl; HDL-C 40.4 ± 10.8 mg/dl; tryglicerides 220.2 ± 147.8 mg/dl), subsequently subdivided into diabetics (29 men and 14 women) and non-diabetics (32 men and 31 women). The entire group and the two subgroups were compared with a control group including 54 healthy subjects (35 men and 19 women; mean age 41.3 ± 7.4 years).

The **fifth group** included 91 subjects with obesity (58 men and 33 women; mean age 52.2 ± 11.4 years; waist circumference 115.3 ± 11.6 cm; BMI 35.64 ± 5.06 kg/m²; SBP 132.9 ± 14.6 mmHg; DBP 81.3 ± 10.4 mmHg; fasting blood glucose 115.2 ± 49.8 mg/dl; total cholesterol 208.4 ± 54.2 mg/dl; LDL-C 130.1 ± 49.0 mg/dl; HDL-C 40.5 ± 11.11 mg/dl; triglycerides 212.1 ± 154.1 mg/dl), subsequently subdivided into two subgroups according to BMI (51 with BMI 30–35 kg/m² and 40 with BMI > 35 kg/m² respectively). The group of obese subjects as well as the two subgroups were compared with a control group including 44 subjects with normal weight (26 men and 18 women, BMI < 25 kg/m²).

The **sixth group** included 48 subjects (36 men and 12 women; mean age 49.7 ± 1.6 years) with OSAS. The anthropometric characteristics and the oxygen status parameters were: BMI 35.37 ± 7.31 kg/m²; waist circumference 118.8 ± 16.1 cm; neck circumference 44.41 ± 4.53 cm; apnea/hypopnea index (AHI) 38.47 ± 25.66 ; mean nocturnal SO₂ $91.1 \pm 3.68\%$; oxygen desaturation index (ODI) 39.34 ± 29.03 . This group of OSAS subjects was subdivided according to the AHI value in two subgroups: Low ($L = 21$ subjects with AHI < 30) and high ($H = 27$ subjects with AHI > 30). The entire group and the two subgroups were compared with a control group including 59 normal subjects (43 men and 16 women; mean age 36.24 ± 8.25 years).

The **seventh group** included 27 subjects (15 men and 12 women; mean age 58.25 ± 7.6 years) with clinically stable CKD in conservative therapy, at stages 2–5 according to the KDOQI classification. In this group the cause of CKD was diabetic nephropathy in 6 patients, nephroangiosclerosis in 5, chronic glomerulonephritis in 3, polycystic kidney disease in 2; the cause was unknown in 11 patients. In this group creatinine was 3.15 ± 1.70 mg/dl, creatinine clearance 26.63 ± 17.05 ml/min, leukocyte count $7721 \pm 3092/\mu\text{l}$, haemoglobin 12.60 ± 2.15 g/dl. This group was compared with a control group of 26 subjects (17 men and 9 women; mean age 43.54 ± 6.92 years).

The **eighth group** included 31 subjects (16 men and 15 women; mean age 61.5 ± 12.8 years) with CKD on HD treatment. In this group, the cause of CKD was nephroangiosclerosis in 9 patients, diabetic nephropathy in 6, chronic glomerulonephritis in 5; the cause was unknown in 11 patients. Dialysis duration was 48.5 ± 35.7 months. This group was compared with a control group of 26 subjects (17 men and 9 women; mean age 43.54 ± 6.92 years).

The **ninth group** included 50 subjects (45 men and 5 women) aged < 46 years with recent AMI. In this group the mean age was 40.4 ± 4.8 years. The time interval between AMI onset and the examination was 13.0 ± 7.0 days. AMI subjects were subdivided according to the number of risk factors (family history of coronary artery disease, smoking, hypercholesterolemia, diabetes mellitus, arterial hypertension) into 3 subgroups: 14 of them had 0/1 risk factor, 21 had 2 risk factors and 15 had

128 3 to 5 risk factors. The 44 subjects in which coronary angiography was performed were subdivided
129 into 3 subgroups on the basis of the extent of coronary lesions: 9 subjects did not show any significant
130 coronary stenosis, 22 subjects had a single-vessel disease and 13 subjects had a multi-vessel disease.
131 The control group included 42 subjects (35 men and 7 women; mean age 38.6 ± 5.3 years) without
132 signs of acute or chronic vascular disease.

133 2.2. Protein oxidation

134 Plasma PC groups were examined using an enzyme-linked immunosorbent assay (ELISA) kit
135 (BioCell PC test kit, Enzo Life Sciences AG, Switzerland) that uses the classic PC reagent 2,4-
136 dinitrophenyl-hydrazine (DNP), which reacts with the PC forming a stable hydrazone product. In
137 brief, plasma samples were incubated with DNP, then plasma proteins were nonspecifically adsorbed
138 to the wells of an ELISA plate. Unconjugated DNP and non-protein constituents were washed away.
139 The absorbed proteins were probed with a biotinylated anti-DNP antibody, followed by streptavidin-
140 linked horseradish peroxidase. A chromatin reagent was added, and the reaction was stopped by adding
141 an acid solution. Absorbance for each well was measured at 450 nm and related to a standard curve
142 prepared for serum albumin, containing increasing proportions of hypochlorous acid-oxidized protein,
143 calibrated colorimetrically. Total protein concentration in plasma samples was evaluated by the method
144 of Lowry et al [11].

145 2.3. Statistical analysis

146 Data were expressed as means \pm SD; the difference between each group and the respective control
147 group (N) was examined according to the Student's *t* test for unpaired data. The statistical difference
148 between control subjects and TS subdivided according to the type of practiced sport (endurance, mixed
149 or power) was estimated according to the 1-way analysis of variance (ANOVA) integrated with the
150 Bonferroni test. The same statistical approach was used for obese subjects subdivided according to the
151 BMI value, for OSAS subjects subdivided according to the AHI value and for AMI subjects subdivided
152 according to the number of risk factors and the extension of coronary lesions. The evaluation of PC
153 before and after dialysis was performed using the Student's *t* test for paired data. The correlations were
154 effected using the linear regression test and the null hypothesis was rejected for *p* values > 0.05 .

155 3. Results

156 3.1. Trained subjects

157 In the whole group of TS a significant decrease in PC groups was observed [12] in comparison
158 with sedentary controls (SC) (TS = 0.351 ± 0.079 nmol/mg prot; SC = 0.405 ± 0.116 nmol/mg prot)
159 (Fig. 1). Dividing the whole group of TS according to the type of sport (endurance, mixed and power),
160 we noted a significant decrease in endurance (0.321 ± 0.045 nmol/mg prot; $p < 0.001$ vs SC) and in
161 mixed (0.330 nmol/mg prot $p < 0.01$ vs SC) but not in power TS (0.416 ± 0.072 nmol/mg prot; not
162 significant vs SC).

163 3.2. Mild essential hypertension

164 In this small group of subjects with mild essential hypertension, we did not observe [13] any variation
in PC groups in comparison with normal controls ($N = 0.440 \pm 0.134$ nmol/mg prot; Hypertensives

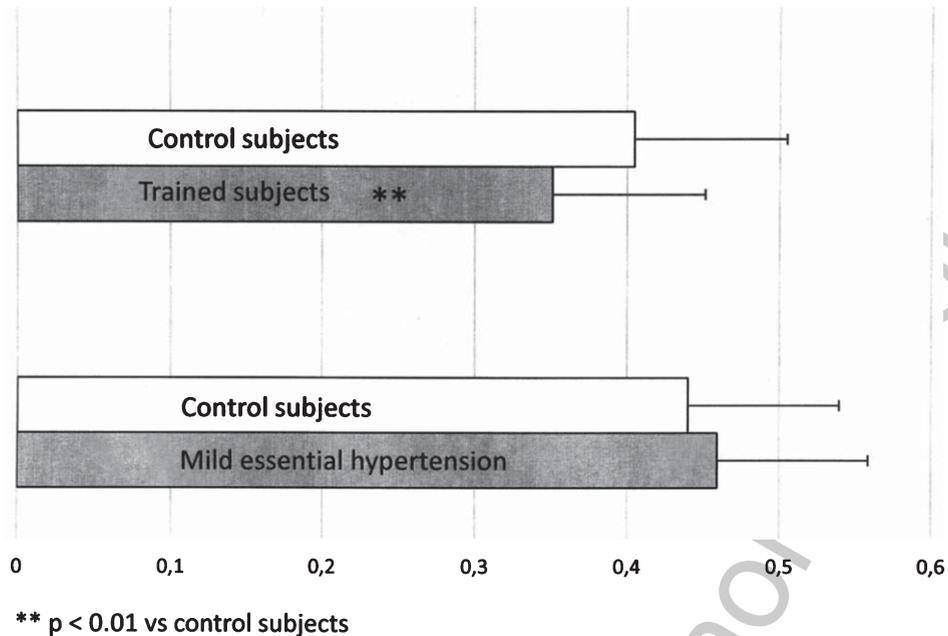


Fig. 1. Protein carbonyl groups in trained subjects and in patients with mild essential hypertension, compared with their respective control groups.

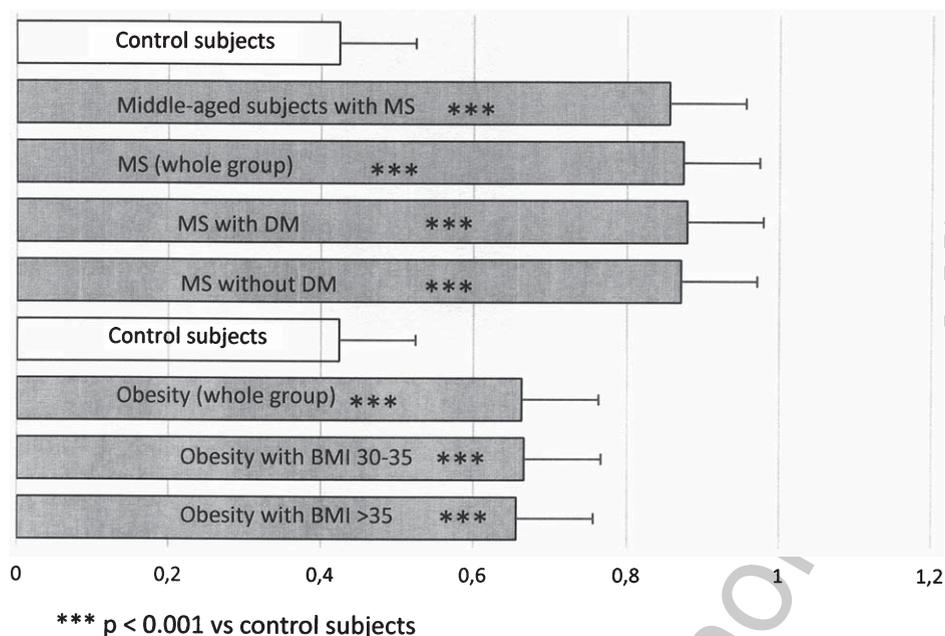
0.459 ± 0.136 nmol/mg prot) (Fig. 1). In hypertensives and in normal controls the PC were not related to age, BMI, metabolic parameters and blood pressure values.

3.3. Middle aged subjects with MS (MAMS)

In this group of middle-aged subjects with MS (MAMS), we observed [14] a significant increase in PC in comparison with normal subjects ($N = 0.424 \pm 0.139$ nmol/mg prot; MAMS = 0.856 ± 0.164 nmol/mg prot; $p < 0.001$) (Fig. 2). In this group, we only found a positive correlation between PC and total cholesterol level ($p < 0.05$).

3.4. Metabolic syndrome (MS)

In the entire group of MS subjects an increase in PC was present ($N = 0.424 \pm 0.139$ nmol/mg prot; MS = 0.874 ± 0.157 nmol/mg prot; $p < 0.001$) in comparison with normal subjects [15] (Fig. 2). The same datum was also observed in MS subjects with diabetes mellitus (DMMS) ($N = 0.424 \pm 0.139$ nmol/mg prot; DMMS = 0.879 ± 0.168 nmol/mg prot; $p < 0.001$) and in MS subjects without DM (NDMMS) ($N = 0.424 \pm 0.139$ nmol/mg prot; NDMMS = 0.871 ± 0.151 nmol/mg prot, $p < 0.001$) (Fig. 2). No difference was found, instead, between diabetic and nondiabetic subjects with MS. In the entire group of MS subjects, we observed a positive correlation between PC and HDL-cholesterol ($p < 0.05$); PC were not correlated with age, anthropometric profile, blood pressure values and metabolic pattern in DMMS, and only a positive correlation between PC and fasting blood glucose level ($p < 0.01$) in NDMMS subjects was observed.



MS = Metabolic Syndrome; DM = Diabetes mellitus; BMI = Body Mass Index

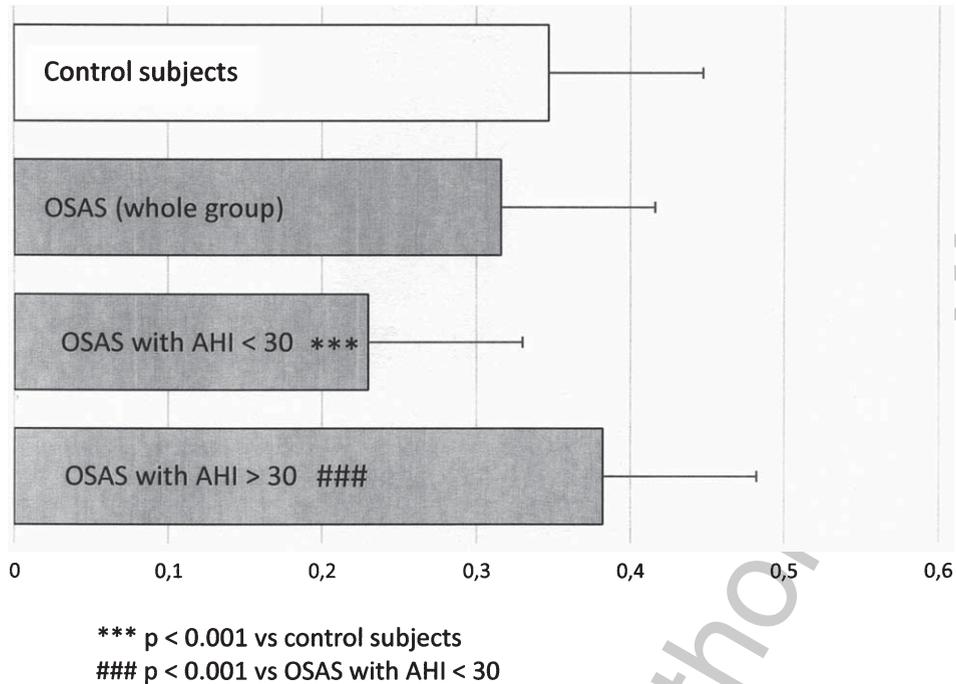
Fig. 2. Protein carbonyl groups in middle-aged subjects with metabolic syndrome, in the whole group of patients with metabolic syndrome and in obese subjects, compared with their respective control groups.

3.5. Obese subjects

In the group of obese subjects (O), we found [16] a significant increase in PC ($N=0.424 \pm 0.139$ nmol/mg prot; $O=0.663 \pm 0.314$ nmol/mg prot; $p < 0.001$) (Fig. 2). Splitting the entire group of obese subjects according to BMI (BMI 30–35 kg/m² and BMI > 35 kg/m²) and using the 1-way ANOVA analysis integrated with the Bonferroni test, we observed that the values of PC groups differed significantly ($p < 0.0001$) among normal controls, obeses with BMI 30–35 kg/m² (0.666 ± 0.318 nmol/mg prot) and obeses with BMI > 35 kg/m² (0.656 ± 0.312 nmol/mg prot) although no statistical difference was noted between the two subgroups of obeses (Fig. 2).

3.6. Obstructive sleep apnea syndrome (OSAS)

In the whole group of OSAS subjects no variation in PC groups was observed in comparison with normal controls ($N=0.347 \pm 0.094$ nmol/mg prot; OSAS = 0.316 ± 0.120 nmol/mg prot) (Fig. 3). Subdividing the subjects with OSAS in accordance with the AHI values, we found that PC were significantly increased in the H subgroup (0.382 ± 0.099 nmol/mg prot) in comparison with the L subgroup (0.230 ± 0.088 nmol/mg prot) and the entire group of OSAS subjects (0.316 ± 0.120 nmol/mg prot) (Fig. 3). In the entire group of OSAS patients, we observed a positive correlation between PC and neck circumference ($r=0.61$, $p < 0.0001$) and a positive correlation between PC and waist circumference ($r=0.35$, $p < 0.02$). We also found a positive correlation between PC and AHI ($r=0.68$, $p < 0.0001$), and between PC and ODI ($r=0.63$, $p < 0.0001$), and a negative correlation between PC and mean oxygen saturation ($r=-0.46$, $p < 0.001$) [17].



OSAS = Obstructive Sleep Apnea Syndrome; AHI = Apnea/Hypopnea Index

Fig. 3. Protein carbonyl groups in OSAS subjects, compared with the control group.

3.7. Chronic kidney disease (CKD) on conservative treatment

In this group of CKD undergoing conservative treatment (CT-CKD), we found an increase in PC in comparison with normal controls ($N = 0.440 \pm 0.134$ nmol/mg prot; CT-CKD = 0.709 ± 0.107 nmol/mg prot; $p < 0.01$) [18] (Fig. 4). In CT-CKD subjects there was a negative correlation between PC and creatinine clearance ($r = -0.46$, $p < 0.02$). PC were not correlated with creatinine levels, leukocyte count and haemoglobin level.

3.8. Chronic kidney disease (CKD) on haemodialysis treatment

In this group of CKD on haemodialysis treatment (HD-CKD), we observed a marked increase in PC in comparison with normal controls ($N = 0.440 \pm 0.134$ nmol/mg prot; HD-CKD = 1.230 ± 0.192 nmol/mg prot; $p < 0.001$) (Fig. 4). The standard haemodialysis session was followed by a further increase in PC (HD-CKD before dialysis = 1.230 ± 0.129 nmol/mg prot; HD-CKD after dialysis = 1.394 ± 0.352 nmol/mg prot; $p < 0.01$) [18] (Fig. 4).

3.9. Juvenile myocardial infarction (JMI)

In JMI, PC groups were significantly increased in comparison with normal controls ($N = 0.422 \pm 0.129$ nmol/mg prot; JMI = 0.605 ± 0.179 nmol/mg prot; $p < 0.001$) (Fig. 4). No significant difference in PC was observed between STEMI (0.593 ± 0.189 nmol/mg prot) and NSTEMI (0.594 ± 0.134 nmol/mg prot). Subdividing JMI subjects according to the number of risk factors, no significant difference was observed between the 3 subgroups; similarly, no difference was observed

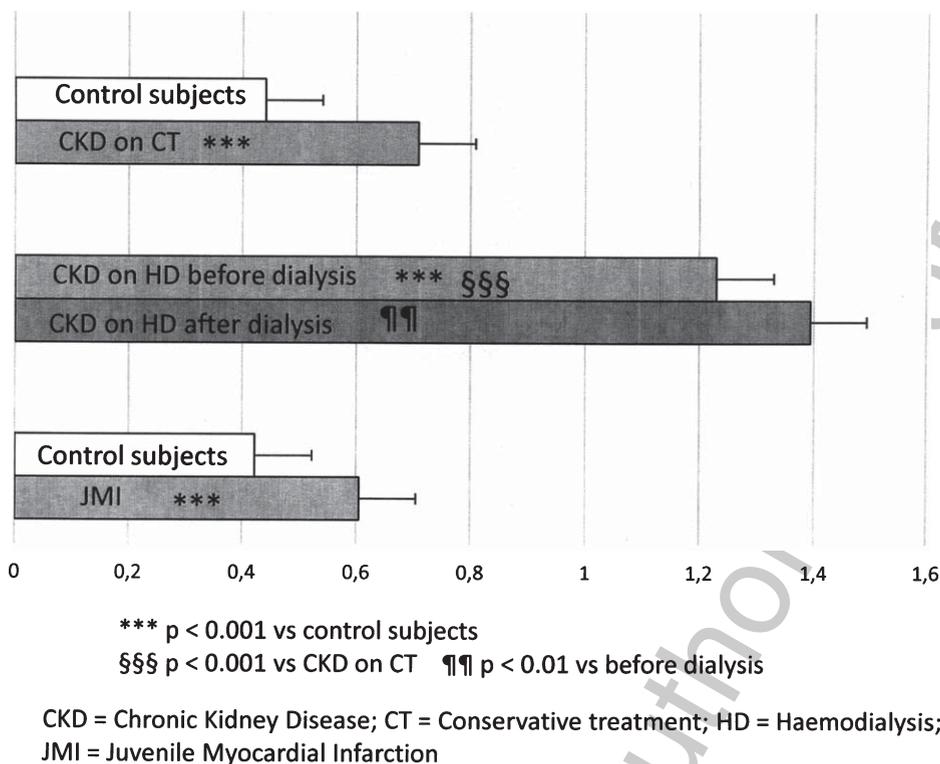


Fig. 4. Protein carbonyl groups in patients with chronic kidney disease on conservative or hemodialysis treatment and in patients with juvenile myocardial infarction, compared with their respective control groups.

219 subdividing the subjects according to the number of stenosed coronary vessels. No correlation was
 220 found between ejection fraction (evaluated by echocardiography) and PC in all JMI subjects nor in the
 221 subgroups of STEMI and NSTEMI subjects [19].

222 4. Discussion

223 In trained subjects the data of our research show a significantly lower concentration of PC in compar-
 224 ison with sedentary controls, even though the difference was significant only in those who practiced
 225 endurance and mixed sports while no variation was found in those practicing power sports. Other
 226 authors [20, 21] found a similar trend of PC groups in TS. However, none of these studies considered
 227 a comparison between the different types of training. To date, several studies have examined the vari-
 228 ation of PC groups produced by an acute bout of exercise [22–25]; only a few have tested PC groups
 229 at rest in trained versus untrained subjects. In TS at rest the protein oxidation markers are reduced,
 230 whereas the indicators of lipid peroxidation are often increased [21, 26–28]. This datum is apparently
 231 a paradox, since epidemiological studies clearly indicate that a regular physical activity is associated
 232 with a lower incidence of oxidative-related disease, such as atherosclerosis, cancer and neurodegener-
 233 ative diseases [29–31]. The simultaneous evidence of increased lipid oxidation and decreased protein
 234 oxidation, observed in TS at rest by us [27] and by other authors [21], have no univocal explanation.
 235 The cause may be a greater protection of protein molecules from oxidative damage, as well as a more
 236 effective repair and/or degradation of damaged proteins.

237 Our finding in mild essential hypertension shows no variation of PC groups in comparison with
238 normal controls. This datum is different from those observed by other authors who observed an increase
239 in PC groups in hypertensives, although it must be noted that in such research subjects with hypertension
240 were much older [32]; in other studies, the patients had higher blood pressure values [33–35]. However,
241 in the papers by Simic [35] and Nandeeshya [34], an increase in PC groups was found in subjects with
242 grade I hypertension. Recently, other authors [36] described a significant increase of PC groups in
243 sustained and white coat hypertension. Previously [37], we had found an increase in lipid peroxidation
244 in a group of hypertensive subjects, including all the subjects evaluated for plasma PC groups. This
245 contrasting datum might suggest that protein oxidation may develop at later stages during the clinical
246 course of essential hypertension; its evaluation at the early stage of the disease might underestimate
247 the impact of oxidative stress in hypertensive subjects.

248 Very interesting are the data regarding the increase of PC groups in middle-aged subjects with MS.
249 We had previously examined the role of oxidative stress in MS and the relative contribution of all the
250 MS components [38]. To date, there is a lack of significant data regarding the profile of oxidative stress
251 in middle-aged subjects with MS. None of the subjects belonging to this group had dietary restrictions
252 before testing for oxidative stress parameters and none had been taking antioxidant supplements; only
253 8 subjects out of this group were taking statins. It must be underlined that in this small group of
254 middle-aged MS subjects we also found an increase in lipid peroxidation, an increase in nitric oxide
255 metabolites and a significant decrease in total antioxidant status [38].

256 In MS patients not selected for age, there was an increase of the PC groups, unaffected by diabetes
257 mellitus. To date, the behaviour of protein oxidation has been examined mainly in the principal com-
258 ponents of MS (obesity, diabetes mellitus, hypertension and dyslipidemia); only Sebekova [39] and
259 Meaney [40] evaluated protein oxidation in MS. Previously [41], the authors of the current research
260 had undertaken the same research approach.

261 In our group of 106 MS subjects, the trend of PC groups was evaluated for the entire group, and for the
262 two subgroups with or without diabetes mellitus. PC groups were correlated with age, anthropometric
263 profile, blood pressure values and glycometabolic profile. In agreement with other authors [42, 43], we
264 found no correlation between PC groups and age, but others [44, 45] demonstrated such association.
265 Differently from other authors [33, 35, 46], we observed no correlation between PC groups and blood
266 pressure values. In agreement with other authors [47], PC groups were not statistically correlated with
267 BMI and waistline. However, others [42] found a positive correlation between protein oxidation, waist
268 circumference and waist-to-hip ratio in juvenile overweight and obesity. In the whole group of MS
269 subjects, we found a correlation between PC groups and HDL-cholesterol, not easily explainable, that
270 may be fortuitous. We observed a correlation between PC groups and fasting blood glucose levels
271 only in nondiabetic subjects with MS. Also this datum might be occasional, since no relation has been
272 observed between protein oxidation and fasting blood glucose levels in both types of diabetes [48] and
273 in severely obese children [47].

274 In our group of adult obese subjects the behaviour of protein oxidation is similar to that of lipid
275 peroxidation [16]. The alteration of the oxidative stress observed by us in obesity confirms previous
276 research papers [49–58]. This trend was observed not only in the entire group but also in the two
277 subgroups (BMI 30–35 kg/m² or BMI > 35 kg/m²). In accordance with other authors [42], a positive
278 correlation between PC groups and TBARS was found, both in the entire group and in the two subgroups
279 of adult obese subjects. The study of correlations with the anthropometric variables only showed a
280 negative correlation of uncertain meaning between PC groups and waist circumference. Some authors
281 [59] examined PC groups in a small group of severely obese patients (BMI = 48 ± 9 kg/m²) before
282 and six months after bariatric surgery, noting a marked decrease of their concentration ($p < 0.01$). In
283 clinical practice alterations in the oxidative status may be corrected by weight loss, even though other
284 therapeutic strategies deserve consideration.

285 In OSAS subjects, the behaviour of PC groups was influenced by the severity of the condition. In fact,
286 protein oxidation markedly differentiated subjects with mild to moderate OSAS from those with severe
287 OSAS. Previous research has underlined the correlation between cardiovascular complications and the
288 severity of OSAS [60–63]. In OSAS subjects we found a positive correlation between PC groups and
289 both neck and waist circumference. These findings agree with those of other authors [64] and contrast
290 with others [65]. Another aspect of our research involved the interrelationship between the PC groups
291 and the polysomnographic parameters. In contrast with some authors [66], but in agreement with
292 others [64], we found a positive correlation between PC groups and AHI values, a positive correlation
293 between PC groups and ODI and a negative correlation between PC groups and mean oxygen saturation.
294 It is known that protein oxidation is especially dependent on the hypoxia-reoxygenation episodes that
295 characterize OSAS [67, 68]. The treatment of OSAS using continuous positive airway pressure (cPAP)
296 allows the modification this parameter. An antioxidant therapy, for example consisting of vitamin C
297 and N-acetylcysteine, could be recommended in OSAS as a complementary treatment, in particular in
298 subjects with low adherence to cPAP [69].

299 For CKD in conservative treatment the increase in PC groups observed by us is in agreement
300 with the data observed by other authors [70–73]. In our small group of CT-CKD subjects, we found a
301 negative correlation between PC groups and creatinine clearance [18]. In the study by Matsuyama et al.
302 [72], a group of CT-CKD subjects was subdivided into 4 groups according to the value of creatinine
303 clearance, and a positive correlation between carbonyl stress and the degree of renal dysfunction
304 was demonstrated. Other authors [73] correlated the levels of PC groups with the estimated glomerular
305 filtration rate without finding any significant result. Other researchers [74] did not observe any variation
306 of PC groups in CT-CKD subjects compared with normal controls. Another study [75] included children
307 with CKD ($n = 65$, age range 1.4–18.6), who were subdivided according to CKD stage; only few subjects
308 had an increase of PC group concentration.

309 CKD subjects undergoing haemodialysis (HD) are at risk of high oxidative stress and systemic
310 inflammation, which aggravate the cardiovascular disease. We examined the PC groups in 31 HD-
311 CKD subjects before and after a HD session. The levels of PC groups were increased in comparison
312 with normal controls before, and especially after the HD session. The data obtained by us [18] agree
313 with those of some authors [71, 74, 76, 77] although others [70] did not find any difference in PC
314 groups before and after a standard HD session [70]. We did not note any difference in PC groups after
315 subdividing HD-CKD subjects according to their dialysis vintage or the type of filter employed for
316 HD.

317 To date, literature data are controversial about the influence of dialysis vintage on protein oxidation.
318 Some authors [78] observed increased PC groups in subject on HD longer than 12 months, while others
319 [77] did not find any variation. In the same study [77] HD was ineffective in controlling oxidative
320 stress in uremia. With regard to the filters, some authors [76] reported a decrease in PC groups after
321 using a high flux membrane, although no difference between polysulfone and cellulase membranes was
322 observed. More biocompatible and antioxidant filters, such as vitamin E-coated polysulfone membrane
323 [79, 80] have been proposed. The use of peritoneal dialysis may impact protein oxidation positively
324 [71, 81]. PC groups decreased significantly after kidney transplantation [82]. Considering the behavior
325 of protein oxidation in CKD on conservative and on HD treatment, it is useful to pursue therapeutic
326 strategies able to mitigate or to oppose such unfavourable event. In fact, it involves the activation of
327 granulocytes, monocytes and vascular endothelial cells [83] and accelerates the atherosclerotic process
328 by promoting further oxidative stress and inflammation [84].

329 In JMI there was an evident increase of PC groups, both in STEMI and NSTEMI subjects. The
330 subdivision according to the number of risk factors and the extent of coronary disease did not show
331 significant differences, although an opposite trend emerged: PC groups tended to be higher in patients
332 with more risk factors and in those with less coronary lesions. An increased protein oxidation has been

333 observed by several authors [85–89] in AMI subjects not selected for age. Our data do not confirm
334 the correlation between protein oxidation and the severity of coronary disease, which was previously
335 described by other authors [88]. There are many studies on the carbonyl scavenger activity played by
336 some molecules [90–93]; in particular, L-arginine seems to reduce the level of protein oxidation in
337 ischemic heart disease [94, 95].

338 In *conclusion*, we evaluated protein oxidation in different conditions by means of the same method.
339 PC groups were reduced in trained subjects and did not show any variation in mild essential hyperten-
340 sion. However, they were significantly higher in middle-aged MS subjects, in MS subjects non selected
341 for age and in adult obese subjects. In MS subjects the behaviour of PC groups was not influenced by
342 diabetes mellitus; similarly, in obese subjects they were unaffected by the obesity degree. In OSAS
343 subjects the PC groups were significantly increased only in the subgroup with AHI > 30. They were
344 increased in CKD subjects on conservative treatment and negatively correlated with creatinine clear-
345 ance. In CKD subjects on HD treatment, the PC groups increased significantly after the HD session.
346 In this respect, useful could be the employment of more biocompatible and antioxidant filters, such as
347 vitamin E-coated polysulfone membranes. The marked increase of PC groups observed in young AMI
348 patients is consistent with our previous findings about lipid peroxidation, total antioxidant status and
349 nitric oxide metabolites in the same clinical condition [96].

350 Conflict of interest

351 The authors declare that they have no conflict of interest regarding the publication of this paper.

352 References

- 353 [1] Frijhoff J, Winyard PG, Zarkovic N, et al. Clinical relevance of biomarkers of oxidative stress. *Antioxid Redox Signal*.
354 2015;23(14):1144-70.
- 355 [2] Hecker M, Wagner AH. Role of protein carbonylation in diabetes. *J Inherit Metab Dis*. 2018;41(1):29-38.
- 356 [3] Kalousova M, Zima T, Tesar V, et al. Advanced glycoxidation end products in chronic diseases—clinical chemistry and
357 genetic background. *Mutat Res*. 2005;579(1-2):37–46.
- 358 [4] Witko-Sarsat V, Gausson V, Nguyen A-T, et al. AOPP-induced activation of human neutrophil and monocyte oxidative
359 metabolism: A potential target for N-acetylcysteine treatment in dialysis patients. *Kidney Int*. 2003;64(1):82–91.
- 360 [5] Li ZH, Liu SX, Hou FF, Wang YQ. Effect of advanced oxidation protein products on nitric oxide production in mouse
361 peritoneal macrophages. *Nan Fang Yi Ke Da Xue Xue Bao* 2006;26(5):558-60.
- 362 [6] Yuan F, Liu SX, Tian JW. Advanced oxidation protein products induce reactive oxygen species production in endothelial
363 cells. *Di Yi Jun Yi Da Xue Xue Bao*. 2004;24(12):1350-2.
- 364 [7] Dalle Donne I, Rossi R, Colombo R, et al. Biomarkers of oxidative damage in human disease. *Clin Chem*.
365 2006;52(4):601-23.
- 366 [8] Chouchani ET, James AM, Fearnley IM, et al. Proteomic approaches to the characterization of protein thiol modification.
367 *Curr Opin Chem Biol*. 2011;15(1):120-8.
- 368 [9] Giustarini D, Rossi R, Milzani A, et al. S-Glutathionylation: From redox regulation of protein functions to human
369 diseases. *J Cell Mol Med*. 2004;8(2):201-12.
- 370 [10] Dalle-Donne I, Aldini G, Carini M, et al. Protein carbonylation, cellular dysfunction and disease progression. *J Cell*
371 *Mol Med*. 2006;10(2):389-406.
- 372 [11] Lowry OH, Rosebrough NJ, Farr AL, et al. Protein measurement with the Folin phenol reagent. *J Biol Chem*.
373 1951;193(1):265-75.
- 374 [12] Lo Presti R, Canino B, Montana M, et al. Protein carbonyl groups in trained subjects. *Clin Hemorheol Microcirc*.
375 2012;51(2):111-6.
- 376 [13] Caimi G, Mulè G, Hopps E, et al. Protein oxidation in mild essential hypertension. *Clin Hemorheol Microcirc*.
377 2012;50(3):193-5.

- 378 [14] Hopps E, Lo Presti R, Noto D, et al. Oxidative status in nondiabetic middle-aged subjects with metabolic syndrome:
379 Preliminary data. *Nutr Metab Cardiovasc Dis.* 2013;23(5):e17-8.
- 380 [15] Caimi G, Hopps E, Noto D, et al. Protein oxidation in a group of subjects with metabolic syndrome. *Diabetes Metab*
381 *Syndr.* 2013;7(1):38-41.
- 382 [16] Caimi G, Canino B, Montana M, Urso C, Calandrino V, Lo Presti R, Hopps E. Lipid peroxidation, protein oxidation,
383 gelatinases, and their inhibitors in a group of adults with obesity. *Horm Metab Res.* 2019;51(6):389-95.
- 384 [17] Hopps E, Canino B, Calandrino V, et al. Lipid peroxidation and protein oxidation are related to the severity of OSAS.
385 *Eur Rev Med Pharmacol Sci.* 2014;18(24):3773-8.
- 386 [18] Caimi G, Carollo C, Hopps E, et al. Protein oxidation in chronic kidney disease. *Clin Hemorheol Microcirc.*
387 2013;54(4):409-13.
- 388 [19] Caimi G, Canino B, Incalcaterra E, et al. Behaviour of protein carbonyl groups in juvenile myocardial infarction. *Clin*
389 *Hemorheol Microcirc.* 2013;53(4):297-302.
- 390 [20] Bloomer RJ, Fisher-Wellman KH. Blood oxidative stress biomarkers: Influence of sex, exercise training status, and
391 dietary intake. *Gend Med.* 2008;5(3):218-28.
- 392 [21] Falone S, Mirabilio A, Pennelli A, et al. Differential impact of acute bout of exercise on redox- and oxidative damage-
393 related profiles between untrained subjects and amateur runners. *Physiol Res.* 2010;59(6):953-61.
- 394 [22] Alessio HM, Hagerman AE, Fulkerson BK, et al. Generation of reactive oxygen species after exhaustive aerobic and
395 isometric exercise. *Med Sci Sports Exerc.* 2000;32(9):1576-81.
- 396 [23] Berzosa C, Gómez-Trullén EM, Piedrafita E, et al. Erythrocyte membrane fluidity and indices of plasmatic oxidative
397 damage after acute physical exercise in humans. *Eur J Appl Physiol.* 2011;111(6):1127-33.
- 398 [24] Michailidis Y, Jamurtas AZ, Nikolaidis MG, et al. Sampling time is crucial for measurement of aerobic exercise-induced
399 oxidative stress. *Med Sci Sports Exerc.* 2007;39(7):1107-13.
- 400 [25] Sentürk UK, Gündüz F, Kuru O, et al. Exercise-induced oxidative stress leads hemolysis in sedentary but not trained
401 humans. *J Appl Physiol (1985).* 2005;99(4):1434-41.
- 402 [26] Balakrishnan SD, Anuradha CV. Exercise, depletion of antioxidants and antioxidant manipulation. *Cell Biochem Funct.*
403 1998;16(4):269-75.
- 404 [27] Caimi G, Canino B, Amodeo G, et al. Lipid peroxidation and total antioxidant status in unprofessional athletes before
405 and after a cardiopulmonary test. *Clin Hemorheol Microcirc.* 2009;43(3):235-41.
- 406 [28] Marzatico F, Pansarasa O, Bertorelli L, et al. Blood free radical antioxidant enzymes and lipid peroxides following
407 long-distance and lactacidemic performances in highly trained aerobic and sprint athletes. *J Sports Med Phys Fitness.*
408 1997;37(4):235-9.
- 409 [29] Lynch BM, Neilson HK, Friedenreich CM. Physical activity and breast cancer prevention. *Recent Results Cancer Res.*
410 2011;186:13-24.
- 411 [30] Radak Z, Hart N, Sarga L, et al. Exercise plays a preventive role against Alzheimer's disease. *J Alzheimers Dis.*
412 2010;20(3):777-83.
- 413 [31] Sofi F, Capalbo A, Cesari F, et al. Physical activity during leisure time and primary prevention of coronary heart disease:
414 An updated meta-analysis of cohort studies. *Eur J Cardiovasc Prev Rehabil.* 2008;15(3):247-57.
- 415 [32] Kedziora-Kornatowska K, Czuczejko J, Pawluk H, et al. The markers of oxidative stress and activity of the antioxidant
416 system in the blood of elderly patients with essential arterial hypertension. *Cell Mol Biol Lett.* 2004;9(4A):635-41.
- 417 [33] Caner M, Karter Y, Uzun H, et al. Oxidative stress in human in sustained and white coat hypertension. *Int J Clin Pract.*
418 2006;60(12):1565-71.
- 419 [34] Nandeesh H, Sathiyapriya V, Bobby Z, et al. Altered oxidant-antioxidant status in non-obese men with moderate
420 essential hypertension. *Indian J Med Sci.* 2007;61(6):326-31.
- 421 [35] Simic DV, Mimic-Oka J, Pljesa-Ercegovac M, et al. Byproducts of oxidative protein damage and antioxidant enzyme
422 activities in plasma of patients with different degrees of essential hypertension. *J Hum Hypertens.* 2006;20(2):149-55.
- 423 [36] Yıldırım E, İpek E, Bavunoğlu I, et al. The impact of protein oxidation on sustained and white coat hypertension. *Anatol*
424 *J Cardiol.* 2017;17(3):210-16.
- 425 [37] Caimi G, Mulè G, Hopps E, et al. Nitric oxide metabolites and oxidative stress in mild essential hypertension. *Clin*
426 *Hemorheol Microcirc.* 2010;46(4):321-5.
- 427 [38] Hopps E, Noto D, Caimi G, et al. A novel component of the metabolic syndrome: The oxidative stress. *Nutr Metab*
428 *Cardiovasc Dis.* 2010;20(1):72-7.
- 429 [39] Sebeková K, Boor P, Valachovicová M, et al. Association of metabolic syndrome risk factors with selected markers of
430 oxidative status and microinflammation in healthy omnivores and vegetarians. *Mol Nutr Food Res.* 2006;50(9):858-68.
- 431 [40] Meaney E, Vela A, Samaniego V, et al. Metformin, arterial function, intima-media thickness and nitroxidation in
432 metabolic syndrome: The mefisto study. *Clin Exp Pharmacol Physiol.* 2008;35(8):895-903.
- 433 [41] Hopps E, Caimi G. Protein oxidation in metabolic syndrome. *Clin Invest Med* 2013;36(1):E1-8.

- 434 [42] Krzystek-Korpacka M, Patryn E, Boehm D, et al. Advanced oxidation protein products (AOPPs) in juvenile overweight
435 and obesity prior to and following weight reduction. *Clin Biochem.* 2008;41(12):943-9.
- 436 [43] Li G, Liu L, Hu H, et al. Age-related carbonyl stress and erythrocyte membrane protein carbonylation. *Clin Hemorheol*
437 *Microcirc.* 2010;46(4):305-11.
- 438 [44] Pandey KB, Rizvi SI. Markers of oxidative stress in erythrocytes and plasma during aging in humans. *Oxid Med Cell*
439 *Longev.* 2010;3(1):2-12.
- 440 [45] Stadtman ER. Protein oxidation in aging and age-related diseases. *Ann N Y Acad Sci.* 2001;928:22-38.
- 441 [46] Chen K, Xie F, Liu S, et al. Plasma reactive carbonyl species: Potential risk factor for hypertension. *Free Radic Res.*
442 2011;45(5):568-74.
- 443 [47] Codoñer-Franch P, López-Jaén AB, De La Mano-Hernández A, et al. Oxidative markers in children with severe obesity
444 following low-calorie diets supplemented with mandarin juice. *Acta Paediatr.* 2010;99(12):1841-6.
- 445 [48] Kalousová M, Skrha J, Zima T. Advanced glycation end-products and advanced oxidation protein products in patients
446 with diabetes mellitus. *Physiol Res* 2002;51(6):597-604.
- 447 [49] Uzun H, Konukoglu D, Gelisgen R, et al. Plasma protein carbonyl and thiol stress before and after laparoscopic gastric
448 banding in morbidly obese patients. *Obes Surg.* 2007;17(10):1367-73.
- 449 [50] Amirkhizi F, Siassi F, Minaie S, et al. Is obesity associated with increased plasma lipid peroxidation and oxidative stress
450 in women? *ARYA Atheroscler J.* 2007;2:189-192.
- 451 [51] Stefanović A, Kotur-Stevuljević J, Spasić S, et al. The influence of obesity on the oxidative stress status and the
452 concentration of leptin in type 2 diabetes mellitus patients. *Diabetes Res Clin Pract.* 2008;79(1):156-63.
- 453 [52] Tinahones FJ, Murri-Pierri M, Garrido-Sánchez L, et al. Oxidative stress in severely obese persons is greater in those
454 with insulin resistance. *Obesity (Silver Spring).* 2009;17(2):240-6.
- 455 [53] Frohnert BI, Sinaiko AR, Serrot FJ, et al. Increased adipose protein carbonylation in human obesity. *Obesity (Silver*
456 *Spring).* 2011;19(9):1735-41.
- 457 [54] Venturini D, Simão AN, Scripes NA, et al. Evaluation of oxidative stress in overweight subjects with or without metabolic
458 syndrome. *Obesity (Silver Spring).* 2012;20(12):2361-6.
- 459 [55] D'Archivio M, Annuzzi G, Vari R, et al. Predominant role of obesity/insulin resistance in oxidative stress development.
460 *Eur J Clin Invest.* 2012;42(1):70-8.
- 461 [56] Cătoi AF, Pârvu A, Galea RF, et al. Nitric oxide, oxidant status and antioxidant response in morbidly obese patients:
462 The impact of 1-year surgical weight loss. *Obes Surg.* 2013;23(11):1858-63.
- 463 [57] Gaxiola-Robles R, Bitzer-Quintero OK, Méndez-Rodríguez LC, et al. Lipid peroxidation and the response of the
464 antioxidant defense system in the obese type 2 diabetic compared with the non-obese type 2 diabetic. *Nutr Hosp.*
465 2013;28(6):1905-11.
- 466 [58] Bollineni RC, Fedorova M, Blüher M, et al. Carbonylated plasma proteins as potential biomarkers of obesity induced
467 type 2 diabetes mellitus. *J Proteome Res.* 2014;13(11):5081-93.
- 468 [59] Sledzinski T, Goyke E, Smolenski RT, et al. Decrease in serum protein carbonyl groups concentration and maintained
469 hyperhomocysteinemia in patients undergoing bariatric surgery. *Obes Surg.* 2009;19(3):321-6.
- 470 [60] Wang X, Ouyang Y, Wang Z, et al. Obstructive sleep apnea and risk of cardiovascular disease and all-cause mortality:
471 A meta-analysis of prospective cohort studies. *Int J Cardiol.* 2013;169(3):207-14.
- 472 [61] Marin JM, Carrizo SJ, Vicente E, et al. Long-term cardiovascular outcomes in men with obstructive sleep apnoea-
473 hypopnoea with or without treatment with continuous positive airway pressure: An observational study. *Lancet*
474 2005;365(9464):1046-53.
- 475 [62] Yaggi HK, Concato J, Kernan WN, et al. Obstructive sleep apnea as a risk factor for stroke and death. *N Engl J Med.*
476 2005;353(19):2034-41.
- 477 [63] Punjabi NM, Caffo BS, Goodwin JL, et al. Sleep-disordered breathing and mortality: A prospective cohort study. *PLoS*
478 *Med.* 2009;6(8):e1000132.
- 479 [64] Vatanserver E, Surmen-Gur E, Ursavas A, et al. Obstructive sleep apnea causes oxidative damage to plasma lipids and
480 proteins and decreases adiponectin levels. *Sleep Breath.* 2011;15(3):275-82.
- 481 [65] Papandreou C. Levels of TBARS are inversely associated with lowest oxygen saturation in obese patients with OSAS.
482 *Sleep Breath* 2013;17(4):1319-22.
- 483 [66] Mancuso M, Bonanni E, LoGerfo A, et al. Oxidative stress biomarkers in patients with untreated obstructive sleep apnea
484 syndrome. *Sleep Med.* 2012;13:632-6.
- 485 [67] Kent BD, Ryan S, McNicholas WT. Obstructive sleep apnea and inflammation: Relationship to cardiovascular co-
486 morbidity. *Respir Physiol Neurobiol* 2011;178(3):475-81.
- 487 [68] Lavie L, Lavie P. Molecular mechanisms of cardiovascular disease in OSAHS: The oxidative stress link. *Eur Respir J.*
488 2009;33(6):1467-84.

- 489 [69] Lira AB, de Sousa Rodrigues CF. Evaluation of oxidative stress markers in obstructive sleep apnea syndrome and
490 additional antioxidant therapy: A review article. *Sleep Breath*. 2016;20(4):1155-60.
- 491 [70] Dursun E, Dursun B, Süleymanlar G, et al. Carbonyl stress in chronic renal failure: The effect of haemodialysis. *Ann*
492 *Clin Biochem*. 2005;42(Pt 1):64-6.
- 493 [71] Li Z, Su BH, Mi XH, et al. Inflammation, oxidative stress and carbonyl stress in uremic patients. *Sichuan Da Xue Xue*
494 *Bao Yi Xue Ban*. 2006;37(1):123-5.
- 495 [72] Matsuyama Y, Terawaki H, Terada T, et al. Albumin thiol oxidation and serum protein carbonyl formation are progres-
496 sively enhanced with advancing stages of chronic kidney disease. *Clin Exp Nephrol*. 2009;13(4):308-15.
- 497 [73] Oberg BP, McMenamin E, Lucas FL, et al. Increased prevalence of oxidant stress and inflammation in patients with
498 moderate to severe chronic kidney disease. *Kidney Int*. 2004;65(3):1009-16.
- 499 [74] Rutkowski P, Małgorzewicz S, Słominska E, et al. Interrelationship between uremic toxicity and oxidative stress. *J Ren*
500 *Nutr*. 2006;16(3):190-3.
- 501 [75] Drożdż D, Kwinta P, Sztefko K, et al. Oxidative Stress Biomarkers and Left Ventricular Hypertrophy in Children with
502 Chronic Kidney Disease. *Oxid Med Cell Longev*. 2016;2016:7520231.
- 503 [76] Bordoni V, Piroddi M, Galli F, et al. Oxidant and carbonyl stress-related apoptosis in end-stage kidney disease: Impact
504 of membrane flux. *Blood Purif*. 2006;24(1):149-56.
- 505 [77] Pupim LB, Himmelfarb J, McMonagle E, et al. Influence of initiation of maintenance hemodialysis on biomarkers of
506 inflammation and oxidative stress. *Kidney Int*. 2004;65(6):2371-9.
- 507 [78] Köken T, Serteser M, Kahraman A, et al. Changes in serum markers of oxidative stress with varying periods of
508 haemodialysis. *Nephrology (Carlton)*. 2004;9(2):77-82.
- 509 [79] Andrulli S, Di Filippo S, Manzoni C, et al. Effect of synthetic vitamin E-bonded membrane on responsiveness to
510 erythropoiesis-stimulating agents in hemodialysis patients: A pilot study. *Nephron Clin Pract*. 2010;115(1):c82-9.
- 511 [80] Bargnoux AS, Cristol JP, Jaussent I, et al. Vitamin E-coated polysulfone membrane improved red blood cell antioxidant
512 status in hemodialysis patients. *J Nephrol*. 2013;26(3):556-63.
- 513 [81] Boulanger E, Moranne O, Wautier MP, et al. Changes in glycation and oxidation markers in patients starting peritoneal
514 dialysis: A pilot study. *Perit Dial Int*. 2006;26(2):207-12.
- 515 [82] Aveles PR, Criminácio CR, Gonçalves S, et al. Association between biomarkers of carbonyl stress with increased
516 systemic inflammatory response in different stages of chronic kidney disease and after renal transplantation. *Nephron*
517 *Clin Pract*. 2010;116(4):c294-9.
- 518 [83] Guo ZJ, Niu HX, Hou FF, et al. Advanced oxidation protein products activate vascular endothelial cells via a RAGE-
519 mediated signaling pathway. *Antioxid Redox Signal*. 2008;10(10):1699-712.
- 520 [84] Liu SX, Hou FF, Guo ZJ, et al. Advanced oxidation protein products accelerate atherosclerosis through promoting
521 oxidative stress and inflammation. *Arterioscler Thromb Vasc Biol*. 2006;26(5):1156-62.
- 522 [85] Bagatini MD, Martins CC, Battisti V, et al. Oxidative stress versus antioxidant defenses in patients with acute myocardial
523 infarction. *Heart Vessels*. 2011;26(1):55-63.
- 524 [86] Barsotti A, Fabbi P, Fedele M, et al. Role of advanced oxidation protein products and Thiol ratio in patients with acute
525 coronary syndromes. *Clin Biochem*. 2011;44(8-9):605-11.
- 526 [87] Feng Y, Shen C, Ma G, et al. Prolonged pain to hospital time is associated with increased plasma advanced oxidation
527 protein products and poor prognosis in patients with percutaneous coronary intervention for ST-elevation myocardial
528 infarction. *Heart Vessels*. 2010;25(5):374-8.
- 529 [88] Serdar Z, Serdar A, Altin A, et al. The relation between oxidant and antioxidant parameters and severity of acute
530 coronary syndromes. *Acta Cardiol*. 2007;62(4):373-80.
- 531 [89] Skvarilová M, Bulava A, Stejskal D, et al. Increased level of advanced oxidation products (AOPP) as a marker of
532 oxidative stress in patients with acute coronary syndrome. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub*.
533 2005;149(1):83-7.
- 534 [90] Galvani S, Coatrieux C, Elbaz M, et al. Carbonyl scavenger and antiatherogenic effects of hydrazine derivatives. *Free*
535 *Radic Biol Med*. 2008;45(10):1457-67.
- 536 [91] Li G, Tang T, Peng M, et al. Direct reaction of taurine with malondialdehyde: Evidence for taurine as a scavenger of
537 reactive carbonyl species. *Redox Rep*. 2010;15(6):268-74.
- 538 [92] Negre-Salvayre A, Coatrieux C, Ingueneau C, et al. Advanced lipid peroxidation end products in oxidative damage to
539 proteins. Potential role in diseases and therapeutic prospects for the inhibitors. *Br J Pharmacol*. 2008;153(1):6-20.
- 540 [93] Vidart J, Wajner SM, Leite RS, et al. N-acetylcysteine administration prevents nonthyroidal illness syndrome in patients
541 with acute myocardial infarction: A randomized clinical trial. *J Clin Endocrinol Metab*. 2014;99(12):4537-45.
- 542 [94] Tripathi P, Chandra M, Misra MK. Oral administration of L-arginine in patients with angina or following myocardial
543 infarction may be protective by increasing plasma superoxide dismutase and total thiols with reduction in serum
544 cholesterol and xanthine oxidase. *Oxid Med Cell Longev*. 2009;2(4):231-7.

- 545 [95] Tripathi P, Misra MK. Therapeutic role of L-arginine on free radical scavenging system in ischemic heart diseases.
546 Indian J Biochem Biophys. 2009;46(6):498-502.
- 547 [96] Lo Presti R, Carollo C, Montana M, et al. Lipid peroxidation and total antioxidant status in juvenile myocardial infarction.
548 Clin Hemorheol Microcirc. 2008;38(2):93-6.

Uncorrected Author Proof