

PhD in Pharmaceutical Sciences

"Doctor Europeus"

Synthesis and antitumor activity of new tetracyclic systems containing the pyrrole ring

Settore scientifico disciplinare CHIM/08

Dr. Barbara Parrino

TUTOR Chiar.mo Prof. Girolamo Cirrincione

PhD Coordinator: Chiar.mo Prof. Girolamo Cirrincione

CYCLE XXIII - 2009-2011

INDEX

| INTRODUCTION | pag.1 |
|---|--------|
| AIM OF THE WORK | pag.22 |
| SYNTHESIS | pag.23 |
| BIOLOGICAL DATA | pag.32 |
| EXPERIMENTAL DATA | pag.46 |
| SYNTHESIS OF VENTILATONE A (NOTTINGHAM) | pag.74 |
| EXPERIMENTAL DATA VENTILATONE A | pag.80 |
| BIBLIOGRAPHY | pag.85 |

INTRODUCTION

Cancer is a growing problem estimated to have about seven million new incidences per year in all over the world. About one in four people will get it in some form during their life time, and at the present time about one in five of all deaths are due to cancer.

Cancer is a term which describes a group of perhaps 120 different diseases which share some broad similarities. In these diseases, a single cell begins to divide uncontrollably forming a tumour that can involve also the adjacent tissues and eventually bits of this tumour break off and form new tumours (this is known as metastasis).

These features differentiate cancer cells from normal cells that are kept under tight control by a number of different biological mechanisms that are still being explored. We do know that cell division is controlled by a relatively small group of enzymes. Some of these operate to form a communication network, relaying growth signals from the surface of the cell to its DNA and telling it when to begin dividing.

Others work as a surveillance team, preventing a cell with damaged DNA from reproducing by first repairing the damage or by instructing it to die. However, sometimes the damage (mutation) occurs in the DNA that codes for these enzymes, so that they are themselves defective. Such a cell will divide uncontrollably, and produce daughter cells that do the same.

However is almost always impossible to prove exactly what caused a cancer in any individual, because most cancers have multiple possible causes; cancers are primarily an environmental disease with 90-95% of cases attributed to environmental factors and 5-10% due to genetics. *Environmental* means any cause that is not genetic, not merely pollution. Common environmental factors that contribute to cancer death include tobacco (25-30%), diet and obesity (30-35%), infections(15-20%), radiation (both ionizing and non-ionizing, up to 10%), stress, lack of physical activity, and environmental pollutants.

Most cancers are initially recognized either because signs or symptoms appear or through cancer screening but definitive diagnosis requires histological examination and evaluation of specific markers that can be useful to establish prognosis and individual treatments.

Many management options for cancer exist including: chemotherapy, radiation therapy,

surgery, immunotherapy, monoclonal antibody therapy and other methods. Which treatments are used depends upon the type of cancer, the location and grade of the tumour, and the stage of the disease, as well as the general state of a person's health.

Complete removal of the cancer without damage to the rest of the body is the goal of treatment for most cancers. Sometimes this can be accomplished by surgery, but the propensity of cancers to invade adjacent tissue or to spread to distant sites by microscopic metastasis often limits its effectiveness. Surgery often required the removal of a wide surgical margin and the effectiveness of chemotherapy is often limited by toxicity to other tissues in the body. Radiation can also cause damage to normal tissue. For these reasons cancer research is the intense scientific effort to understand disease processes and discover possible therapies.

The drugs used to combat cancer belong to one of two broad categories. The first is cytotoxic (cell killing) drugs and the second is cytostatic (cell stabilising drugs). Both categories lead to a reduction in the size of the tumour because cancer cells (for various reasons) have such a high mortality rate that simply preventing them from dividing will lead to a reduction in the population.

Cytotoxic drugs work by interfering with DNA replication. Because cancer cells are rapidly dividing, they are rapidly synthesizing new DNA and if this is damaged the cell will die.

There are three main groups of molecules that can be used to interfere with DNA replication:

• antimetabolites: molecules that appear to be nucleotides and so are incorporated into DNA, leading to non-functional DNA.

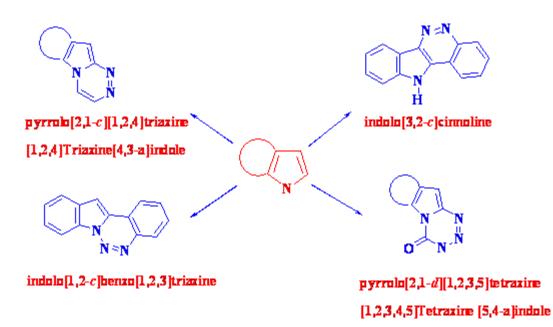
• alkylating agents: molecules that permanently attach to the DNA, distorting its shape. Unfortunately these also attach to many other molecules in cells.

• DNA-binding agents: molecules that attach to the DNA chain, break it, disengage from the chain and then attach to another chain to repeat the process. These usually function in conjunction with an enzyme.

Examples of compounds showing antitumor activity are polycondenced nitrogen heterocycles. They found infact wide range of pharmaceutical applications.

From several years the research group where I carried out my PhD thesis has been interested in the synthesis and biological evaluation of polycondensed nitrogen heterocycles, containing pyrrole or indole moieties, endowed with antineoplastic activity and, being planar systems, they potentially might intercalate between the base pairs of double-stranded DNA.

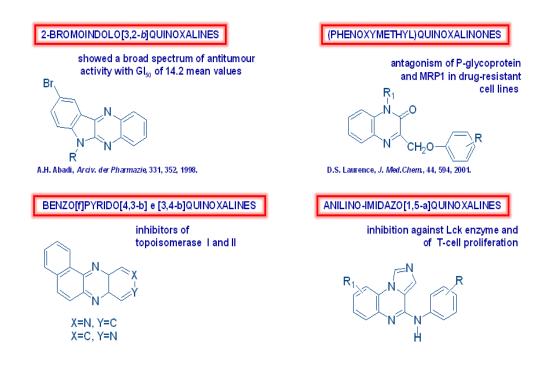
Pyrrolo[2,1-c][1,2,4]triazine, [02EJMC267] [1,2,4]triazine[4,3-a]indole, and [04ACR3775] showed antiproliferative activity with GI₅₀ at low micromolar concentrations against a wide range of human cell lines (panel of about 60 cell lines). Indolo[1,2-*c*]benzo[1,2,3]triazine, [99JMC2561] and indolo[3,2-*c*]cinnoline, [99BMC1591] exhibited potent antiproliferative activity, reaching GI₅₀ values even at nanomolar level against leukaemia cell lines and solid tumours cell lines. They also showed antibacterial activity being two magnitude orders more active than Pyrrolo[2,1-*d*][1,2,3,5]tetrazine, [03BMC2371] streptomycin. and [1,2,3,5]tetrazine[5,4-a]indole, [05BMCL295] showed as well antiproliferative activity from micromolar to nanomolar concentration.



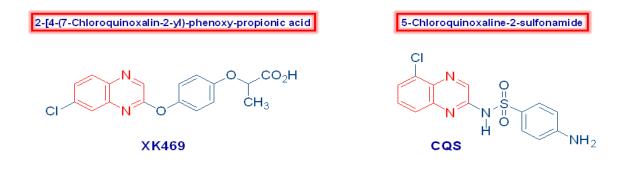
Quinoxalines represent an important class of compounds that are found in a variety of biologically and medicinally useful agents. Infact, the quinoxaline portion is the pharmacophore moiety of a large number of molecules.

For instance, substituted (phenoxymethyl)quinoxalin-2-ones demonstrated excellent antagonism of P-glycoprotein and MRP1 in drug-resistant cell lines [01JMC594];

quinoxaline ring condensed with a pyridine moiety led to benzo[f]pyrido [4,3-b] and pyrido[3,4-b]quinoxalines that are inhibitors of topoisomerase I and II [95ACDD277]. Incorporation of indole ring gave 5-substituted 2-bromoindolo[3,2-b]quinoxaline that showed a broad spectrum of antitumor activity with two biochemical mechanism-based screens (cdc2 kinase and cdc25 phosphatase) with IC₅₀ of 70 and 25 μ M respectively [98ADP352]. (Anilinophenyl)imidazo[1,5-a]quinoxalines derivatives showed inhibition against Lck enzyme and T-cell proliferation [02BMC3153].

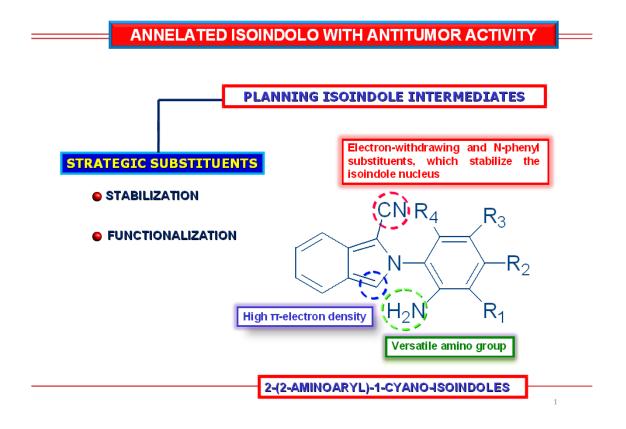


Some quinoxaline derivatives found their way in clinical application. For example the two known antineoplastic quinoxaline topoisomerase II inhibitors, chloroquinoaline-phenoxy-propionic acid (**XK469**) [07IND147], [08EJC1684], [08IND331] and chloroquinoxalinesulfonamide (**CQS**) [06IND343].

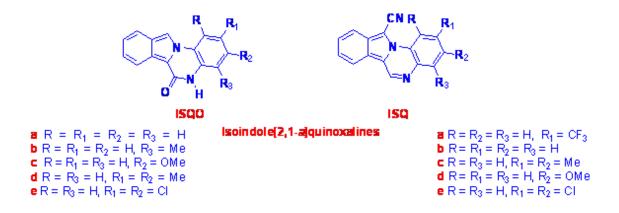


For all these reasons, in the last years, our research group have been interested to the synthesis of isoindolo[1,2-a]quinoxalines.

Isoindole derivatives are quite unstable, in fact, they rapidly darken and resinify at room temperature, especially in the presence of air.

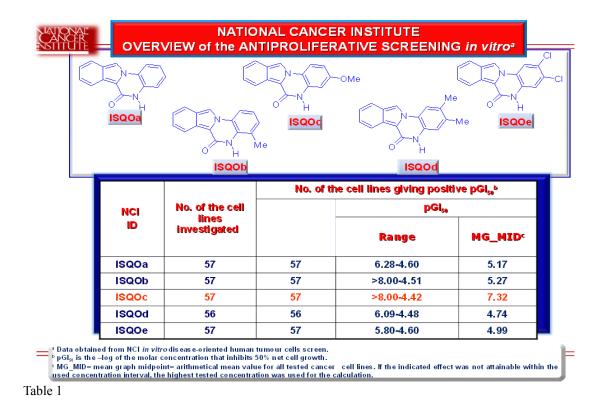


Therefore, isoindolo[1,2-a]quinoxalines. were synthesized from isoindole intermediates which would have strategic substituents: to both stabilize and functionalize the isoindole structure. Were synthesized compounds having an electron-withdrawing substituent at C-1, which stabilize the isoindole structure, high π -electron density at the position 3, also a phenyl substituted moiety on the nitrogen bearing a versatile amine group.



Isoindole[2,1-a]quinoxalines **ISQO** and **ISQ** have been synthesized and all derivatives were evaluated by the National Cancer Institute (NCI, Bethesda, MD) in the 3-cell line (MCF7-breast, NCI-H460-nonsmall cell lung and SF-268-CNS) one dose primary anticancer assay (10^{-4} M) (Table 1) [08JMC2387].

All five isoindoloquinoxalines **ISQO** were selected for evaluation against the full NCI panel of approximately 60 human cancer cell lines and were determined the following parameters: GI₅₀,TGI and LC₅₀. An evaluation of the data reported in the table 1 point out that all five derivatives exhibited antineoplastic activity against all the human cell lines. The most active compound was the 3-methoxy derivative **ISQOc** for which the pGI₅₀ mean value was 7.32. From the data reported, it was supposed that substituents that enriched the π -electron density of the benzene moiety of the quinoxaline system improved the antineoplastic activity. Derivatives **ISQO**, with the exception of **ISQOe**, were particularly effective against leukaemia subpanels; in fact the calculated pGI₅₀ mean value is higher than that of the overall cell lines.



The mean graph of the most active compound (table 2), shows that 60% of the treated cell lines are sensitive to compound **ISQOc** at nanomolar concentration. The most sensitive subpanels are leukaemia and colon cancer, for which the average value of pGI_{50} is 8.00.

Interestingly, also the TGI of some cell lines reaches the nanomolar level and in some cell lines the LC_{50} was measured in the low micromolar range.

The antiproliferative effects of isoindolo-quinoxaline derivatives were also evaluated in three human cancer cell lines overexpressing P-glicoprotein drug efflux and showed to be equally resistant to vinblastine, doxorubicin and sublines resistant to different drugs.

In order to clarify the mechanism(s) of action of isoindoloquinoxaline, the effects of different concentration of the most active compound **ISQOc** on cell cycle progression of Jurkat cells ($pGI_{50} = 8.39$) after 24 h of drug exposure were studied.

| Panel/Cel Line | Log ₁₀ GI60 | Log ₁₀ TGI | Log ₁₀ LC50 |
|---|---|--|--|
| Leukemia CCRF-CEM HL-60(TB) K-562 MOLT-4 RPMI-8226 SR Non-Smal Cell Lung Cancer A549/ATCC | < -8.00 < -8.00 < -8.00 < -8.00 < -8.00 < -8.00 < -8.00 | -6.90 -4.70 -4.80 -5.47 -6.47 -5.69 -5.77 | > 4.00 4.12 4.12 4.16 > 4.00 4.12 |
| EKVX H0IP-82 NCI-H226 NCI-H23 NCI-H32M NCI-H320 NCI-H322 Colon Cancer | -8.64 | 4.53 4.62 4.34 3.4,00 5.57 3.4,00 4. | > 4.00 4.17 > 4.00 > 4.00 > 4.00 > 4.00 > 4.00 |
| COLO 205 HCC-2998 HCT-116 HCT-15 HT29 KM12 | -5.50 < -8.00 < -8.00 -7.50 -8.71 < -8.00 | > -4.00 -5.92 -4.21 > -4.00 | > 4.00 -5.33 -5.38 > 4.00 > 4.00 |
| SW-620 CNS Cancer SF-268 SF-295 SF-539 | < -8.00 < -8.00 < -8.00 | -5.77 | -523 -4.06 > 4.00 |
| SNB-19 SNB-75 U251 Melanoma | < 8.00 -5.37 -5.60 < 8.00 | -7.39 -4.29 -5.13 -4.84 | -5.39 > 4.00 > 4.00 -4.28 |
| LOX INVI MALME-3M M14 SK-MEL-2 SK-MEL-28 SK-MEL-5 UACC-257 UACC-62 | < -8.00 < -8.00 < -8.00 -5.91 < -8.00 -4.80 -6.57 | -5.51 -4.83 -5.46 -4.57 -5.99 -4.51 -4.77 | 4.70 > 4.00 -5.44 4.66 > 4.00 -5.44 -5.44 -1.13 -4.13 -4.33 |
| Ovarian Cancer IGROV1 OVCAR-3 OVCAR-4 OVCAR-5 OVCAR-8 SK-0V-3 Renal Cancer | -7.88 -8.00 -5.43 < -8.00 < -8.00 < -8.00 < -5.29 | 4.87 < -8.00 > -4.00 -5.71 -5.07 > -4.00 | -4.06 > -4.00 -4.11 > -4.00 > 4.00 > 4.00 |
| 786-0 A438 ACHN CAKI-1 RXF 393 SN12C TK-10 | < -8.00 -6.85 < -8.00 < -8.00 - 6.55 < -8.00 - 6.55 | -5.75 4.78 4.73 4.50 -5.57 -5.05 -5.05 -5.60 | -524 -4.34 -4.26 > -4.00 > -4.00 > -4.00 |
| UO-31 Prostate Cancer PC-3 DU-145 Breast Cancer | < -8.00 | -524 -6.14 -4.94 | -4.55 > -4.00 -4.36 |
| MCF7 NCI/ADR-RES MDA-MB-231/ATCC HS 578T MDA-MB-435 T-47D | -6.85 < -8.00 -6.68 < -8.00 < -8.00 -4.42 | -4.80 < -8.00 -6.10 -4.97 > -4.00 | > 400 > 400 > 400 > 400 > 400 > 400 > 400 > 400 |
| MG MID Deta Range | | -529 271 4.0 +3 +2 +1 0 -1 -2 -3 | 427 1.17 1.44 +3 +2 +1 0 -1 -2 -3 |

Table 2 Mean graph of compound **ISQOc**

Treatment of Jurkat cells with **ISQOc** at the concentration of 500 nM led to profound changes of the cell cycle profile (fig.1).

Untreated cells showed a classical pattern of proliferating cells proportionally distributed in G1 (48%), S (35%) and G2/M (16%) phases.

On the contrary, a clear G2/M arrest pattern was observed with a concomitant decrease of G1 phase.

It was also interesting to note the appearance of a hypodiploid peak (sub-G1) indicative of apoptosis, which reached the value of 35% at the concentration of 62.5 nM.

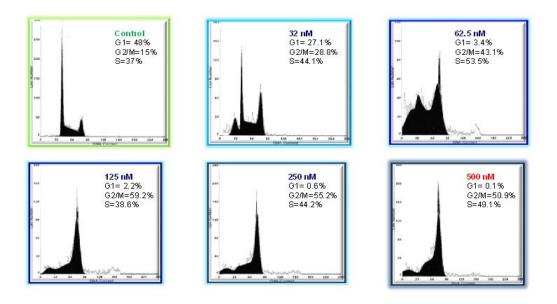


Fig.1

Compound **NSC 734237** (**ISQOc**) was also checked by immunofluorescence microscopy to study possible interactions with microtubules.

For these experiments was used a tumor cell line of small lung carcinoma (A-549) which was previously demonstrated to be sensible to these compounds, the microtubule network in control cells exhibited normal organization and arrangement.

On the contrary, compound NSC 734237 (ISQOc), at different concentrations (5 μ M and 2.5 μ M) disrupted the tubulin network.

Cells showed an evident characteristic "rounded up" morphology caused by disaggregation of microtubule in both interphase and mitotic phases after 24 h of

treatment.

These effects resemble that of vinblastine chosen as reference compound.

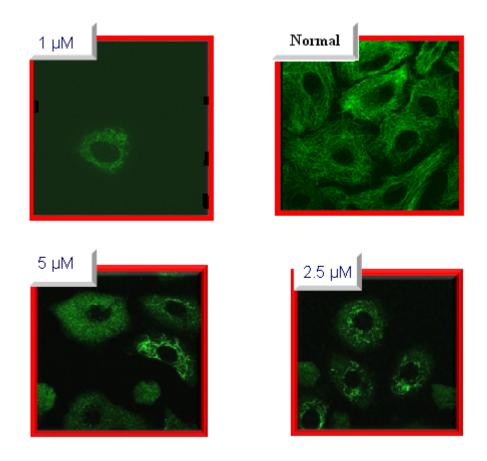


Fig.2

Immunofluorescence microscopy showed that **ISQOc** after 24 h of incubation completely disrupted the tubulin network. Polymerization was followed by fluorescent enhacement because of incorporation of a fluorescent reported into the microtubules as polymerization occurred. **ISQOc** investigated clearly induce a concentration dependent inhibition of tubulin polymerization (fig.2). In particular a complete inhibition of microtubule assembly was observed for **ISQOc** at a concentration similar to that of colchicine

By using DNA relaxation assay, it was determined that isoindoloquinoxalines inhibited topoisomerase I catalytic activity but not topoisomerase II activity.

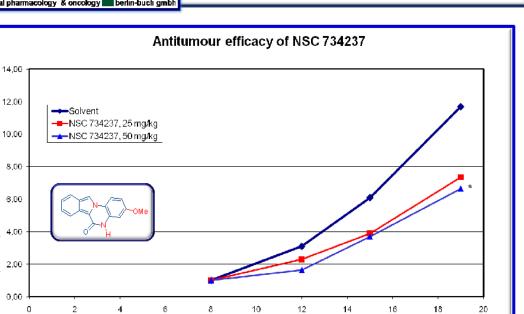
Further studies showed that **ISQOc** is a potent inducer of apoptosis in Jurkat cell line. The induction of apoptosis is associated with dissipation of the mitochondrial transmembrane potential, production of reactive oxygen species and cardiolipin oxidation and activation of caspase-3 and caspase-9.

Due to these really interesting data the Drug Discover Committee of the European Organization of Research and Treatment of Cancer decided to start the in vivo tests on compound **ISQOc**.

The influence of **ISQOc** on ADR-resistant MT-3 breast tumors was also investigate.

For this study the ADR-resistant, human breast carcinoma MT-3 grown as xenograft in nude mice was used. Mice (eight) bearing palpable MT-3/ADR tumors were treated 5-times weekly, for 2 weeks intra peritoneal with 25 or 50 mg/kg of ISQ3 or with the solvent (DMSO). Results showed that compound **ISQOc** induced a dose-dependent and significant tumor growth inhibition accompanied by a moderate and dose-dependent body weight reduction (fig.3).





Days after tumor transplantation

Fig.3

Median) Relative tumor volume

The expression of CD31 and Ki-67 was also investigated by immunohistochemistry. **ISQOc** showed significant anti-angiogenic effects in MT-3/ADR xenografts. Except from micro vessel number a significant reduction of micro vessel ratio and size was

1

even found for decreased in tumor treated with 50 mg/kg **ISQOc**. The expression of the proliferation marker Ki-67 was also significantly diminished due to treatment with **ISQOc**. This effect was not dose-dependent and found for both doses (fig.4). These findings are concordant with the observed anti-tumoral efficacy of **ISQOc** in MT-3/ADR xenografts.

Isoindole quinoxalines represent potential compounds for the therapy of cancer (probably including such tumours with acquired resistance to conventional cytostatics) with anti-angiogenic and anti-proliferative activity even at low doses and after a relatively short treatment period.

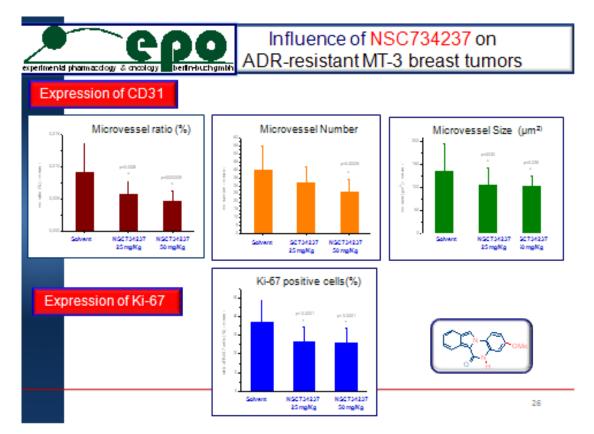


Fig.4

Molecular modeling

Considering the interesting results obtained for **ISQOa-e**, preliminary molecular modeling studies were performed in order to investigate the ability of new quinoxalines to bind Topoisomerase 1 and α , β tubulin, the two principal targets of **ISQO** compounds. To build the modeling, docking studies on literature compounds similar to **ISQO**, with antitubuline activity and with topoisomease 1 inhibition, were performed and the model was applied on new synthesized compounds.

1.Tubulin model

Colchicine is a tubulin assembly inhibitor that interacts with α , β tubulin dimers at distinct interfaces between the two subunits. These interactions cause microtubule destabilization and subsequent cellular apoptosis.

In 2004, Ravelli et al. reported the structure of tubulin in complex with DAMAcolchicine and with the stathmin-like domain (SLD) of RB3 (PDB 1SA0), at 3.5 Å resolution [04NAT198]. In this structure the colchicine binding site is located at the α , β interface (Fig.5).

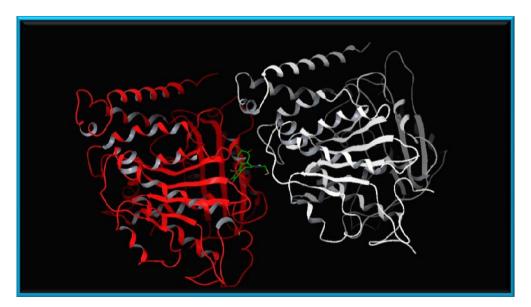


Fig.5

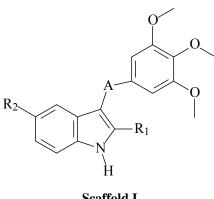
Strands S8 and S9, loop T7 and helices H7 and H8 of β-tubulin surround DAMAcolchicine, which also interacts with loop T5 of the neighbouring α –subunit. This structure was selected to dock new isoindole quinoxaline and ISQO compounds and it was useful in other works present in literature [08JMC4620], [05JMC6107], [09BMC6279].

a) Docking on literature compounds

To validate the model, structures with antitubulinic activity for which are reported IC_{50} (µM) for tubulin inhibition were docked [09JMC7512]. These structures are similar to **ISOO.** They have an indole portion linked to an aromatic portion with different substituents.

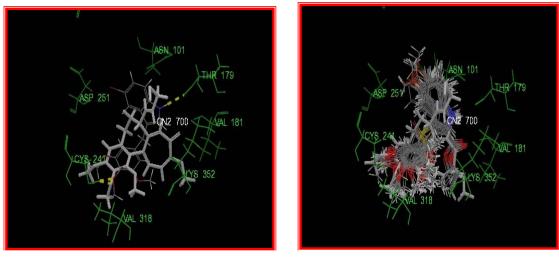
A number of 43 compounds with a molecular scaffold I, were docked using different conditions: using standard conditions, minimum sized Glide box, using different constraints for hydrogen bonds as VAL 181, ASN 101 and CYS 241.

The best docking was that one in which CYS 241 was used as constraint. The 77% of compounds bind the site through hydrogen bonds between indole NH and THF 179 residue and between -OMe groups and CYS 241 (Fig.6-7).



Scaffold I

The rest of other compounds bind the site orienting the indole portion in a different way. This is probably due to the free rotation around A atom (Fig.8).







Although most compounds bind the site in the same way and are good fitted, a correlation between IC_{50} and docking score was not found. This is probably due to a imperfect relationship between tubulin inhibition and inhibition of colchicine binding as data reported show [09JMC7512].

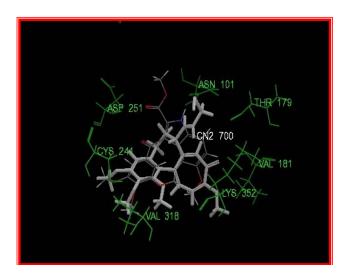
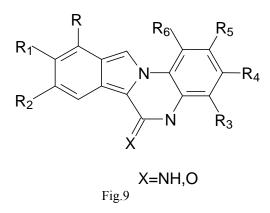


Fig.8

b) Docking on ISQ compounds

The same protein structure 1SA0 was used to dock quinoxaline compounds. Also in this case, structures were docked in different conditions: using standard conditions, minimum sized glide box, maximum sized glide box and using THF 179, VAL 181, ASN 101, and CYS 241 as constraints. A number of 28 compounds were docked. The best docking was that one in which VAL 181 was used as constraint. Compounds bind the active site depending on their structural features.



The docked compounds can be divided into 2 groups depending on their poses. The first group includes compounds with high glide score in which C=X group is involved in H-bonds with VAL 181 (Fig.10.1). Belong to this group compounds bearing the methoxyl group on the quinoxaline potion in different position from R_4 .

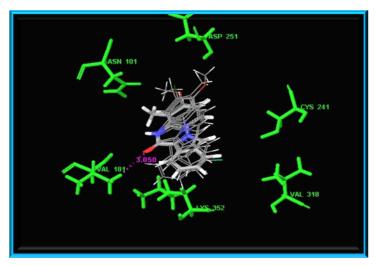


Fig.10

The second group includes compounds with medium-high glide score in which R_4 group is involved in H-bond with VAL 181 and R_1 is near to CYS 241 residue (Fig.11). Belong to this group compounds bearing methoxy or hydroxy group in R_4 ; Hydroxyl derivatives showed higher docking score values compared to the corresponding methoxyl derivatives

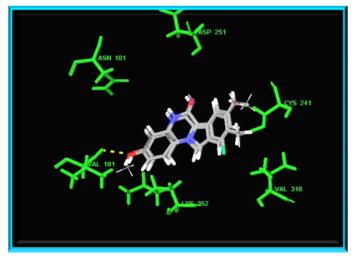


Fig.11

Only for some **ISQO** docked compounds, are known pGI_{50} values [08JMC2387]. For most of these compounds a correlation between glide score and pGI_{50} was found but not for all. This is due to pGI_{50} values that are related to both antitubuline and anti-Topoisomerase 1 activity.

According to the previous docking and to data reported in literature, compounds that bind Val 181 residue and interact in the same time with CYS 241 residue should be more active.

Comparing results, in both docking we can notice that the aromatic portion is often oriented to the CYS 241 residue and this feature could be responsible to confer better antitubuline activity.

| ISQ | Sustituents | Docking score |
|-------|--|------------------|
| 5m | X=O, R ₄ =R ₂ =Me,R=R ₅ =R ₆ =H R ₄ =OH | -7,616457 |
| 5q | X=O, R=F R ₁ =R ₂ =R ₃ =R ₆ =R ₆ =H R ₄ =OH | -7,530704 |
| 5p | X=O, R=R ₁ =R ₂ =R ₅ =R ₅ =H R ₂ =OMe R ₄ =OH | -7,518082 |
| 5g | X=0, R=R ₃ =R ₆ =R ₆ =H R ₁ =R ₂ =OMe R ₄ =OMe | -7,446021 |
| 4m | X=NH, R=R ₃ =R ₆ =R ₆ =H R ₁ =R ₂ =Me R ₄ =OH | -7,414586 |
| 4q | X=NH, R=F, R ₁ =R ₂ =R ₃ =R ₆ =R ₆ =H, R ₄ =OH | -7,311471 |
| 50 | X=0, R=R ₂ =R ₃ =R ₆ =R ₆ =H R ₁ =OMe R ₄ =OH | -7,169742 |
| 5k | X=O, R=R ₁ =R ₂ =R ₃ =R ₄ =R ₆ =H R ₅ =OMe | -7,142603 |
| 40 | X=NH, R=R ₂ =R ₃ =R ₆ =R ₆ =H R ₃ =OMe R ₄ =OH | -7,072346 |
| 5j | X=0, R=R ₁ =R ₂ =R ₅ =R ₅ =H R ₂ =OMe R ₄ =OMe | -7,043955 |
| 5n | X=0, R=R ₃ =R ₅ =R ₅ =H R ₁ =R ₂ =0Me R ₄ =0H | -7,035969 |
| 5f | X=O, R ₁ =R ₂ =Me,R=R ₃ =R ₆ =R ₆ =H R ₄ =OMe | -6,799753 |
| 5h | X=0, R=F, R ₁ =R ₂ =R ₃ =R ₆ =R ₆ =H R ₄ =OMe | -6,622998 |
| ISQOb | X=0, R=R ₁ =R ₂ =R ₃ =R ₆ =H R ₄ =R ₅ =Me | -6,254501 |
| 51 | X=0, R=R ₂ =R ₃ =R ₆ =R ₆ =H R ₁ =OMe R ₄ =OMe | -6,203328 |
| ISQOe | X=O, R=R ₁ =R ₂ =R ₃ =R ₈ =H R ₄ =R ₈ =CI | -6,014557 |
| ISQOc | X=0, R=R ₁ =R ₂ =R ₃ =R ₅ =R ₆ =H R ₄ =OMe | -5,887919 |
| 51 | X=0, R=R ₁ =R ₂ =R ₄ =R ₆ =H R ₃ =R ₆ =OMe | -5,815474 |
| ISQOd | X=0, R=R ₁ =R ₂ =R ₄ =R ₅ =R ₅ =H R ₃ =OMe | -5,803996 |
| ISQOa | X=0, R=R ₁ =R ₂ =R ₃ =R ₄ =R ₅ =R ₆ =H | -5,434076 |

Table 3

2.Topoisomerase 1 model

DNA-topoisomerase 1 is an important enzyme responsible to relax supercoiled DNA for transcription, replication, and mitosis [98SCI1504].

The X-ray ternary human Topoisomerase 1-DNA- Topotecan structure (fig.12) isolated by Staker and co-workers [02PNAS15387] shows that Topotecan intercalates at the site of DNA cleavage and bind the enzyme through H-bond with ASP533 residue.

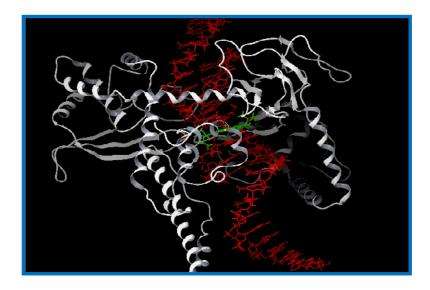


Fig.12

Because of structural analogy with **ISQO** compounds this X-ray structure with PDB code 1K4T was used for docking.

a) Docking on literature structures

To validate the model, structures taken from two different articles [08JMC4609], [06BMCL4395] (group A and B) and with anti-Topoisomerase 1 activity for which are reported qualitative data for topoisomerase 1 inhibition, were docked.

They are very similar to **ISQO** compounds. They contain an aromatic planar system in which a quinoxalinone portion is present as **ISQO** compounds.

The structures were docked in different conditions: using standard conditions, using compound protonated forms, neutral forms, using minimum and maximum sized glide box, using ARG 364 as constraint.

The best docking for group A was that one in which constraint ARG 364 and neutral ligands were used. A total number of 20 compounds were docked. Some molecules (12 of 20 compounds) respond to one way of binding in which pyridine N is oriented and interacts with H-bond with ARG 364. Compounds that no respond to this pose are the ones that have not electron pair available on this atom. Comparing structures with TTG docked structure we can see a very good superimposition (Fig.13).

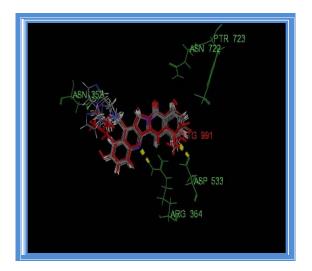


Fig.13

The best docking for group B was that one in which a minimum sized glide box and no ligand protonated forms were used. A total number of 17 compounds were docked. All molecules interact with ARG 364 through H-bonds but orienting the planar system in different ways.

10 of 17 compounds bind the active site interacting through H-bond between C=O group of five-membered ring and ARG 364 (Fig.14).

b) Docking on ISQO compounds

The same protein X-ray structure 1K4T was used for ISQO compounds docking.

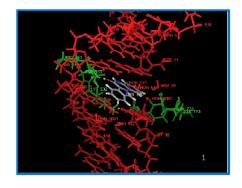
Also in this case all structures were docked using different conditions: using minimum and maximum sized Glide box, using standard condition and using ARG 364 as constraint.

The best docking was that where ARG 364 was used as constraint.

All compounds bind the active site in very similar ways interacting through C=X group with ARG 364. All molecules are in the same plane but additional interactions are responsible to orient molecules in different ways.

| ISQ | Substituents | Docking |
|------|--|-----------|
| | | score |
| 40 | X=NH, R=R ₂ =R ₃ =R ₅ =R ₆ =H R ₁ =OMe R ₄ =OH | -9,684861 |
| 50 | X=0, R=R ₂ =R ₃ =R ₅ =R ₆ =H R ₁ =OMe, R ₄ =OH | -9,627385 |
| 5n | X=0, R=R ₃ =R ₅ =R ₆ =H R ₁ =R ₂ =OMe, R ₄ =OH | -9,181279 |
| 5p | X=0, R=R ₁ =R ₃ =R ₅ =R ₆ =H R ₂ =OMe, R ₄ =OH | -9,142769 |
| 5q | X=0, R=F, R ₁ =R ₂ =R ₃ =R ₅ =R ₆ =H, R ₄ =OH | -9,001540 |
| 4q | X=NH, R=F, R ₁ =R ₂ =R ₃ =R ₅ =R ₆ =H, R ₄ =OH | -8,993757 |
| 5m | X=0, R ₁ =R ₂ =Me,R=R ₃ =R ₅ =R ₆ =H, R ₄ =OH | -8,910252 |
| 4m | $X=NH, R=R_3=R_5=R_6=HR_1=R_2=Me, R_4=OH$ | -8,876916 |
| SQOc | X=0, R=R ₁ =R ₂ =R ₃ =R ₅ =R ₆ =H, R ₄ =OMe | -8,799884 |
| 51 | X=0, R=R ₁ =R ₂ =R ₄ =R ₅ =H, R ₃ =R ₆ =OMe | -8,760821 |
| SQOb | X=0, R=R ₁ =R ₂ =R ₃ =R ₆ =H R ₄ =R ₅ =Me | -8,510735 |
| 5k | X=0, R=R ₁ =R ₂ =R ₃ =R ₄ =R ₆ =H R ₅ =OMe | -8,329139 |
| 5f | X=0, R ₁ =R ₂ =Me,R=R ₃ =R ₅ =R ₆ =H, R ₄ =OMe | -8,288526 |
| 5h | X=0, R=F, R ₁ =R ₂ =R ₃ =R ₅ =R ₆ =H, R ₄ =OMe | -8,232042 |
| 5j | X=0, R=R ₁ =R ₃ =R ₅ =R ₆ =H R ₂ =R ₄ =OMe | -8,212415 |
| SQOd | X=0, R=R ₁ =R ₂ =R ₄ =R ₅ =R ₆ =H, R ₃ =OMe | -8,155822 |
| SQOa | X=0, R=R ₁ =R ₂ =R ₃ =R ₄ =R ₅ =R ₆ =H | -8,078321 |
| SQOe | X=0, R=R ₁ =R ₂ =R ₃ =R ₆ =H R ₄ =R ₅ =Cl | -8,002569 |
| 5i | X=0, R=R ₂ =R ₃ =R ₅ =R ₆ =H R ₁ =R ₄ =OMe | -7,957684 |
| 5g | X=0 , R=R ₃ =R ₆ =R ₆ =H R ₁ =R ₂ =R ₄ =OMe | -6,261761 |







c) Comparing results

Comparing docking for ISQ compounds and literature compounds we can see how different structures are able to bind the active site in similar way interacting with ARG 364 residue. Also on this case compounds bearing hydroxyl group in para position had higher docking score value than the corresponding methoxyl derivatives.

AIM OF THE WORK

The aim of my work, considering the interesting results obtained by **ISQOc**, was the synthesis of new tetracyclic systems containing the pyrrole ring in order to study:

- 1 how the methoxy group position, on the quinoxaline moiety, can influence the anticancer activity;
- 2 how different substituents on the isoindole ring can influence the anticancer activity maintaining the methoxy group in *para* position;
- 3 how modifications on isoindole quinoxaline ring system can led to new more hydrophilic compounds reducing the use of DMSO in biological assays. In particular:

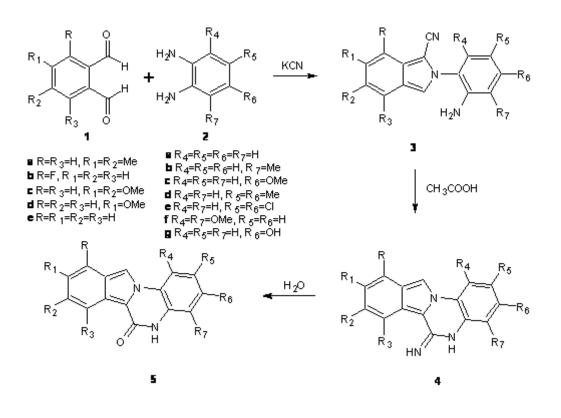
a) it was introduced an hydroxyl group in para position on the aromatic portion and it was studied how different substituents on isoindole ring can influence the anticancer activity;

b) it was introduced a N atom on the isoindole ring that could be protonated at physiological pH and it was studied how different substituents on aromatic portion can influence the anticancer activity;

c) it was introduced a N atom on the aromatic portion that could be protonated at physiological pH and it was studied how different substituents on isoindole ring can influence the anticancer activity;

d) were isolated the more hydrophilic imino forms of the corresponding quinoxalinones compounds synthesized.

SYNTHESIS



| Compounds | Subtituents |
|--------------|--|
| 3,4a | $R = R_1 = R_2 = R_3 = R_4 = R_5 = R_6 = R_7 = H;$ |
| 3,4b | $R = R_1 = R_2 = R_3 = R_4 = R_5 = H; R_7 = Me;$ |
| 3,4 c | $R = R_1 = R_2 = R_3 = R_4 = R_5 = R_7 = H; R_6 = OMe;$ |
| 3,4d | $R = R_1 = R_2 = R_3 = R_4 = R_7 = H; R_5 = R_6 = Me;$ |
| 3,4e | $R = R_1 = R_2 = R_3 = R_4 = R_7 = H; R_5 = R_6 = C1;$ |
| 3,4,5f | $R=R_3=R_4=R_5=R_7=H; R_1=R_2=Me; R_6=OMe;$ |
| 3,4,5g | $R=R_3=R_4=R_5=R_7=H; R_1=R_2=R_6=OMe;$ |
| 3,5h | $R_1 = R_2 = R_3 = R_4 = R_5 = R_7 = H; R = F; R_6 = OMe;$ |
| 3,5i | $R=R_2=R_3=R_4=R_5=R_7=H; R_1=R_6=OMe;$ |
| 3,5j | $R=R_1=R_3=R_4=R_5=R_7=H; R_2=R_6=OMe;$ |
| 3,4,5k | $R=R_1=R_2=R_3=R_4=R_7=H; R_5=OMe;$ |
| 3,4,51 | $R=R_1=R_2=R_3=R_5=R_6=H; R_4=R_7=OMe;$ |
| 3,4,5m | $R=R_3=R_4=R_5=R_7=H; R_1=R_2=Me; R_6=OH;$ |
| 3,4,5n | $R=R_3=R_4=R_5=R_7=H; R_1=R_2=OMe; R_6=OH;$ |
| 3,4,50 | $R=R_2=R_3=R_4=R_5=R_7=H; R_1=OMe; R_6=OH;$ |
| 3,5р | $R=R_1=R_3=R_4=R_5=R_7=H; R_2=OMe; R_6=OH;$ |
| 3,4,5q | $R_1 = R_2 = R_3 = R_4 = R_5 = R_7 = H; R = F; R_6 = OH;$ |

Scheme 1

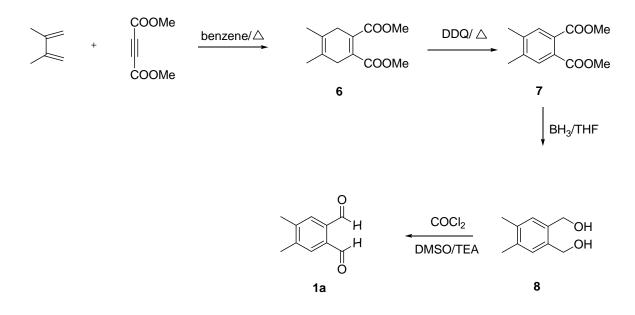
Considering the interesting results obtained in molecular docking studies, first it was planned to synthesize compounds in which the methoxyl group in para position in **ISQOc** compound was maintained in order to study how different substituents on isoindole portion can influence the anticancer activity and some compounds in which the methoxyl group position was changed.

It was also planned to synthesize new quinoxalines in which the methoxyl group in para position was replaced for an hydroxyl group in order to improve the solubility and to limitate the use of DMSO in biological screening. (Scheme 1)

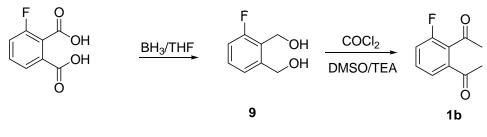
Isoindole quinoxalines **4** and **5** were synthesized from 2-(2-aminoaryl)1-cyanoisoindoles **3**. These intermediates were prepared by a Strecker type synthesis between different phtaloyldicarboxaldehydes **1** and substituted o-phenylendiamines **2** in the presence of potassium cyanide.

The not commercial available phthaloyldicarboxaldehydes were synthesized

Substituted fluoro-phthaloyldicarboxaldehyde and *ortho-* dimethylphthaloyldicarboxaldehyde were prepared by Swern oxidation of the corresponding dimethanols **8** and **9**. The reaction was carried out using oxalyl chloride activated by dimethyl sulfoxide and trietilamine as base in DCM at -78°C; the resulting phthaloyldicarboxaldehydes **1a** and **1b** were obtained in good yields (80-87%) [94S1035]. (Scheme 2-3)

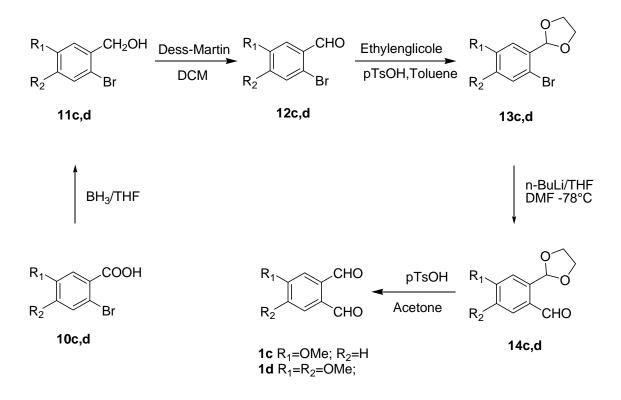


Derivatives **8** and **9** were prepared in good yields (80-99%) from the reduction either the corresponding dimethyl-ester **7** or the fluoro-carboxylic acid commercial available using BH₃ in THF solution. The dimethyl-ester **7** was previously obtained by a Diels-Alder reaction from dimethyl acetylenedicarboxylate and 2,3-dimethylbuta-1,3-diene in benzene or toluene under reflux to obtain compound **6** in excellent yield (92%); it was then oxidized using DDQ in chlorobenzene (2,3—dichloro-5,6-dicyano-benzo-1,4quinone). (Scheme 2-3)



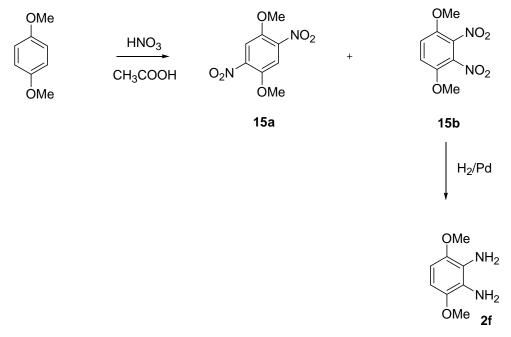
Scheme 3

Mono- and di-methoxy phthaloylcarboxaldehydes **1c** and **1d** were obtained in excellent yields (90-99%) from the suitable bromo-benzoic acids **10**. They were reduced using BH₃ solution in THF at O°C to the corresponding methanol derivatives **11c**. Subsequent Dess-Martin oxidation furnished the phthaloylcarboxaldehydes **12** in very good yields (90-98%). Aldehydes **12** were protected with ethylene glycol using Dean-Stark trap and compounds **13** were obtained in good yields (90%). Compounds **13** were formylated to compounds **14** using nBuLi in THF at -78°C and DMF and then deprotected at rt in acetone using p-TsOH to afford the corresponding phthaloylcarboxaldehydes **1c** and **1d**. (Scheme 4)



Scheme 4

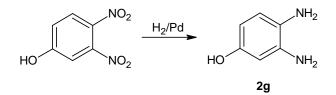
All phthaloylcarboxaldehydes, including the not substituted commercial available, were reacted with different phenilendiamine (**2a-g**); for the synthesis of 3,6-dimethoxy-1,2-phenylendiamine **2f**, the 1,4-dimethoxy-benzene was treated with acetic anhydride and nitric acid 70% at -15°C for few minutes.



Scheme 5

The resulting yellow precipitate, a mixture of the two isomers **15a-b**, was filtered off using acetic acid and purified in column. Isolated compound **15b**, it was reduced in methanol with Pd to the diamine **2f** . (Scheme 5)

The 3,4-diamino phenol 2g was obtained from reduction of the corresponding dinitro compound using H₂/Pd and in methanol. (Scheme 6)

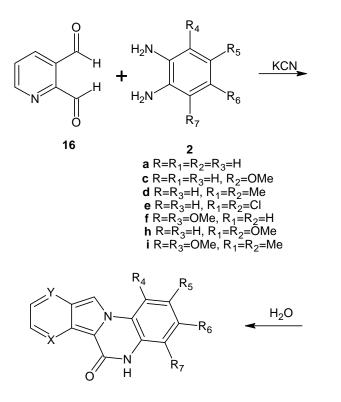


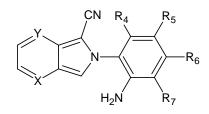
Scheme 6

All the key intermediates were cyclized in acetic acid to give new isoindoloquinoxaline derivatives in good yields (**4a-q** and **5f-q**). (Scheme 1)

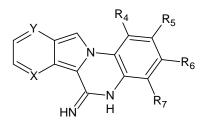
Then we planned to synthesize new tetracyclic systems introducing a nitrogen atom that protonated at physiological pH could improve quinoxalines solubility.

Were synthesized compounds **18a-m** and **19a** starting from a reaction between pyridine dicarboxy aldehyde **16** and different phenylendiamines **2** in the presence of potassium cyanide. In almost all cases a mixture of the two cyano intermediate isomers was obtained due to pyridine aldehyde asymmetry.





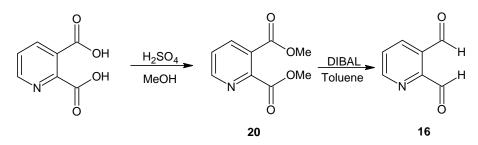




| Compounds | Substituents |
|-----------|--|
| 19a | R ₄ =R ₇ =H; R ₅ =R ₆ =Me; Y=CH; X=N |
| 19b | R ₄ =R ₇ =H; R ₅ =R ₆ =Me; Y=N; X=CH |
| 19c | R ₄ =R ₇ =H; R ₅ =R ₆ =Cl; Y=CH; X=N |
| 19d | R ₄ =R ₇ =H; R ₅ =R ₆ =Cl; Y=N; X=CH |
| 19e | R ₄ =R ₇ =OMe; R ₅ =R ₆ =H; Y=CH; X=N |
| 19f | R ₄ =R ₇ =OMe; R ₅ =R ₆ =H; Y=N; X=CH |
| 19g | $R_4=R_7=H; R_5=R_6=OMe; Y=CH; X=N$ |
| 19h | R ₄ =R ₇ =H; R ₅ =R ₆ =OMe; Y=N; X=CH |
| 19i | R ₄ =R ₇ =OMe; R ₅ =R ₆ =Me; Y=CH; X=N |
| 19j | R ₄ =R ₇ =OMe; R ₅ =R ₆ =Me; Y=N; X=CH |
| 19k | R ₄ =R ₅ =R ₆ =R ₇ =H; Y=CH; X=N |
| 191 | R ₄ =R ₅ =R ₆ =R ₇ =H; Y=N; X=CH |
| 19m | R ₄ =R ₅ =R ₇ =H; R ₆ =OMe; Y=N; X=CH |
| 18a | R ₄ =R ₅ =R ₇ =H; R ₆ =OMe; Y=N; X=CH |

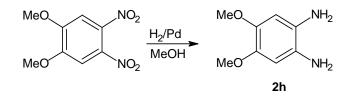
Scheme 7

Pyridine-2,3-dicarbaldehyde **16** was prepared according to literature [68BSCF4117] from the corresponding carboxylic acid that was dissolved in methanol and a catalytic amount of sulfuric acid to led to compound **20**. Compound **20** was then reduced to dicarbaldehyde **16** using DIBAL in toluene at -78°C. (Scheme 8)



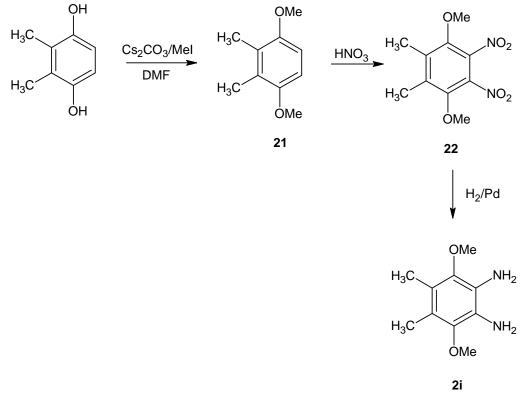
Scheme 8

Pyridine-2,3-dicarbaldehyde was reacted with different phenylendiamines (**2a**, **2c-f**, **2hi**). For the synthesis of 4,5-dimethoxy-o-phenylendiamine **2h**, not commercial available, the corresponding dinitro compound was reduced in methanol using H₂/Pd. (Scheme 9)



Scheme 9

3,6-dimethoxy-4,5-dimethyl-phenylendiamine **2i** was synthesized from 2,3dimethylbenzene-1,4-diolo that was methylated in DMF using Cs_2CO_3 as base in good yield (70%). Compound **21** was treated with nitric acid 70% sol. at -15°C and the resulting compound **22**, obtained in 80% yield, reduced in methanol to the diamine **2i** using H₂/Pd. (Scheme 10)

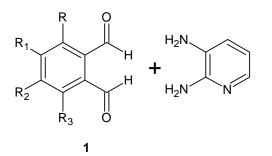


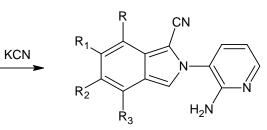
Scheme 10

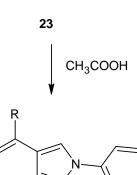
All the key intermediates **17** were cyclized in acetic acid to give new azaisoindoloquinoxaline derivatives (**18a-m** and **19a**) in good yields. (Scheme 7)

Then we also try to introduce a nitrogen atom on the aromatic portion to obtain the new pyrido-pyrazino-isoindole system of type **24** and **25**.

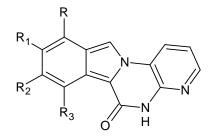
In this case different aldehydes **1** were reacted with pyridine-2,3-diamine to afford the intermediates **23** that were cyclized in acetic acid using a catalytic amount of DMF to led compound **24a-c** and **25a-c** in good yields. (Scheme 11)

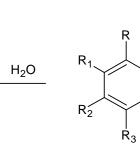






-N H





25

24

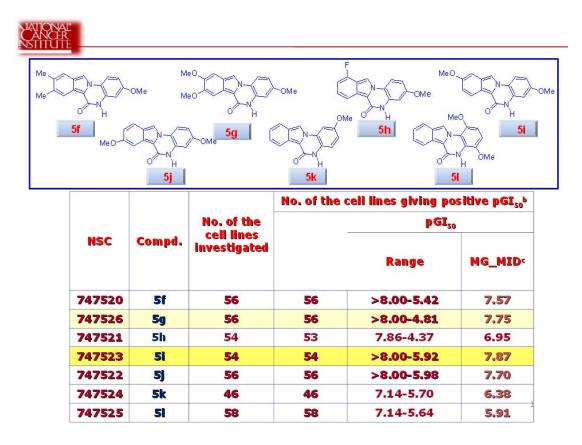
ΗŃ

| Compounds | Substituents |
|-----------|-------------------------|
| 25a | $R=R_1=R_2=R_3=H;$ |
| 24a | $R=R_1=R_2=R_3=H;$ |
| 25b | $R=R_3=H; R_1=R_2=Me;$ |
| 24b | $R=R_3=H; R_1=R_2=Me;$ |
| 25c | $R=R_3=H; R_1=R_2=OMe;$ |
| 24c | $R=R_3=H; R_1=R_2=OMe;$ |

Scheme 11

BIOLOGICAL DATA

Compound **5f-1** were submitted to the National Cancer Institute of Bethesda (NCI) and tested in a full panel of about 60 different tumour cell lines.





All the new derivatives resulted to be more potent than the lead compound **ISQOc**.

They showed potent antitumor activity against the total number of cell lines investigated with pGI_{50} mean values from 5.91 to 7.87.

The most active derivative was compound 5i, whose pGI₅₀ mean value was 7.87.

Is interesting to note that the two derivatives **5k** and **5l**, in which the methoxy group position was changed, are the less active although they showed pGI_{50} of 6.38 and 5.91 respectively (table 5).

In fig.15 is reported the mean graph of the most active compound **5i** (NSC 747523) whose GI_{50} value reached nanomolar level on 89% of the cell lines and 20% of the cell lines at TGI.

| National Cancer Institute Developmental Therapeutics Program | | NSC :747523/1 | Units :Molar | SSPL :0GOB | EXP. ID :0807RS68 |
|--|--|---|--------------|--|-------------------|
| Mean Graphs | | Report Date :March 07, 2009 | | Test Date :July 21, 2008 | |
| Panel/Cell Line | Log ₁₀ GI50 GI50 | Log ₁₀ TGI TGI Log | | Log ₁₀ LC50 LC50 | |
| Leukemia CCRF-CEM HL-60(TB) K-562 MOLT-4 SR Non-Small Cell Lung Cancer A549/ATCC EKVX HOP-62 HOP-92 NCI-H23 NCI-H23 NCI-H23 NCI-H23 NCI-H23 NCI-H460 NCI-H522 Colon Cancer COLO 205 HCC-2998 HCT-116 HCT-15 HT29 KM12 SW-620 CNS Cancer SF-286 SF-285 S | < -8.00 | -6.00 -6.00 -6.98 -6.98 -5.81 -5.83 -5.83 -5.83 -5.83 -5.83 -5.83 -5.83 -5.83 -5.83 -5.84 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.98 -5.99 -5.90 -5.87 -5.87 -5.87 -5.87 -5.87 -5.87 -5.87 -5.88 -5.63 -5.89 -5.89 -5.80 -5.81 -5.77 -5.82 -7.71 -5.85 </td <td></td> <td>$\begin{array}{cccccccccccccccccccccccccccccccccccc$</td> <td></td> | | $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | |
| MID Delta Range | -7.87 0.13 2.08 +3 +2 +1 0 -1 -2 -3 | -6.3 1.7 3.57 +3 +2 +1 | 0 -1 -2 -3 | -5.31 2.69 4.0 +3 +2 +1 0 | |

Fig.15

| National Cancer Institute Dev | elopmental Therapeutics Program | NSC :747526/1 | Units :Molar | SSPL :0GOB | EXP. ID :0807RS68 |
|--|--|---|--------------|--|-------------------|
| Mean Graphs | | Report Date :May 13, 2011 | | Test Date :July 21, 2008 | |
| Panel/Cell Line | Log ₁₀ GI50 GI50 | Log ₁₀ Tai Tai | | Log ₁₀ LC50 LC50 | |
| Leukemia CCRF-CEM HL-60(TB) K-562 MOLT-4 RPMI-8226 SR Nor-6mail Cel Lung Cancer A549iATCC EKVX HOP-62 HOP-52 HOP-52 HOP-52 HOP-52 NCI+4226 NCI+4256 NCI+4256 NCI+4256 NCI+425 Colon Cancer COLO 205 HCT-15 HCT- | cb ₁₀ 000 cb0 < +8.00 | 100 5.91 4 5.89 5.89 4.800 5.84 5.84 4.75 4.75 4.75 4.75 4.75 4.75 4.75 4.75 4.75 4.75 4.75 4.75 4.75 4.87 4.82 5.98 7.11 5.91 7.12 5.79 7.85 4.63 5.85 4.63 5.85 | | 5.18 5.23 5.30 5.11 5.34 4.30 4.32 4.32 4.33 4.32 4.33 4.33 4.40 5.37 4.13 4.23 4.40 5.37 4.13 4.23 4.23 4.23 4.35 5.34 5.34 5.34 5.34 5.34 5.35 5.34 5.34 5.34 5.34 5.35 5.34 5.34 5.35 5.34 5.35 5.34 5.35 5.34 5.35 5.34 5.35 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.34 5.35 5.35 5.34 5.35 5.34 5.35 5.35 5.34 5.35 5.35 5.34 5.35 5.34 5.35 5.34 5.35 5.34 5.35 5.34 5.35 5.34 5.35 5.34 5.35 5.34 5.34 5.35 5.34 5.35 5.34 5.34 5.35 5.34 5.34 5.35 5.34 5.34 5.35 5.34 5.34 5.34 5.35 5.34 5.35 5.34 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.35 5.34 5.35 5.35 5.35 5.35 5.35 5.35 5.35 5.34 5.35 | |
| MALME-3M MI4 MDA-MB-435 SK-MEL-2 SK-MEL-3 UACC-357 UACC-52 Ovarian Cancer IGROV1 OVCAR-3 OVCAR-3 OVCAR-5 OVCAR-6 NCIADR-RES | < -8.00 < -8.00 < -8.00 -7.50 < -8.00 -7.50 < -8.00 -7.32 -7.52 < -8.00 < -8.00 < -8.00 < -8.00 < -8.00 | 4.88 4.43 4.58 4.95 4.53 4.57 4.57 5.86 4.80 4.80 4.80 4.80 4.80 4.80 4.80 4.80 | | 525 4.02 4.32 4.19 4.06 4.23 4.33 4.23 4.23 4.22 5.36 5.81 4.31 4.37 4.39 7.32 4.32 | |
| 8K-0V-3 Renal Cancer 785-0 A438 ACHN CAKI-1 RXF 393 8N12C TK-10 UO-31 Frostate Cancer PO-3 DU-145 Breast Cancer | -7.29 < -8.00 < -8.00 < -8.00 < -8.00 -7.88 -7.38 -7.38 < -8.00 < -8.00 < -8.00 < -8.00 < -8.00 | 4.58 4.79 6.96 4.78 < 8.00 4.75 4.86 4.72 5.71 7.57 | | 4.33 4.52 4.39 4.64 5.42 4.37 4.39 4.34 4.39 4.34 4.39 4.34 | |
| MGF7 MDA-MB-231/ATCC H0 5751 BT-549 T-47D | -7.51 -7.51 < 8.00 < 8.00 -4.81 | -5.88 -7.27 -7.45 -4.45 | | 5.32 4.47 4.40 4.48 4.10 | |
| _MID Delta Range | -7.75 0.25 3.19 +3 +2 +1 0 -1 -2 -3 | -5.74 2.26 3.57 +3 +2 +1 (|) | -4.75 2.57 3.32 +8 +2 +1 0 | 1 2 3 |

Fig.16

In fig.16 is also reported the mean graph of compound **5g** (NSC 740526) that was selected for in vivo test. For the pharmacologist infact the two compounds are very similar but **5g** was synthesized in better yield and, for this reason, selected for in vivo tests.

In vivo test performed on Ovcar-3 tumor showed that the compound at the dose of 15 mg/kg induced a significant reduction of the tumor although accompanied by a decrease of the body weight which was however recovered (fig.17)

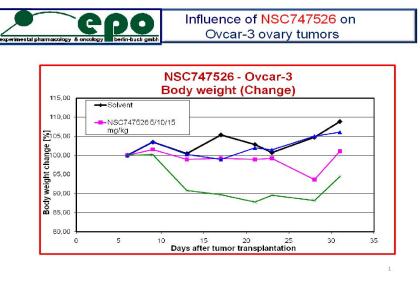


Fig.17

Also for this compound the expression of CD31 and Ki-67 was investigated by immunohystochemistry (fig.18).

Compound **5i** showed significant anti-angiogenic effects in Ovcar-3 tumors. It was observed a dose dependent significant reduction of the micro vessel number, micro vessel ratio and size.

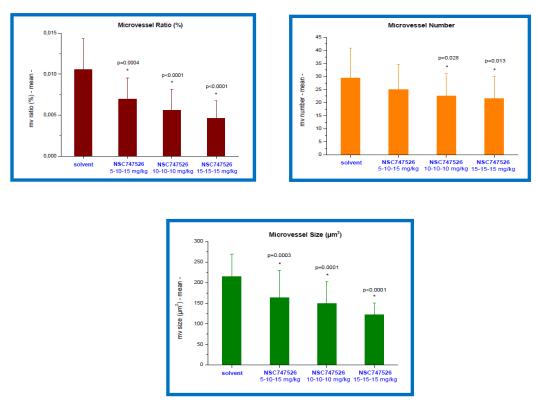


Fig.18

The expression of the proliferation marker Ki-67 was also significantly diminished and in this case was dose-dependent (fig.19)

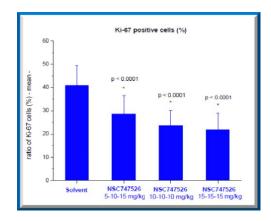
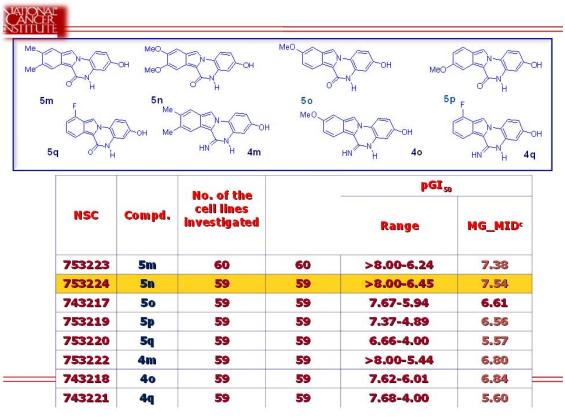


Fig.19

Also compounds **5m-q** and **4m**, **4o** and **4q** were selected by the NCI for the total screening on the full tumour cell lines panel.



| Table (| 2 |
|---------|---|
|---------|---|

All compounds showed antiproliferative activity on all tested cell lines from micromolar to nanomolar concentrations. The most active compound was 5n whose pGI₅₀ mean value was 7.54 (table 6). The imino forms 4m, 4q and 4o showed antiproliferative activity from micromolar to nanomolar concentrations.

Also the imino **4a-g**, **4k-l** and **4n** were isolated and selected for the one-dose prescreening by the NCI and the most interesting compounds **4g** and **4n** were selected for the total-screening at five dose concentrations.

For compound **4g** (NSC 760286) was calculated a pGI_{50} mean value of 7.44 and its mean graph showed it is very active on colon cancer cell line that are sensitive even at nanomolar concentration (fig.20).

| National Cancer Institute Developmental Therapeutics Program Mean Graphs | | | NSC :760286/1 | Units :Molar | | SSPL :0GOB | EXP. ID :1108NS03 |
|--|--|---------------|--|--------------|---|--|-------------------|
| | | | Report Date :November 24, 2011 | | | Test Date :August 15, 2011 | |
| Panel/Cell Line | Log ₁₀ G150 | G/50 | Log ₁₀ TGI | TGI | l | .og ₁₀ LC50 | LC50 |
| Leukemia CCRF-CEM HL-60(TB) K-562 MOLT-4 RPMI-8226 Non-Smail Cell Lung Cancer A549/ATCC EKVX HOP-62 | -7.99 -6.86 < -8.00 -6.60 -7.59 -7.55 -6.53 < -8.00 | | -4.96 -6.33 -4.92 -4.92 -4.97 -4.70 -4.72 -4.70 -4.73 | | | -4.18 > -4.00 | |
| ENVA HOP-62 HOP-92 NCH423 NCH423 NCH482 NCH460 NCH4522 Colon Cancer COLO 205 HCC-2398 | -4.84 < -8.00 < -8.00 -7.91 < -8.00 < -8.00 | | -4.72 -4.70 -4.73 -4.53 -4.54 -4.54 -4.64 -4.97 < -8.00 -6.86 | - | | -4.03 -4.36 -4.21 -4.21 -4.25 -4.44 -4.31 -4.35 -4.29 -4.78 | |
| HCC-2998 HCT-116 HCT-15 HT29 KM12 SW-620 CNS Cancer SF-286 SF-295 SF-295 SF-539 SNB-75 SNB-75 SNB-75 L1251 | < -8.00 -7.36 < -8.00 < -8.00 -7.85 < -8.00 -7.85 < -8.00 < -8.00 < -8.00 | | -6.86 -4.78 -4.86 -4.88 -4.97 -4.90 -4.70 -4.78 | | | -4.78 -4.35 -4.38 -4.34 -4.18 -4.30 -4.21 -4.21 -4.31 -4.16 | |
| SF-295 SF-539 SNB-19 SNB-75 U251 Melanoma LOX IMVI MALME-3M | < -8.00 -7.89 -7.76 < -8.00 -7.39 < -8.00 -5.03 < -8.00 | | -4.78 -4.89 -6.66 -4.74 -4.54 -4.80 -4.82 -4.42 | | | -4.62 -4.27 > -4.00 -4.31 | - |
| M14 MDA-MB-435 SK-MEL-2 SK-MEL-5 UACC-257 UACC-62 Overging Concer | < -8.00 -4.97 < -8.00 -7.83 < -8.00 | | < -8.00 -4.55 -4.68 -6.21 -4.55 -4.66 | - | | -4.13 -4.28 -4.12 -4.23 -5.06 -4.14 -4.10 | _ |
| OVCAR-3 OVCAR-3 OVCAR-5 OVCAR-5 OVCAR-5 SV-0V-3 Renal Cancer 785.0 | -7.01 < -8.00 -6.97 -7.13 -7.38 < -8.00 < -8.00 -7.21 | 1 | -4.76 -4.73 -4.73 -4.83 -6.49 | Ļ | | -4.03 -4.38 -4.27 -4.36 -4.31 -4.31 -4.37 -4.59 | |
| A498 ACHN CAKI-1 RXF 393 SN12C TK-10 UO-31 Prostate Cancer | -7.21 -7.87 < -8.00 < -8.00 < -8.00 -7.52 -5.74 | | -4.95 -6.23 -4.79 -4.70 -4.70 -4.78 -4.69 -4.74 | | | -4.44 -4.52 -4.39 -4.24 -4.27 -4.36 -4.32 -4.35 | |
| PC-3 DU-145 Breast Cancer | < -8.00 < -8.00 | = | -4.77 -4.98 | | | -4.33 -4.47 | • |
| MCF7 MDA-MB-231/ATCC HS 578T BT-549 T-47D MDA-MB-468 | -7.09 -6.48 -7.48 -6.34 -7.47 | = | -4.87 -4.64 -4.56 -4.74 -4.59 -4.90 | | | -4.19 -4.18 >> -4.00 -4.36 -4.23 -4.35 | |
| MID | -7.44 | | -5.08 | | | 4.29 | |
| Delta Range | 0.56 | +1 0 -1 -2 -3 | 2.92 3.58 | +1 0 -1 -2 | 3 | 0.77 1.06 | 0 -1 -2 -3 |

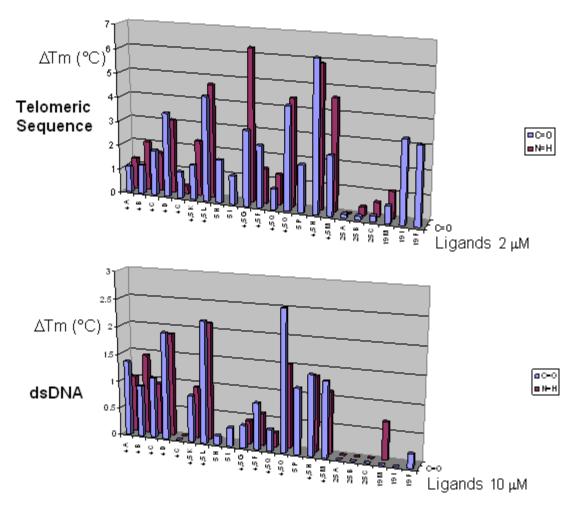
| National Cancer Institute Developmental Therapeutics Program Mean Graphs | | | NSC :760289/1 Units : Molar | | SSPL:0GOB | EXP. ID :1108NS03 |
|--|---|---------------|---|------------|--|----------------------------|
| | | | Report Date :November | 24, 2011 | Test Date : August 15, 2 | Test Date :August 15, 2011 |
| Pane/Cell Line | Log ₁₀ GI50 | GI50 | Log ₁₀ TGI | TGI | Log ₁₀ LC50 LC | 250 |
| Leukemia CCRF-CEM HL-60(TB) K-562 MOIT-4 RPMI-8226 Non-Smail Cell Lung Cancer A549/ATCC FK/X | -6.74 -6.55 -7.51 -6.29 -5.95 | | -5.12 -6.10 -5.29 -5.14 -5.01 | - | > -4.00 -4.07 > -4.00 -4.03 > -4.00 | |
| HOP-92 HOP-92 NCH+226 NCH+226 NCH+322M NCH4522 Colon Cancer COLO 205 HCC-2998 HCT-116 HCT-15 HT29 KM12 SW-620 | -6.27 -5.66 -6.48 -5.52 -7.42 -5.99 -5.73 -6.56 -7.79 | | -4.89 -4.84 -4.92 -4.82 -5.65 -5.51 -4.81 -5.64 | Ę | 4 22 4.33 4.46 4.35 5.15 5.03 4.37 4.42 4.55 | - |
| | -7.22 -6.53 -7.47 -6.57 -6.47 -6.21 -7.44 | | -5.36 -5.01 -5.76 -4.94 -5.41 -5.28 -4.80 | | -4.50 -4.49 -5.32 -4.40 -4.34 -4.47 -4.16 | _ |
| CNS Cancer SF-288 SF-285 SF-539 SNB-19 SNB-75 U251 Melanoma LOX IM/U | -7.37 -6.67 -7.17 -7.02 -7.53 -7.23 | | -4.81 -4.92 -5.96 -4.81 -4.98 -4.97 -5.78 | | -4.34 -4.29 -4.91 -4.34 -4.31 -4.48 | - |
| LOX IIM/I MALME-3M M14 MDA-MB-435 SK-MEL-2 SK-MEL-28 SK-MEL-28 SK-MEL-5 UACC-257 UACC-257 UACC-257 UACC-257 UACC-257 UACC-257 UACC-257 | -7.71 -6.40 -7.20 -7.79 -5.77 -7.88 -7.75 -5.88 -7.63 | | -5.78 -4.62 -5.16 -4.88 -5.10 -4.81 -6.79 -5.15 -5.86 | | 5.18 -4.04 -4.48 -4.23 -4.36 -4.39 -5.59 -4.49 -5.21 | |
| IGROV1 OVCAR-3 OVCAR-4 OVCAR-5 OVCAR-8 NCI/ADR-RES SK-OV-3 Renal Cancer | -6.30 -7.28 -5.77 -6.23 -5.78 -7.13 | | -4.80 -5.49 -4.87 -4.89 -4.82 -4.95 -4.93 | | -4.05 -4.88 -4.43 -4.43 > -4.00 -4.14 -4.46 | |
| 786-0 A 498 ACHN CAKI-1 RXF 393 SN12C TK-10 UO-31 Prostate Cancer | -6.39 -6.93 -7.07 -6.61 -7.89 -6.90 -5.94 -6.73 | ł | -5.60 -6.07 -5.62 -5.44 -4.92 -4.90 -5.61 | | -5.02 -5.34 -4.78 -4.37 -4.94 -4.38 -4.44 -4.74 | |
| PC-3 DU-145 Breast Cancer | -6.56 -6.89 | 1 | -4.81 -5.01 | | -4.31 -4.47 | |
| MCF7 MDA-MB-231/ATCC HS 578T BT-549 T-47D MDA-MB-468 | -6.92 -5.80 -6.45 -5.75 -6.45 -7.01 | | -5.79 -4.82 -4.55 -4.94 | | -5.21 -4.32 > -4.00 -4.46 -4.30 -4.88 | _ |
| | | | | | | |
| _MID Delta Range | -6.73 1.16 2.37 +3 +2 | +1 0 -1 -2 -3 | -5.2 1.59 2.24 | 0 -1 -2 -3 | -4.5 1.09 1.59 | |

Fig. 21

The mean graph of compound **4n** showed a pGI_{50} mean value of 6.73 and that CNS Cancer and melanoma cell line are very sensitive to this compound (fig.21).

For all compounds the possible interaction with nucleic acids was also studied. The thermal DNA denaturation was evaluated in presence of quinoxaline compounds at 2 μ M and 10 μ M concentration. Compounds were tested on human telomeric sequence (AGGGTTAGGGTTAGGGTTAGGGTTAGGGT), c-Myc promoter sequence (Pu24, TGAGGGTGGGGAGGGTGGGGGAAGG) both having a G-quadruplex arrangement, and a double helix random sequence fragment (GGATGTGAGTGTGAGTGTGAGGG).

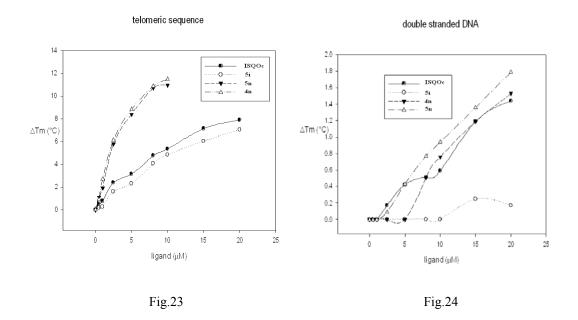
Imino and ketone forms of quinoxalines were compared and results showed that there is not significant difference between the two forms for the interation with G-quadruplex form or doble helix DNA.



For G-quadruplex interation is very important the presence of substituents in position 8 and 9 bearing an electron-negative atom. Infact compounds **4n** and **5n** showed the best results in terms of G-quadruplex interaction contrary to compound **4m** and **5m** that didn't show excellent resuls. Also the presence of hydroxyl group in para position on the quinoxaline portion seems to be more important than methoxyl group for G-quadruplex interation. Same results were obtained for Pu24 promoter recognition.

For compounds **4n**, **5n**, **5i** was evaluated the DNA denaturation relating to compound concentrations compairing the results with ISQOc.

All compounds are able to induce DNA denaturation in a dose dependent way (Fig.23-24).



Studies on circular dichroism to understand how quinoxalines are able to modify nucleic acid structures were also performed. Results showed that all compounds are able to increase the signal at 290 nm for the human telomeric sequence showing an effective interaction. Also in this case the best result was related to compound **4n**.

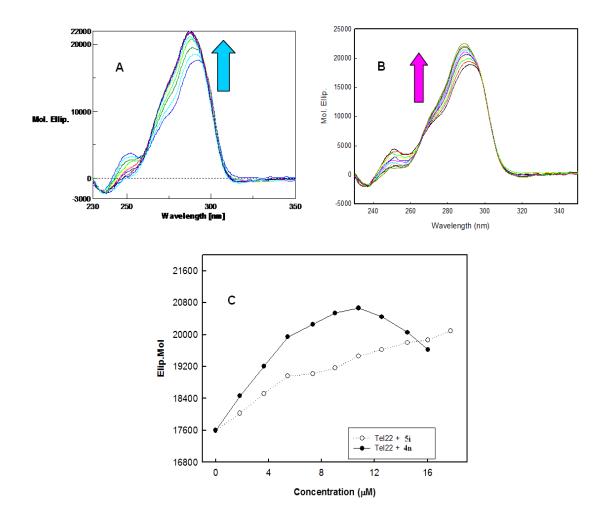


Fig.25. A) TeI22 (4 μ M) in presence of **4n** (0.16 μ M) in TK 10/50 pH 7.4; **B**) TeI22 (4 μ M) in presence of **5i** (0.16 μ M) in TK 10/50 pH 7.4; **C**) CD Titration of TeI22 at 292 nm

About interaction with double helix DNA, compound **4n** causes significant dichroism spectrum alteration in a similar way to Mitoxantrone showing better results than compound **5i**.

Same results were obtained for compound **5n**. Compound **5o** causes three-dimensional DNA structure alteration and compound **5i** seems to be no effective.

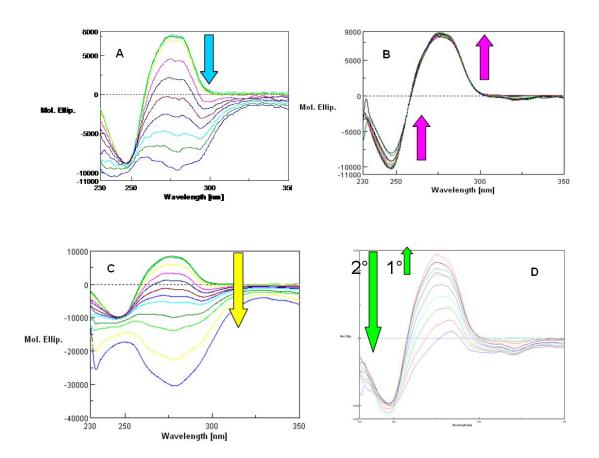


Fig.26. A) ctDNA (96 μ M) in presence of **4n** (0-30 μ M)in TK 10/50 pH 7.4; B) ctDNA (96 μ M) in presence of **5i** (0-30 μ M)in TK 10/50 pH 7.4; C) ctDNA (96 μ M) in presence of **5n** (0-30 μ M)in TK 10/50 pH 7.4; D) ctDNA (96 μ M) in presence of **5o** (0-30 μ M)in TK 10/50 pH 7.4;

Electrophoresis studies were also performed and showed that compound **50**, **5i**, **5n** and **4n** are able to interact with nucleic acids confirming dichroism studies results (Fig.27).

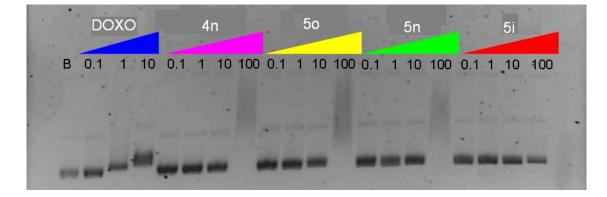
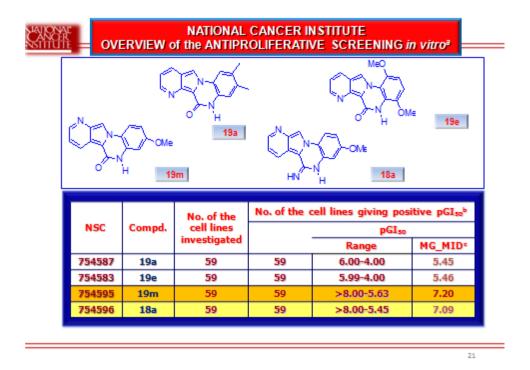


Fig.27





Compounds **18a-m** and **19a** were submitted to the NCI and selected for the prescreening. The most interesting derivatives were **19a**, **18a**, **19e** and **19m**. For all compounds was calculated the pGI_{50} mean value that was of 5.45, 7.09, 5.46 and 7.20 respectively. The most interesting compounds **19m** and **18a** were active at nanomolar concentration in more than 80% of tumor cell lines.

Also compounds **24a-c** and **25a-c** were selected by the NCI for the pre-screening. The most interesting compound was **25b** that was tested on the full panel and whose pGI_{50} mean value was 5.78.

X- Ray studies were also performed on compound 4c to know its exact mesomeric form.

Compound **4c** was crystallized in acetic acid and through its crystallographic structure bond distances were calculated and compared to analogue structures for which the same parameters were known.

Results showed that the double bound is probably located outside the quinoxaline ring system, having the shortest bound distance. (Fig.29)

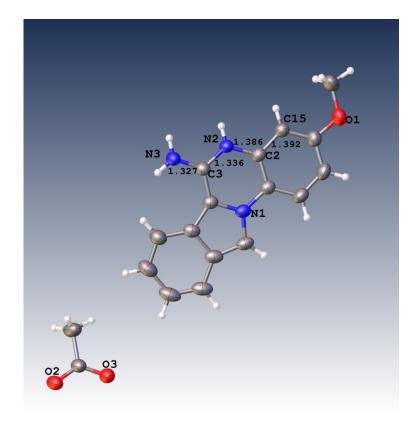


Fig.29

EXPERIMENTAL DATA

All melting points were taken on a Büchi-Tottoli apparatus and are uncorrected; IR spectra were determined in bromoform with a Jasco FT/IR 5300 spectrophotometer. 1H e 13C NMR spectra were measured at 200 and 50.3 MHz, respectively, in DMSO- d6 or CDCl₃ solution, using a Bruker AC 200 MHz spectrometer (TMS as internal reference). Chromatography column was performed with Merk silica gel 230-400 mesh ASTM or BIOTAGE FLASH chromatography or with Büchi Sepacore chromatography module (prepacked cartridge system). Elemental analyses (C, H, N) were within $\pm 0.4\%$ of the theoretical values. Microwave experiments were carried out using a CEM Discover LabmateTM microwave apparatus.

Synthesis of dimethyl 4,5-dimethylciclohexa-1,4-diene-1,2-dicarboxilate 6

To a solution of dimethyl acetilene dicarboxilate (25g, 0.17 mol) in toluene (100 ml) 2,3-dimethylbuta-1,3-diene (13.1g, 0.16 mol) was added under nitrogen atmosphere. The mixture was stirred under reflux overnight. The solvent was evaporated under reduced pressure.

m.p. 64.0-65.0°C; Yield 95%; IR 1732 (CO) cm⁻¹; ¹H NMR CDCl₃ (ppm): 1.69 (s, 3H, CH₃), 3.77 (s, 2H, CH₂), 3.84 (s, 3H, CH₃).

Synthesis of 4,5-dimethylbenzene-1,2-dicarboxilate 7

To a solution of compound 6 (30g, 0.13 mol) in chlorobenzene (500 ml) 2 equivalents of DDQ were added. The mixture was stirred under reflux for 24h. After cooling, diethyl ether was added and the mixture was filtered on celite. Evaporated the solvent under reduced pressure, the crude was purified by chromatography using DCM as eluent.

m.p. 49.0-50.0°C; Yield 92%; IR 1728 (CO) cm⁻¹; ¹H NMR CDCl₃ (ppm): 2.29 (s, 3H, CH₃), 3.81 (s, 3H, CH₃), 7.47 (s, 1H, CH).

Synthesis of 1,2-phenylene-dimethanol 8,9

To a solution of boronhydride (sol 1,5M in THF/diethyl ether, 50ml) at 0°C under argon atmosphere, a solution of the appropriate acid or ester (24 mmol) in THF (40 ml) was added. The mixture was stirred at 0°C for 2h and then under reflux for 20h. After cooling methanol was added to the mixture and the solvent evaporated under reduced pressure. The resulting crude was dissolved in ethyl acetate and washed with a saturated solution of sodium hydrogen carbonate. The organic layer was dried on Na₂SO₄ and evaporated.

(**3-Fluoro-1,2-phenylen)dimethanol 9**: m.p. 73.0°C; yield 99%; ¹H NMR DMSO (ppm): 3.41 (br, 2H, OH), 4.68 (d, 2H, CH₂), 7.05 (m, 3H, Ar), 7.16 (d, 2H, CH₂), 7.30 (t).

(**4,5-Dimetil-1,2-phenylen)dimethanol 8**: m.p. 103.0°C; yield 80%; ¹H NMR CDCl₃ (ppm): 2.24 (s, 3H, CH₃), 3.33 (br, 1H, OH), 4.61 (d, 2H, CH₂), 7.08 (s, 1H, CH).

Synthesis of *o*-phtalaldehydes 1a,1b

To a mixture of DCM (100 ml) and oxalyl chloride (8ml, 88 mol) at -78°C and under argon atmosphere, a mixture of DMSO (13.6 ml, 0.176 mol) and DCM (25 ml) was added dropwise. After 3-5 min a solution of the appropriate 1,2-dimethanol **8,9** (40 mol) in DCM-DMSO (10 ml) was added dropwise. After 30 min 100 ml of triethylamine were added dropwise. After stirring for 10 min the mixture was warmed to rt and poured in water and ice. It was extracted in DCM and the organic layer dried and evaporated. The crude was purified in column using DCM as eluent.

4,5-Dimethyphtalaldehyde 1a: m.p. 95.0-100.0°C; yield 87%; IR 1688 (CHO) cm⁻¹; ¹H NMR CDCl₃ (ppm): 1.77 (s, 3H, CH₃), 7.28 (s, 1H, CH), 10.3 (s, 1H, CHO).

3-Fluorophtaladehyde 1b: m.p. 75.0-80.0°C; yield 80%; IR 1700 (CHO) cm⁻¹; ¹H NMR CDCl₃ (ppm): 7.75 (d), 7.81 (m), 8.04 (m), 10.5 (CHO, 10.6 (CHO).

Synthesis of (2-bromo-phenylen)methanols 11c,d

To a solution of benzoic acid **10c,d** (20 mmol) in THF (50 ml), a solution of BH₃/THF (solution 1.0M in THF, 28.0 ml, 30 mmol) was added dropwise and the mixture was

stirred at rt for 3h and 30 min. After cooling at 0°C a solution of HCl 1M was added and the mixture was extracted with diethyl ether. The organic layer was washed with water and saturated solution of sodium hydrogen carbonate, dried and evaporated.

(**2-Bromo-5-methoxyphenyl)methanol 11c**: m.p. 45.0-46.0°C; yield 99%; IR 3408 (OH) cm⁻¹; ¹H NMR DMSO (ppm): 3.77 (s, 6H, 2xCH₃),4.49 (d, 2H, J = 5.5 Hz, CH₂), 5.47 (d, 1H, J = 5.5 Hz, OH), 6.79 (dd J = 3.1 e 8.7 Hz, H-4), 7.13 (d J = 3.1 Hz, H-6), 7.44 (d J = 8.8 Hz, H-3); ¹³C NMR DMSO (ppm): 55.2 (q), 62.5 (t), 110.9 (s), 113.6 (d), 113.9 (d), 132.5 (d), 142.2 (s), 158.8 (s).

(2-Bromo-4,5-dimethoxyphenyl)methanol 11d: m.p. 95.0°C; yield 90%; IR 3498 (OH) cm⁻¹; ¹H NMR DMSO (ppm): 3.77 (s, 3H, CH₃), 3.77 (s, 3H, CH₃), 4.45 (d, 2H, J = 5.5 Hz, CH₂), 5.35 (d, 1H, J = 5.5 Hz, OH), 7.10 (s, 1H, H-6), 7.12 (s, 1H, H-3); ¹³C NMR DMSO (ppm): 55.5 (q), 55.9 (q), 62.3 (t), 110.7 (s), 111.6 (d), 115.2 (d), 132.9 (s), 148.1 (s), 148.2 (s).

Synthesis of 2-bromo-benzaldehydes 12c,d

To a solution of the appropriate phenyl methanol **11c,d** (1.7 mmol) in DCM, 2.4 mmol (1.9 mmol) of Dess-Martin periodinane are added under argon atmosphere. The mixture was stirred for 2 hours. Diethyl ether was added and the organic phase was washed with 5% solution of $Na_2S_2O_3$ and saturated solution of $NaHCO_3$. The organic phase was dried using Na_2SO_4 and evaporated. The crude was purified in column using DCM as eluent.

2-Bromo-5-methoxybenzaldehyde 12c: m.p. 74.0-75.0°C; yield 98%; IR (CHO) 1682 cm⁻¹; ¹H NMR DMSO (ppm): 3.83 (s, 3H, CH₃), 7.23 (dd, 1H, J = 3.2 e 8.7 Hz, H-4), 7.34 (d, 1H J = 3.2 Hz, H-6), 7.70 (d, 1H J = 8.7 Hz, H-3), 10.2 (s, 1H, CHO); ¹³C NMR DMSO (ppm): 55.7 (q), 113.6 (d), 116.3 (s), 122.7 (d), 133.6 (s), 134.8 (d), 158.9 (s), 191.4 (d).

2-Bromo-4,5-dimethoxybenzaldehyde 12d: m.p. 148.0-149.0 °C; yield 90%; IR (CHO) 1676 cm⁻¹; ¹H NMR DMSO (ppm): 3.83 (s, 3H, CH₃), 3.91 (s, 3H, CH₃), 7.31 (s, 1H, H-6), 7.32 (s, 1H, H-3), 10.06 (s, 1H, CHO); ¹³C NMR DMSO (ppm): 55.6 (q), 56.4 (q), 110.4 (d), 115.9 (d), 119.3 (s), 125.7 (s), 148.6 (s), 154.4 (s), 190.1 (d).

Synthesis of 2-(2-bromo-phenyl)1,3-dioxolanes 13c,d

To a solution of 2-bromo-benzaldehyde **12c,d** (3.3 mmol) in toluene, ethyl glicole (9.2 mmol, d= 1.113, 0.6 ml) and a catalytic amount of *p*-toluensulfonic acid are added. The mixture was stirred at reflux for 24h using Dean-Stark trap. After cooling, the organic phase was washed with a saturated solution of NaHCO₃ and dried and evaporated. The crude was purified by BIOTAGE chromatography using cycloexane/ethyl acetate (98/2) as eluent.

2-(2-Bromo-5-methoxyphenyl)-1,3-dioxolane 13c: oil, yield 90%; ¹H NMR CDCl₃ (ppm): 3.79 (s, 3H, CH₃), 4.05-4.16 (m, 4H, 2xCH₂), 6.04 (s, 1H, CH), 6.78 (dd, 1H J = 3.1 e 8.8 Hz, H-4), 7.16 (d, 1H J = 3.1 Hz, H-6), 7.44 (d, 1H J = 8.8 Hz, H-3); ¹³C NMR CDCl₃ (ppm): 55.5 (t), 65.4 (q), 102.4 (d), 113.0 (d), 113.1 (s), 116.7 (d), 133.6 (d), 137.4 (s), 159.0 (s).

2-(2-Bromo-4,5-dimethoxiphenyl)-1,3-dioxolane 13d: m.p. 98.0-99.0°C; yield 90%; ¹H NMR DMSO (ppm): 3.76 (s, 3H, CH₃), 3.79 (s, 3H, CH₃), 3.95-4.11 (m, 4H, 2xCH₂), 5.85 (s, 1H, CH), 7.05 (s, 1H, H-6), 7.14 (s, 1H, H-3); ¹³C NMR DMSO (ppm): 56.1 (q), 56.4 (q), 65.3 (t), 102.3 (d), 111.1 (d), 113.0 (s), 115.9 (d), 128.5 (s), 148.6 (s), 150.5 (s).

Synthesis of 2-(1,3-dioxolan-2-yl)benzaldehydes 14c,d

To a solution of dioxolane **13c,d** (2.5 mmol) in THF (8ml) at -78°C, *n*-butil lithium (solution 1.6 M in *n*-hexanes, 2.5 mmol, 1.6 ml) are added during 1h. The mixture was stirred for 1h at -78°C. After that, 0.3 ml of DMF are added and the mixture was warmed to rt during 1-5h. A saturated solution of NH₄Cl was added and the mixture extracted in DCM. The organic layer was dried with Na₂SO₄ and evaporated. The crude was purified in column using DCM as eluent.

2-(1,3-Dioxolan-2-yl)-5-methoxybenzaldehyde 14c: oil, yield 86%; IR (CHO) 1682 cm⁻¹; ¹H NMR DMSO (ppm): 3.88 (s, 3H, CH₃), 3.99-4.10 (m, 4H, 2xCH₂), 6.39 (s, 1H, CH), 7.10-7.20 (m, 2H, H-4 e H-6), 7.89 (d, 1H J = 8.5 Hz, H-3), 10.2 (s, 1H, CHO); ¹³C NMR DMSO (ppm): 55.7 (q), 64.9 (t), 99.8 (d), 112.2 (d), 114.4 (d), 127.3 (s), 131.7 (d), 141.8 (s), 163.4 (s), 190.3 (d).

2-(1,3-Dioxolan-2-yl)-4,5-methoxybenzaldehyde 14d: m.p. 77.0-78.0°C; yield 100%; IR (CHO) 1680 cm⁻¹; ¹H NMR DMSO (ppm): 3.98 (s, 3H, CH₃), 3.99 (s, 3H,

CH₃), 4.00-4.12 (m, 4H, 2xCH₂), 6.33 (s, 1H, CH), 7.22 (s, 1H, H-6), 7.41 (s, 1H, H-3), 10.3 (s, 1H, CHO); ¹³C NMR DMSO (ppm): 55.6 (q), 55.8 (q), 64.8 (t), 99.9 (d), 109.5 (d), 110.0 (d), 127.2 (s), 134.0 (s), 149. 1(s), 153.0 (s), 189.9 (d).

Synthesis of orto-phtaldehydes 1c,d

To a solution of orto-benzylaldehyde **14c**, **d** (1.5 mmol) in acetone (17 ml) a catalytic amount of *p*-toluensulfonic acid was added. The solution was stirred for 15 minutes at rt. Evaporated the solvent, the resulting residue was dissolved in DCM and it was washed with a solution of Na_2CO_3 and brine. The crude was purified by chromatography using DCM as eluent.

5-Methoxyphtaldehyde 1c: m.p. 37.1°C; yield 60%; IR (CHO) 1695 cm⁻¹; ¹H NMR CDCl₃ (ppm): 3.96 (s, 3H, CH₃), 7.23 (dd, 1H J = 2.6 e 8.5 Hz, H-4), 7.44 (d, 1H J = 2.6 Hz, H-6), 7.94 (d, 1H J = 8.5 Hz, H-3), 10.3 (s, 1H, CHO), 10.6 (1H, s, CHO); ¹³C NMR CDCl₃ (ppm): 56.0 (q), 114.9 (d), 118.7 (d), 129.4 (s), 134.6 (d), 138.6 (s), 163,9 (s), 191.1 (d), 192.0 (d).

4,5-Dimethoxyphtaldehyde 1e: m.p. 169.4-170.0; yield 88%; IR (CHO) 1680 cm⁻¹; ¹H NMR CDCl₃ (ppm): 3.94 (s, 6H, 2xCH₃), 7.54 (1, 2H, 2xCH), 10.5 (s, 2H, 2xCHO); ¹³C NMR CDCl₃ (ppm): 11.7 (d), 131.2 (s), 153.2 (s), 191.7 (d).

Synthesis of 1-Cyano-2-(2-aminophenyl)isoindoles 3

To a solution of sodium hydrogen sulfite (1.56 g, 0.015 mol) in water (38 ml), phthalaldehyde **1e** (2 g, 0.015 mol) was added. The mixture was stirred until the solid was dissolved, and the appropriate 1,2-phenylenediamine **2a-f** (0.015 mol) was added. The reaction was heated on a steam bath for 30 min at 40°C, then KCN (3.39 g, 0.052 mol) in water (8.0 ml) was added, and the mixture was heated for an additional 90 min. The solid formed upon cooling was filtered and purified by chromatography. In the case of derivative **3e**, the 1,2-phenylenediamine **1e** was first dissolved in DMF (15 ml) and then added to the reaction mixture.

1-Cyano-2-(2-aminophenyl)isoindole 3a: this compound was purified by flash chromatography by using DCM as eluent to give a solid: yield 65%; m.p. 150.0-151.0

°C; IR 3473, 3363 (NH₂), 2202 (CN) cm-1; 1H NMR (DMSO-d6) (ppm) 5.19 (2H, bs, NH₂), 6.70 (t, 1H, J=7.2 Hz, H-5'), 6.94 (d, 1H, J=7.2 Hz, H-2), 7.15-7.19 (m, 3H, H-4', H-5, H-6'), 7.31 (t, 1H, J=8.2 Hz, H-6), 7.66 (d, 1H, J=8.2 Hz, H-4), 7.78 (d, 1H, J=8.2 Hz, H-7), 7.86 (s, 1H, H-3); 13C NMR (DMSO-d6) ppm: 93.9 (s), 114.0 (s), 115.8 (d), 116.1 (d), 117.5 (d), 121.5 (d), 122.2 (d), 122.4 (d), 122.5 (s), 124.1 (s), 125.5 (d), 127.8 (d), 130.4 (d), 131.2 (s), 144.1 (s).

1-Cyano-2-(2-amino-3-methylphenyl)isoindole 3b: this compound was purified by chromatography using DCM-cyclohexane (7:3) as eluent to give compound **3b**:

yield 40%; m.p. 62.0-63.0 °C; IR 3481, 3388 (NH₂), 2200 (CN) cm-¹; ¹H NMR (CDCl₃) ppm: 2.25 (s, 3H, CH₃), 3.55 (s, 2H, NH₂), 6.80 (t, 1H, J=7.6 Hz, H-5'), 7.10 (d, 1H, J=7.6 Hz, H-6'), 7.14 (d, 1H, J=7.6 Hz, H-4'), 7.23 (t, 1H, J=8.1 Hz, H-5), 7.31 (t, 1H, J=8.1 Hz, H-6), 7.44 (s, 1H, H-3), 7.69 (d, 1H, J=8.1 Hz, H-4), 7.73 (d, 1H, J=8.1 Hz, H-7); 13C NMR (CDCl3) ppm: 17.6 (q), 92.2

(s), 113.0 (s), 117.9 (d), 118.3 (d), 120.5 (d), 120.9 (d), 123.2 (d), 123.8 (s), 124.1 (s), 124.6 (s), 125.5 (d), 125.9 (d), 131.8 (s), 131.9 (d), 140.5 (s).

1-Cyano-2-(22-amino-42-methoxyphenyl)isoindole 3c: this compound was purified by chromatography by using DCM as eluent to give a solid: yield 95%; m.p. 116.0-117.0°C; IR 3465, 3375 (NH2), 2198 (CN) cm⁻¹; ¹H NMR (CDCl₃) ppm: 3.64 (bs, 2H, NH₂), 3.80 (s, 3H, CH₃), 6.38 (bs, 1H, H-3'), 6.42 (d, 1H, J=7.9 Hz, H-5'), 7.12 (d, 1H, J=7.9 Hz, H-6'), 7.14 (t, 1H, J=8.6 Hz, H-5), 7.29 (t, 1H, J=8.6 Hz, H-6), 7.41 (s, 1H, H-3), 7.66 (d, 1H, J=8.6 Hz, H-4), 7.70 (d, 1H, J=8.6 Hz, H-7); 13C NMR (CDCl3) ppm: 55.4 (q), 95.4 (s), 101.4 (d), 104.5 (d), 113.9 (s),

117.5 (s), 118.2 (d), 120.8 (d x 2), 123.1 (d), 124.5 (s), 125.8 (d), 128.7 (d), 131.7 (s), 143.3 (s), 161.5 (s).

1-Cyano-2-(2'-amino-4',5'-dimethylphenyl)isoindole 3d: this compound was purified by chromatography by using DCM as eluent to give a solid: yield 85%; m.p. 119-120 °C; IR 3465, 3375 (NH₂), 2202 (CN) cm⁻¹; ¹H NMR (CDCl₃) ppm: 2.17 (s, 3H, CH₃), 2.22 (s, 3H, CH₃), 3.44 (s, 2H, NH₂), 6.64 (s, 1H, H-3'), 6.95 (s, 1H, H-6'), 7.12 (t, 1H, J=7.6 Hz, H-5), 7.27 (t,1H, J=7.6 Hz, H-6), 7.38 (s, 1H, H-3), 7.64 (d, 1H, J=7.6 Hz, H-4), 7.68 (d, 1H, J=7.6 Hz, H-7);

¹³C NMR (CDCl₃) ppm: 18.6 (q), 19.7 (q), 95.1 (s), 114.0 (s), 118.1 (d), 118.2 (d), 120.6 (d), 120.9 (d), 121.8 (s), 123.1 (d), 124.5 (s), 125.8 (d), 127.0 (s), 128.2 (d), 131.7 (s), 139.6 (s), 139.7 (s).

1-Cyano-2-(2'-amino-4',5'-dichlorophenyl)isoindole 3e: this compound was purified by chromatography by using DCM as eluent to give a solid: yield 77%; m.p. 71.0-72.0 °C; IR 3394, 3340 (NH₂), 2200 (CN) cm⁻¹; ¹H NMR (DMSO-*d*6) ppm: 5.75 (2H, s, NH₂), 7.12 (1H, s, H-3'), 7.16 (1H, t, J=8.4 Hz, H-5), 7.32 (1H, t, J=8.4 Hz, H-6), 7.58 (1H, s, H-6'), 7.66 (1H, d, J= 8.4 Hz, H-4), 7.78 (1H, d, J=8.4 Hz, H-7), 7.91 (1H, s, H-3);

¹³C NMR (DMSO-*d*6) ppm: 94.0 (s), 113.8 (s), 115.8 (s), 116.2 (d), 117.5 (d), 121.6 (d), 121.7 (s), 122.4 (d), 122.6 (d), 124.2 (s), 125.8 (d), 129.6 (d), 131.2 (s), 132.9 (s), 144.7 (s).

1-Cyano-5,6-dimethyl-2-(2'-amino-4'-methoxiphenyl)-isoindole 3f: m.p. 179.0-180.0°C; yield 55%; IR: 3487, 3392 (NH₂), 2195 (CN) cm⁻¹; ¹H NMR CDCl₃ (ppm): 2.34 (3H, s, CH₃), 2.38 (3H, s, CH₃), 3.63 (2H, s, NH₂), 3.79 (3H, s, CH₃), 6.35-6.42 (1H, m), 7.11 (1H, d), 7.25 (1H, s), 7.20 (1H, t, J=7.6 Hz, H-5), 7.40-7.45 (2H, m);

¹³C NMR CDCl₃ (ppm): 20.5 (q), 20.9 (q), 55.5 (q), 94.3 (s), 101.4 (d), 104.4 (d), 114.5 (s), 117.2 (d), 117.7 (s), 119.5 (d), 119.9 (d), 124.0 8s), 128.8 (d), 131.4 (s), 133.2 (s), 136.4 (s), 143.5 (s), 161.4 (s).

1-Cyano-5,6-dimethoxy-2-(2'-amino-4'-methoxyphenyl)-isoindole 3g: m.p. 223.0-224.0°C; yield 60%; IR: 3469, 3371 (NH₂), 2200 (CN) cm⁻¹; ¹H NMR CDCl₃ (ppm): 3.64 (2H, s, NH₂), 3.82 (3H, s, CH₃), 3.93 (3H, s, CH₃), 3.98 (3H, s, CH₃), 6.39-6.44 (2H, m), 6.93 (1H, d,), 6.90 (1H, s), 6.95 (1H, s), 7.21 (1H, s);

¹³C NMR CDCl₃ (ppm): 55.5 (q), 55.9 (q), 56.1 (q), 94.7 (s), 96.2 (d), 98.3 (d), 101.4 (d), 104.4 (d), 114.5 (s), 117.8 (s), 119.7 (s), 119.8 (s), 128.2 (d), 128.9 (d), 143.5 (s), 148.8 (s), 151.1 (s), 161.3 (s)

Cyano-7-fluoro-2-(2'-amino-4'-methoxyphenyl)isoindole 3h: m.p. 118.0-119.0°C; yield 45%; IR: 3481, 3381 (NH₂), 2212 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 3.65 (2H, bs, NH₂), 3.81 (3H, s, CH₃), 6.39-6.45 (2H, m), 6.80-7.46 (5H, m);

¹³C NMR CDCl₃ (ppm): 52.5 (q), 91.0 (s), 102.9 (d), 103.4 (d), 113.9 (s), 116.0 (d), 117.9 8s), 120.5 (d), 123.9 (d), 124.6 (s), 127.8 (d), 130.4 (s), 131.2 (s), 135.2 (s), 140.5 (s), 159.4 (s).

1-Cyano-6-methoxy-2-(2'-amino-4'-methoxyphenyl)- isoindole 3i: m.p. 146.0-147.0°C; yield 42%; IR: 3477, 3423 (NH₂), 2200 (CN) cm⁻¹; ¹H NMR CDCl₃ (ppm): 3.65 (2H, s, NH₂), 3.82 (3H, s, CH₃), 3.89 (3H, s, CH₃), 6.38-6.45 (2H, m), 6.85 (1H, dd,), 6.93 (1H, d), 6.93 (1H, d), 7.14 (1H, d), 7.32 (1H, d), 7.55 (1H, dd);

¹³C NMR CDCl₃ (ppm): 55.4 (q), 55.5 (q), 95.0 (d), 101.5 (d), 104.5 (d), 114.5 (s), 117.7 (s), 118.4 (d), 120.6 (s), 121.2 (d), 122.2 (d), 128.8 8d), 133.2 (s), 143.4 (s), 158.5 (s), 161.4 (s).

1-Cyano-5-methoxy-2-(2'-amino-4'-methoxyphenyl)- isoindole **3**j: m.p. 144.0-145.0°C; yield 40%; IR: 3465, 3375 (NH₂), 2198 (CN) cm⁻¹; ¹H NMR CDCl₃ (ppm): 3.63 (2H, s, NH₂), 3.83 (3H, s, CH₃), 3.85 (3H, s, CH₃), 6.38-6.45 (2H, m), 6.90 (1H, d,), 7.00 (1H, dd), 7.12-7.27 (3H, m), 7.61 (1H, d);

¹³C NMR CDCl₃ (ppm): 55.3 (q), 55.5 (q), 97.2 (d), 101.4 (d), 104.5 (d), 117.7 (s), 118.1 (s), 119.5 (d), 119.6 (d), 120.8 (d), 124.9 (s), 127.9 (s), 128.8 (d), 143.3 (s), 156.2 (s), 161.4 (s).

2(2-amino-3,6-dimethoxyphenyl-2H-isoindole-1-carbonitrile 31: m.p. 161.5-162.7°C; yield 20%; IR 3486,3389 (NH2), 2202 (CN) cm⁻¹; ¹H NMR DMSO-d6 (ppm):3.61 (3H, s, OCH₃), 3.81 (3H, s, OCH₃), 4.73 (2H, s, NH₂), 6.35 (1H, d, J=8.9 Hz, H-5'), 6.95 (1H, d, J=8.9 Hz, H-4'), 7.12 (1H, t, J=6.3 Hz, H-4), 7.29 (1H, t, J=6.3 Hz, H-3), 7.63 (1H, d, J=8.5 Hz, H-5), 7.73-7.78 (2H, m, H-2, H-6);

¹³C NMR CDCl₃ (ppm): 97.16 (d), 111.37 (d), 111.84 (s), 114.09 (s), 117.59 (d), 121.61 (d), 122.13 (d), 122.58 (d), 124.14 (s), 125.37 (d), 131.00 (s),135.69 (s), 135.75 (s), 141.09 (s), 149.43 (s).

Synthesis of 1-cyano-2-(2'-amino-4'-methixy-phenyl)-isoindole 3k

To a solution of sodium hydrogen sulfite (1.56 g, 0.015 mol) in water (38 ml), phthalaldehyde **1e** (2 g, 0.015 mol) was added. The mixture was stirred until the solid was dissolved, and the appropriate 1,2-phenylenediamine **2c** (0.015 mol) was added. The reaction was heated on a steam bath for 30 min at 40°C, then KCN (3.39 g, 0.052 mol) in water (8.0 ml) was added, and the mixture was heated for an additional 90 min. The solid formed upon cooling was filtered and purified by chromatography in DCM.

1-cyano-2-(2'-amino-4'-methoxy-phenyl)-isoindole 3k: m.p. 105.0-106.0°C; yield 42%; IR: 3452, 3365 (NH₂), 2200 (CN) cm⁻¹; ¹H NMR CDCl₃ (ppm): 3.38 (2H, bs, NH₂), 3.77 (3H, s, CH₃), 6..82-6.96 (3H, m, H-3', H-4' e H-6'), 7.12 (1H, d, J=7.9 Hz, H-6'), 7.17(1H, t, J=8.0 Hz, H-5), 7.27 (1H, t, J=8.0 Hz, H-6), 7.47 (1H, s, H-3), 7.67.7.74 (2H, m, H-4 e H-7);

¹³C NMR CDCl₃ (ppm): 55.9 (q), 95.0 (s), 112.7 (d), 113.8(s), 117.8 (d), 118.2 (d), 118.3(d), 120.4 (d), 120.9 (d), 123.3 (d), 124.4 (s), 124.5 (s), 126.0 (d), 131.8 (s), 135.7 (s), 152.5 (s).

Synthesis of 1-cyano-2-(2'-amino-4'-hydroxyphenyl)isoindoles 3

To a solution of sodium hydrogen sulfite (1.56 g, 0.015mol) in water (38 ml), phthalaldehyde **1a-b**, **1d** (0.015 mol) was added. The mixture was stirred until the solid was dissolved, and the 3,4-diamino phenol **2g** (0.015 mol) was added. The reaction was stirred at rt for 3h, then KCN (3.39 g, 0.052 mol) in water (8.0 ml) was added, and the mixture was stirred at rt for additional 24h. The solid formed upon cooling was filtered and purified by chromatography using DCM as eluent.

1-Cyano-5,6-dimethyl-2-(2'-amino-4'-hydroxyphenyl)-isoindole 3m: m.p. 197.2-197.8°C; yield 10%; IR: 3487 e 3392 (NH₂), 3328 (OH), 2196 (CN) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 2.30 (3H, s, CH₃), 2.34 (3H, s, CH₃), 4.92 (2H, bs, NH₂), 6.12 (1H, dd, J=2.5,8.5 Hz, H5'), 6.32 (1H, d, J=2.5Hz, H3'), 6.94 (1H, d, J=8.5 Hz, H6'), 7.39 (1H, s, H4), 7.47 (1H, s, H7), 7.57 (1H, s, H3), 9.50 (1H, s, OH);

¹³C NMR DMSO-d6 (ppm): 20.0 (q), 20.3 (q), 93.2 (s), 101.6 (d), 103.8 (d), 114.6 (s), 115.4 (s), 116.3 (d), 120.0 (d), 121.5 (d), 123.4 (s), 128.6 (d), 130.7 (s), 131.8 (s), 135.5 (s), 145.1 (s), 159.0 (s).

1-Cyano-6-methoxy-2-(2'-amino-4'-hydroxyphenyl)-isoindole 30: m.p. 211.2-212.0°C; yield 13%; IR: 3430 e 3345 (NH₂), 3303 (OH), 2200 (CN) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.79 (3H, s, OCH₃), 4.92 (2H, bs, NH₂), 6.10 (1H, dd, J=2.5, 8.5 Hz, H5'), 6.30 (1H, d, J=2.5 Hz, H3'), 6.94 (1H, d, J=8.5 Hz, H6'), 6.96 (1H, dd, J=2.2, 9.1Hz, H5), 7.07 (1H, d, J=2.2 Hz, H7), 7.54 (1H, d, J=9.1 Hz, H4), 7.58 (1H, s, H3), 9.49 (1H, s, OH);

¹³C NMR DMSO-d6 (ppm): 55.0 (q), 94.5 (s), 98.2 (d), 101.6 (d), 103.7 (d), 114.2 (s), 115.3 (s), 118.9 (d), 120.0 (d), 121.3 (d), 124.3 (s), 127.1 (s), 128.6 (d), 145.2 (s), 155.1 (s), 159.0 (s).

1-Cyano-5-methoxy-2-(2'-amino-4'-hydroxyphenyl)-isoindole 3p: m.p. 194.4-195.5°C; yield 10%; IR: 3475 e 3372 (NH₂), 3266 (OH), 2198 (CN) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.84 (3H, s, OCH₃), 4.97 (2H, bs, NH₂), 6.09 (1H, dd, J=2.5, 8.5 Hz, H5'), 6.29 (1H, d, J=2.5 Hz, H3'), 6.79 (1H, dd, J=2.2, 9.0 Hz, H5), 6.89 (1H, d, J=2.2

Hz, H7), 6.93 (1H, d, J=8.5Hz, H6'), 7.64 (1H, d, J=9.0 Hz, H4), 7.66 (1H, s, H3), 9.47 (1H, s, OH);

¹³C NMR DMSO-d6 (ppm): 55.2 (q), 93.8 (s), 94.7 (d), 101.6 (d), 103.7 (d), 114.7 (s), 115.3 (s), 116.9 (d), 120.0 (s), 121.3 (d), 123.0 (d), 128.6 (d), 132.3 (s), 145.2 (s), 157.9 (s), 159.0 (s).

1-Cyano-4-fluoro-2-(2'-amino-4'-hydroxyphenyl)-isoindole 3q: m.p. 203.6-204.7°C; yield 20%; IR: 3482 e 3376 (NH₂), 3234 (OH), 2217 (CN) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 5.09 (2H, bs, NH₂), 6.10 (1H, dd, J=2.5, 8.5 Hz, H5'), 6.30 (1H, d, J=2.5 Hz, H3'), 6.98 (1H, d, J=8.5 Hz, H6'), 7.02-7.14 (2H, m, H5,H7), 7.58 (1H, dt, J_F=2.5 Hz, J=6.7 Hz, H6), 7.87 (1H, d, J_F=2.5 Hz, H3), 9.52 (1H, s, OH);

¹³C NMR DMSO-d6 (ppm): 92.1 (d, $J_F=2.8$ Hz), 101.6 (d), 103.7 (d), 108.1 (dd, $J_F=16.7$ Hz), 114.0 (s), 114.7 (s), 117.90 (dd, $J_F=4.3$ Hz), 121.1 (d, $J_F=18$ Hz), 122.4 (dd, $J_F=6.4$ Hz), 123.7 (d), 126.7 (d, $J_F=6.7$ Hz), 128.7 (d), 145.3 (s), 153.7 (d, $J_F=248$ Hz), 159.3 (s).

Synthesis of 1-cyano-2-(2'-amino-4'-hydroxyphenyl)-isoindole 3n

To a solution of sodium hydrogen sulfite (1.56 g, 0.015mol) in water (38 ml), phthalaldehyde **1c** (0.015 mol) was added. The mixture was stirred until the solid was dissolved, and the 3,4-diamino phenol **2g** (0.015 mol) was added. The reaction was stirred at rt for overnight, then KCN (3.39 g, 0.052 mol) in water (8.0 ml) was added, and the mixture was stirred at rt for additional 24h. The solid formed upon cooling was filtered and purified by chromatography using DCM- ethyl acetate 8/2 as eluent.

1-Cyano-5,6-dimethoxy-2-(2'-amino-4'-hydroxyphenyl)-isoindole 3n: m.p. 202.4-203.2°C; yield 30%; IR: 3467 e 3384 (NH₂), 3326 (OH), 2186 (CN) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.79 (3H, s, CH₃), 3.86 (3H, s, CH₃), 4.92 (2H, bs, NH2), 6.09 (1H, dd, J=2.5, 8.5 Hz, H5'), 6.30 (1H, d, J=2.5 Hz, H3'), 6.90 (1H, s, H4), 6.92 (1H, d, J=8.5Hz, H6'), 7.05 (1H, s, H7), 7.48 (1H, s, H3), 9.46 (1H, s, OH);

¹³C NMR DMSO-d6 (ppm): 55.3 (q), 55.5 (q), 93.6 (s), 95.8 (d), 99.3 (d), 101.6 (d), 103.7 (d), 114.8 (s), 115.5 (s), 119.1 (s), 121.4 (d), 127.3 (s), 128.6 (d), 145.2 (s), 148.0 (s), 150.7 (s), 158.9 (s).

Synthesis of isoindolo[2,1a]quinoxalin-6-imino 4

A solution of 1-cyano-2-(2'-amino-phenyl)isoindoles **3** (3 mmol) in acetic acid (10 ml) was stirred under reflux for 30 min. After cooling the mixture was evaporated under reduced pressure and the resulting solid crystallized in ethanol. In case of derivatives **4m**, **4o**, **4q** the precipitate formed after cooling was filtered, collected and crystallized in ethanol.

5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 4a: m.p. 248.5-249.9°C; yield quant.; IR 3367, (NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 6.87 (2H, bs, NH₂), 7.28-7.36 (3H, m, H-4, H-8, H-9), 7.46 (1H, t, J=7.0 Hz, H-3), 7.58 (1H, dd, J=1.2, 8.1 Hz, H-2), 7.86-7.90 (1H, m, H-7), 8.38 (1H, d, J=8.1, H-5), 8.42-8.47 (1H, m, H-10), 8.93 (1H, s, H-6). ¹³C NMR DMSO-d6 (ppm): 117.66 (d), 108.02 (s), 115.64 (d), 119.43 (d), 119.54 (s), 120.16 (d), 122.14 (d), 122.67 (d), 123.06 (d), 123.60 (s), 125.46 (d), 125.72 (s), 126.92 (d), 138.35 (s), 151.40 (s).

4-methyl-5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 4b: m.p. 210.3-211.6°C; yield 95%; IR 3309, 3320 (NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 2.60 (3H, s, CH₃), 6.81 (2H, bs, NH₂), 7.17-7.37 (4H, m, H-3, H-4, H-8, H-9), 7.85-7.89 (1H, m. H-7), 8.22 (1H, d, J=7.7 Hz, H-5), 8.43-8.48 (1H, m, H-10), 8.89 (1H, s, H-6);

¹³C NMR DMSO-d6 (ppm): 18.23 (q), 99.49 (d), 107.97 (s), 113.39 (d), 119.36 (d), 119.51 (s), 120.23 (d), 121.58 (d), 122.15 (s), 122.54 (d), 123.05 (d), 123.28 (s), 123.83 (s), 127.59 (d), 133.39 (s), 150.37 (s).

3-methoxy-5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 4c: m.p. 242.5-243.0°C; yield quant.; IR 3303, 3500(NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.86 (3H, s, OCH₃), 6.83 (2H, bs, NH₂), 6.93 (1H, dd, J=2.8, 8.9 Hz, H-4), 7.04 (1H, d, J=2.7 Hz, H-2), 7.28 (1H, dd, J=2.9, 6.5 Hz, H-5, H-8), 7.84 (1H, m, H-9), 8.29 (1H, d, J=9.0 Hz, H-7), 8.42 (1H, m, H-10), 8.82 (1H, s, H-6).

¹³C NMR DMSO-d6 (ppm): 55.28 (q), 107.51 (d), 107.52 (d), 110.40 (d), 116.61 (d), 117.89 (s), 118.58 (s), 119.22 (d), 119.53 (s), 119.98 (d), 122.36 (d), 122.81 (d), 125.67 (s), 129.93 (s), 151.73 (s), 158.24 (s).

| Crystallographic Data of compound 4c | | | | |
|---|---------------------------------------|--|--|--|
| Empirical formula | $C_{18}H_{17}N_3O_3$ | | | |
| Formula weight | 323.35 | | | |
| Wavelength (Å) / Temperature (K) | 1.54184 / 293(2) | | | |
| Crystal system | monoclinic | | | |
| Space group | $P2_1 / c$ | | | |
| <i>a</i> (Å) | 12.8001(2) | | | |
| <i>b</i> (Å) | 17.7147(3) | | | |
| <i>c</i> (Å) | 6.91504(11) | | | |
| β (deg) | 101.1176(16) | | | |
| Volume (Å ³) | 1538.56(4) | | | |
| Z (molecules/unit cell) | 4 | | | |
| Calculated density (Mg m ⁻³) | 1.396 | | | |
| Absorption coefficient, μ (mm ⁻¹) | 0.796 | | | |
| <i>F</i> (000) | 680.0 | | | |
| Total reflections | 18882 | | | |
| Index ranges (h, k, l) | -15/15, -20/21, -8/8 | | | |
| Independent (unique) reflections / R_{int} | 2754 / 0.0242 | | | |
| Observed reflections $[I > 2\sigma(I)]$ | 2562 | | | |
| Data / parameters / restraints | 2754 / 219 / 0 | | | |
| Goodness–of–fit ^a on F^2 | 1.069 | | | |
| Final <i>R</i> indices $[I > 2\sigma(I)]$ | $R_1^{b} = 0.0458; wR_2^{c} = 0.1313$ | | | |
| Largest difference peak and hole ($e^{A^{-3}}$) 0.28 and -0.21 | | | | |
| ^a Goodness–of–fit = $[\Sigma (w (F_o^2 - F_c^2)^2] / (N_{obsevns} - N_{params})]^{1/2}$, based on all data; | | | | |
| | | | | |

^b $R_1 = \Sigma (|F_o| - |F_c|) / \Sigma |F_o|;$

^c $wR_2 = [\Sigma[w (F_o^2 - F_c^2)^2] / \Sigma[w (F_o^2)^2]]^{1/2}.$

2,3-dimethyl-5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 4d: m.p. 238.2-239.0°C; yield 90%; IR 3362, 3401 (NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 2.34 (3H, s, CH₃), 2.40 (3H, s, CH₃), 6.71 (2H, bs, NH₂), 7.25-7.30 (2H, m, H-8, H-9), 7.37 (1H, s, H-2), 7.83-7.88 (1H, m, H-7), 8.11 (1H, s, H-5), 8.39-8.43 (1H, m, H-10), 8.83 (1H, s, H-6);

¹³C NMR DMSO-d6 (ppm): 19.37 (q), 21.05 (q), 106.91 (d), 108.06 (s), 115.83 (d),

119.26 (d), 119.38 (s), 120.13 (d), 121.52 (s), 122.28 (d), 122.85 (d), 125.70 (s), 125.74 (d), 131.02 (s), 135.46 (s), 136.41 (s), 150.98 (s).

2,3-dichloro-5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 4e: m.p. 241.9-242.6°C; yield quant.; IR 3557, 3398 (NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 7.14 (2H, bs, NH₂), 7.30-7.35 (2H, m, H-8, H-9), 7.69 (1H, s, H-2), 7.83-7.89 (1H, m, H-10), 8.77 (1H, s, H-5), 9.03 (1H, s, H-6);

¹³C NMR DMSO-d6 (ppm): 99.34 (s), 107.72 (d), 109.23 (s), 117.57 (s), 119.69 (s), 119.91 (d), 120.110 (s), 123.11 (d), 123.34 (s), 123.45 (d), 123.50 (s), 125.88 (s), 128.91 (d), 138.81 (d), 152.31 (d).

3-methoxy-8,9-dimethyl-5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 4f: m.p. 243.1-244.0°C; yield 95%; IR 3399, 3398 (NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 2.38 (3H, s, CH₃), 2.42 (3H, s, CH₃), 3.84 (3H, s, OCH₃), 6.74 (2H, bs, NH₂), 6.89 (1H, dd, J=2.8, 8.9 Hz, H-4), 7.00 (1H, d, J=2.7 Hz, H-2), 7.58 (1H, s, H-7), 8.19-8.23 (2H, m, H-5, H-10), 8.64 (1H, s, H-6);

¹³C NMR DMSO-d6 (ppm): 20.23 (q), 20.38 (q), 56.24 (q), 106.31 (d), 106.80 (s), 107.47 (d), 110.07 (d), 116.36 (d), 118.09 (d), 118.96 (d), 119.06 (s), 125.16 (s), 132.14 (s), 132.25 (s), 139.75 (s), 151.62 (s), 157.95 (s).

3-methoxy-8,9-dimethoxy-5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 4g: m.p. 356.8-358.2°C; yield quant; IR 3439, 3366 (NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.84 (3H, s, OCH₃), 3.85 (3H, s, OCH₃), 3.95 (3H, s, OCH₃), 6.77 (2H, bs, NH₂), 6.87 (1H, dd, J=2.8, 8.9 Hz, H-4), 6.97 (1H, d, J=2.7, H-2), 7.14 (1H, s, H-7), 7.63(1H, s, H-10), 8.13 (1H, d, J=9.0 Hz, H-5), 8.55 (1H, s, H-6);

¹³C NMR DMSO-d6 (ppm): 55.22 (q), 55.84 (q), 55.98 (q), 97.66 (d), 99.24 (d), 106.45
(d), 107.37 (s), 107.47 (d), 109.81 (d), 114.98 (s), 115.86 (d), 118.39 (s), 121.36 (s), 139.27 (s), 148.26 (s), 148.40 (s), 151.51 (s), 157.61 (s);

2-methoxy-8,9-dimethoxy-5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 4k: m.p. 182.0-182.3°C; yield quant.; IR 3500, 3347 (NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.94 (3H, s, OCH₃), 6.70 (2H, bs, NH₂), 7.12 (1H, dd, J=2.7, 8.9 Hz, H-3), 7.28-7.33 (2H, m, H-9, H-9), 7.53 (1H, d, J=8.9 Hz, H-2), 7.86-7.90 (1H, m, H-7), 7.91 (1H, d, J=2.6 Hz, H-4), 8.41-8.45 (1H, m, H-10), 8.98 (1H, s, H-6)

¹³C NMR DMSO-d6 (ppm): 55.87 (q), 99.16 (d), 108.07 (d), 115.57 (d). 116.73 (s), 119.57 (d), 120.37 (d), 122.59 (d), 123.80 (d), 125.80 (s), 126.33 (d), 131.68 (s), 149.83

(s), 155.22 (s), 155.74 (s), 159.66 (s).

1,4-dimethoxy-5,6-dihydroisoindolo[2,1-a]quinoxalin-6-amine 41: m.p. 272.4-272.9°C; yield 90%; IR 3352, 3339 (NH₂) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.87 (3H, s, OCH₃), 4.06 (3H, s, OCH₃), 6.87-7.02 (4H, m, NH₂, H-3, H-4), 7.25-7.35 (2H, m, H-8, H-9), 7.90-7.95 (1H, m, H-7), 8.41-8.45 (1H, m, H-10);

¹³C NMR DMSO-d6 (ppm): 55.81 (q), 56.39 (q), 103.27 (d), 107.51 (d), 108.33 (s), 113.91 (d), 114.88 (s), 118.17 (s), 119.71 (d), 119.76 (d), 122.69 (d), 122.89 (d), 125.20 (s), 130.94 (s), 144.58 (s), 147.39 (s), 150.71 (s).

3-idrossi-5*H***-8,9-dimetil-isoindolo[2,1***a***]chinossalin-6-imino 4m**: m.p. 289.1-289.4°C, yield 55%, IR: 3390 (OH), 3284 e 3264 (NH) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 2.38 (3H, s, CH₃), 2.42 (3H, s, CH₃), 6.77 (1H, bs, NH), 6.78 (1H, dd, J=2.6, 8.8 Hz, H4), 6.91 (1H, d, J=2.6 Hz, H2), 7.59 (1H, s, H7), 8.10 (1H, d, J=8.8 Hz, H5), 8.14 (1H, s, H10), 8.57 (1H, s, H6), 9.08 (1H, bs, OH), 9.59 (1H, bs, NH).

¹³C NMR DMSO-d6 (ppm): 20.2 (q), 20.4 (q), 106.3 (d), 106.7 (s), 109.4 (d), 110.9 (d), 116.2 (d), 117.1 (s), 118.1 (d), 119.0 (d), 119.1 (s), 125.1 (s), 132.0 (s), 132.1 (s), 139.6 (s), 151.4 (s), 156.1 (s).

6-amino-8,9-dimethoxy-5,6-dihydroisoindolo[2,1-a]quinoxalin-3-ol, 4n: m.p. 242.5-243.0°C; yield quant.; IR 3677, 3557 (NH₂), 3348 (OH) cm⁻¹; ¹H NMR DMSO-d6: 3.85 (3H, s, OCH₃), 3.92 (3H, s, OCH₃), 6.68 (3H, m, NH2, H-4), 6.86 (1H, d, J=2.4 Hz, H-2), 7.13 (1H, s, H-10), 7.61 (1H, s, H-7), 8.02 (1H, d, J= 8.9 Hz, H-5), 8.49 (1H, s, H-6);

¹³C NMR DMSO-d6 (ppm):55.19 (q), 55.81 (q), 97.65 (d), 99.50 (d), 106.03 (d), 107.32 (s), 109.42 (d), 110.68 (d), 114.86 (d), 115.68 (s), 117.45 (s), 118.66 (s), 121.23 (s), 139.32 (s), 148.12 (s), 148.22 (s), 151.38 (s), 155.73 (s).

3-idrossi-5*H***-8-metossi-isoindolo[2,1***a***]chinossalin-6-imino 40:** m.p. 279.1-280.4 °C, yield 85%, IR: 3306 (OH), 3280 e 3244 (NH) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.92 (3H, s, CH₃), 6.72 (1H, dd, J=2.6, 8.8 Hz, H4), 6.87 (1H, bs, NH), 6.88 (1H, d, J=2.6 Hz, H2), 6.93 (1H, dd, J= 1.9, 9.1 Hz, H9), 7.63 (1H, d, J=1.9 Hz, H7), 7.73 (1H, d, J=9.1 Hz, H10), 8.07 (1H, d, J=8.8 Hz, H5), 8.65 (1H, s, H6), 9.05 (1H, bs, OH), 9.96 (1H, bs, NH).

¹³C NMR DMSO-d6 (ppm): 55.5 (q), 98.2 (d), 107.2 (s), 107.4 (d), 109.3 (d), 110.8 (d), 116.0 (d), 116.9 (d), 117.2 (s), 120.4 (s), 121.7 (d), 139.4 (s), 151.7 (s), 155.9 (s), 156.2 (s), 172.1 (s).

3-idrossi-5*H***-7-fluoro-isoindolo[2,1***a***]chinossalin-6-imino 4q: m.p. 340°C, yield 86 %, IR: 3387 (OH), 3264 e 3252 (NH) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 6.78 (1H, bs, NH), 6.81 (1H, dd, J=2.4, 8.9 Hz, H4), 6.93 (1H, d, J=2.4 Hz, H2), 7.03-7.29 (3H, m, H6-H8-H9), 7.69 (1H, d, J=8.2 Hz, H10), 8.20 (1H, d, J=8.9 Hz, H5) 8.92 (1H, bs, OH), 10.60 (1H, bs, NH). ¹³C NMR DMSO-d6 (ppm): 99.5 (s), 105.8 (d, J_F=4.1 Hz), 106.5 (dd, J_F=22.9 Hz), 108.4 (d), 108.8 (d, J_F=16.8 Hz), 109.5 (d), 111.7 (d), 116.3 (dd, J_F=3.6 Hz), 116.4 (s), 116.8 (d), 123.1 (dd, J_F=8.1 Hz), 128.6 (d, J_F=7.4 Hz), 139.9 (s), 150.2 (s), 154.5 (d, J_F=246.4 Hz).**

Synthesis of isoindole[2,1a]quinoxalin-6-one 5

A solution of 1-cyano-2-(2'-amino-phenyl)isoindoles **3** (3 mmol) in acetic acid (10 ml) was stirred under reflux for 30 min. After cooling the mixture was poured onto ice. The resulting precipitate was filtered off and crystallized in ethanol.

3-Methoxy-5*H***-8,9-dimethyl-isoindole[2,1***a***]quinoxalin-6-one 5f m.p. >410°C; yield 60%; IR: 3435 (NH), 1722 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 2.40 (3H, s, CH₃), 2.45 (3H, s, CH₃), 3.89 (3H, s, CH₃), 7.15 (s, 1H), 7.72 (s, 1H), 8.38-8.46 (m, 3H), 9.11 (s, 1H), 13.46 (1H, bs, NH).**

3-Methoxy-5H-8,9-dimethoxy-isoindolo[2,1*a***]quinoxalin-6-one 5g**: m.p.<410°C; yield 72%; IR: 3556 (NH), 1637 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 3.89 (3H, s, CH₃), 3.94 (3H, s, CH₃), 3.99 (3H, s, CH₃), 7.11-7.31 (2H, m), 7.82 (1H,s), 8.35-8.67 (2H, m), 9.1 (1H, d) 13.14 (1H, bs, NH).**3-Methoxy-5H-10-fluoro-isoindole[2,1***a***]quinoxalin-6-one 5h**: m.p. 246-247°C; yield 58%; IR: 3430 (NH), 1710 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 3.90 (3H, s, CH₃), 7.16-7.46 (2H, m), 7.85 (1H, d), 8.09 (1H, s), 8.47 (1H, d), 9.40 (1H, s), 13.62 (1H, bs, NH).

3,8-Dimethoxy-5H-isoindole[2,1a]quinoxalin-6-one 5i: m.p.298-299°C; yield 60%; IR: 3280 (NH), 1635 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 3.89 (1H, s, OCH₃), 3.95 (3H,s, OCH₃), 7.13-7.30 (3H, m,), 9.08 (1H, s), 13.2 (1H, bs, NH);

3,9-Dimethoxy-5*H***-isoindole[2,1***a***]quinoxalin-6-one 5j**: m.p. 283-284°C; yield 60%; IR: 3309 (NH), 1641 (CO) cm⁻¹; ¹H NMR DMSO (ppm): ¹H NMR DMSO (ppm): 3.89 (1H, s, OCH₃), 3.97 (3H,s, OCH₃), 7.09-7.14 (3H, m, NH₂, H-2'), 7.80-7.93 (3H, m, H-5', H-4', H-6), 8.36 (1H, d J=8.8 Hz, H-5), 8.50 (1H, s, H-3), 9.17 (1H, s, H-2), 12.84 (1H, bs, NH);

4-Methoxy-5*H***-isoindole[2,1***a***]quinoxalin-6-one 5k: m.p. 291°C; yield 90%; IR 3360 (NH), 1634 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 3.90 (3H, s, CH₃), 7.06 (1H, s), 7.09 (2H, m,), 7.45 (1H, s), 7.60 (1H, t), 8.00 (1H, s), 8.37 (2H, m), 13.4 (1H, bs, NH).**

3-Methoxy-5*H***-8,9-dimethoxy-isoindole[2,1***a***]quinoxalin-6-one 5**I: m.p. 282.7-289.0°C; yield 70%; IR: 3314 (NH), 1638 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 4.00 (3H, s, OCH₃), 4.10 (3H, s, OCH₃), 7.15-7.31 (2H, m, H-3', H-6'), 7.47 (1H, t, J=7.5 Hz, H-4), 8.62 (1H, t, J=7.50, H-5), 8.11 (1H, d, J=8.0 Hz, H-3'), 8.55 (1H, d, J=8.4, H-6'), 9.66 (1H, s, H-2), 13.14 (1H, bs, NH).

3-hydroxy-5*H***-8,9-dimethyl-isoindole[2,1***a***]quinoxalin-6-one 5m: m.p. >410°C, yield 88%, %, IR: 3280 (OH), 3189 (NH), 1637 (CO) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 2.38 (3H, s, CH₃), 2.43(3H, s, CH₃), 6.95 (1H, dd, J=2.4, 9.1 Hz, H4), 7.07 (1H, d, J=2.4 Hz, H2), 7.71 (1H, s, H7), 8.30 (1H, d, J=9.1 Hz, H5), 8.32 (1H, s, H10), 9.03 (1H, s, H6), 10.40 (1H, bs, OH), 13.01 (1H, bs, NH).**

3-hydroxy-5*H***-8,9-dimethoxy-isoindolo[2,1***a***]quinoxalin-6-one 5n**: m.p. 287.2-288°C, yield 50%, IR: 3316 (OH), 3264 (NH), 1640 (CO) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.88 (3H, s, CH₃), 3.97 (3H, s, CH₃), 6.93 (1H, dd, J=2.3, 9.0 Hz, H4), 7.04 (1H, d, J=2.3 Hz, H2), 7.26 (1H, s, H7),7.75 (1H, s, H10), 8.21 (1H, d, J=9.0 Hz, H5), 8.44 (1H, s, H6), 8.90 (1H, bs, OH), 13.04 (1H, bs, NH).

3-hydroxy-5*H***-8-methoxy-isoindole[2,1***a***]quinoxalin-6-one 50:** m.p. 335.0-335.7 °C, yield 83%, IR: 3306 (OH), 3280 (NH), 1635 (CO) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.89 (3H, s, CH₃), 6.94 (1H, dd, J=2.3, 9.0 Hz, H4), 7.06 (1H, d, J=2.3 Hz, H2), 7.09 (1H, dd, J= 1.7, 9.1 Hz, H9), 7.80 (1H, d, J=1.7 Hz, H7), 7.89 (1H, d, J=9.1 Hz, H5), 8.26 (1H, d, J=9.0 Hz, H10), 9.11 (1H, bs, OH), 13.05 (1H, bs, NH).

3-hydroxy-5*H***-9-methoxy-isoindole[2,1***a***]quinoxalin-6-one 5p:** m.p.316.4-317.7 °C, yield 89%, IR: 3326 (OH), 3309 (NH), 1641 (CO) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 3.89 (3H, s, CH₃), 7.00 (1H, dd, J=2.4, 9.1 Hz, H2), 7.10 (1H, d, J=2.4 Hz, H2), 7.21 (1H, dd, J= 2.1, 9.1 Hz, H8), 7.32 (1H, d, J=2.1 Hz, H10), 8.33 (1H, d, J=9.1 Hz, H5), 8.45 (1H, s, H6), 8.46 (1H, d, J=9.1 Hz, H7), 9.05 (1H, bs, OH), 13.50 (1H, bs, NH).

3-hydroxy-5*H***-7-fluoro-isoindole[2,1***a***]quinoxalin-6-one 5q:** m.p. 362-363°C; yield 87%, IR: 3265 (OH), 3180 (NH), 1642 (CO) cm⁻¹; ¹H NMR DMSO-d6 (ppm): 7.02 (1H, dd, J=2.1, 9.0 Hz, H4), 7.12 (1H, d, J=2.1 Hz, H2), 7.31-7.48 (2H, m, H8-H9), 7.84 (1H, d, J=7.2 Hz, H10), 8.38 (1H, d, J=9.0 Hz, H5), 9.35 (1H, d, J_F=1.9 Hz, H6), 10.69 (1H, bs, OH), 14.11 (1H, bs, NH).

Synthesis of diethyl pyridine-2,3-dicarboxilate 20

Pyridin 2,3- dicarboxilic acid (12 mmol, 2g) was suspended in methanol (8 ml). To the mixture, sulfuric acid conc. (0.8 ml) was added and it was refluxed for 24h. The solvent was evaporated under reduced pressure and to the resulting oil a saturated solution (20 ml) of NaHCO3 was added until neutralization. It was extracted in ethyl acetate, dried and evaporated under reduced pressure.

Dimethyl pyridine-2,3-dicarboxilate, 20: 55.0-56.0°C; yield 78%; IR 1734 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 3.88 (6H, s, 2xCH₃), 7.73 (1H, dd, J=4.8, 8.0 Hz, H-5), 8.32 (1H, d, J=8.0 Hz, H-4, 8.83 (1H, d, J=4.8 Hz, H-6); ¹³C NMR DMSO-d6 (ppm): 52.72 (q), 52.94 (q), 124.88 (s), 125.57 (d), 137.87 (d), 150.57 (s), 152.29 (d), 165.07 (s), 166.37 (s).

Synthesis of pyridine-2,3-dicarbaldehyde 16

To a stirred solution of compound 20 (9.7 mmol, 1.9 g) in toluene (68 ml), cooled at - 78°C and under nitrogen atmosphere, DIBAL 1M solution (26 ml) was added drop wise. After 30 min a solution of diethyl ether and acetic acid (10 ml, 1/1) was added and the mixture was stirred for 15 min more at -78°C and then warmed to rt in 1h. The resulting precipitate was filtered and suspended in toluene and filtered again (5 times). The organic layers were evaporated under reduced pressure. It was crystallized in ethanol.

Pyridine-2,3-dicarbaldehyde 16: Yield 60%; IR 1697, 1710 cm⁻¹; ¹H NMR DMSO (ppm): 7.46-9.01 (3H, m, H-4, H-5, H-6), 10.19 (1H, s, CHO); 10.59 (1H, s, CHO). ¹³C NMR DMSO-d6 (ppm): 127.78 (d), 135.93 (d), 153.27 (d), 191.63 (d), 194.39 (d).

Synthesis of 1,4-dimethoxy-2,3-dimethylbenzene 21

To a stirred solution of 2,3-dimethyl-hydroquinone (3.6 mmol, 0.5 g) in DMF (6 ml), Cs_2CO_3 (10.0 mmol, 3.3 g) and methylen iodide (10 ml) were added. The mixture was stirred at rt overnight. It was poured in water and ice, extracted in DCM, dried and evaporated.

1,4-dimethoxy-2,3-dimethylbenzene, 21: m.p. 78.0°C; yield 70%; ¹H NMR DMSO (ppm): 2.07 (6H, s, CH₃), 3.70 (6H, s, OCH₃), 6.71 (2H, s, H-5, H-6); ¹³C NMR

DMSO-d6 (ppm): 11.83 (q), 55.58 (q), 107.91 (d).

Synthesis of 1,4-dimethoxy-2,3-dimethyl-5,6-dinitrobenzene 22

To a stirred solution of compound **21** (3 mmol, 0.5 g) in acetic anhydride (4ml), cooled at 0°C, nitric acid 90% (0.7 ml) was added drop wise. The mixture was warmed at 90°C for 1h and then cooled to rt and poured onto water and ice. The resulting precipitate was filtered off.

1,4-dimethoxy-2,3-dimethyl-5,6-dinitrobenzene, 22: m.p. 155.0°C; yield 80%; ¹H NMR DMSO (ppm): 2.31 (6H, s, CH₃), 3.84 (6H, s, OCH₃); ¹³C NMR DMSO-d6 (ppm): 13.40 (q), 63.17 (q), 139.14 (s), 146.00 (s).

Synthesis of 3,6-dimethoxy-4,5-dimethyl-1,2-diamine, 2i

Method A: to a mixture of Sn (1.9 g) in HCl conc. compound **22** (2 mmol, 0.51 g) was added and it was stirred at rt for 72h. It was poured onto ice and extracted in ethyl acetate. It was dried, evaporated and purified in column using ethyl acetate as eluent. Yield 30%

Method B: to a solution of compound 22 (3.7 mmol, 0.95g) in methanol (20 ml) a catalytic amount of 10% Pd/C is added to the mixture that is hydrogenated on Paar for 24h at rt. The catalyst is filtered off and the filtrate evaporated under reduced pressure. Yield 96%

3,6-dimethoxy-4,5-dimethyl-1,2-diamine, 2i: mp. 114.0°C; IR 3410, 3332 (NH₂) cm⁻¹; ¹H NMR DMSO (ppm): 2.01 (3H, s, CH₃), 3.56 (3H, s, OCH₃), 4.17 (4H, bs, NH₂); ¹³C NMR DMSO-d6 (ppm): 11.67 (q), 58.72 (q), 116.45 (s), 125.84 (s), 141.68 (s).

<u>Synthesis of 1,4-dimethoxy-2,5-dinitrobenzene and 1,4-dimethoxy-2,3-</u> <u>dinitrobenzene, 15a-b</u>

To a stirring solution of 1,4-dimethoxybenzene (10 mmol, 1.38 g) in acetic anhydride (10 ml) cooled at -10°C, a mixture of nitric acid and acetic anhydride (8.8 ml, 1/1) previously cooled at -15°C was added drop wise. The resulting yellow precipitate was filtered off and purified in column cycloexane-ethyl acetate 9/1,

1,4-dimethoxy-2,5-dinitrobenzene, 15a: m.p. 189.0-190.0°C; ¹H NMR DMSO (ppm): 3.95 (6H, s, 2xCH₃), 7.71 (1H, s, H-3, H-6); ¹³C NMR DMSO-d6 (ppm): 57.80 (q), 119.11 (d), 144.80 (s), 132.65 (s).

1,4-dimethoxy-2,3-dinitrobenzene, 15b: m.p. 210.0-211.0°C; ¹H NMR DMSO (ppm): 3.92 (6H, s, 2xCH₃), 7.95 (1H, s, H-5, H-6); ¹³C NMR DMSO-d6 (ppm): 57.58 (q), 110.93 (d), 141.42 (s), 142.75 (s).

Synthesis of 6-(2-amino)-6H-pyrrolo[3,4-b]pyridine-4-carbonitrile 17:

To a solution of sodium hydrogen sulfite (1.56 g, 0.015 mol) in water (3.8 ml), pyridine-2,3-dicarbaldehyde **16** (0.015 mol) was added. The mixture was stirred until the solid was dissolved, and the appropriate 1,2-phenylenediamine **2a**, **2c-i** (0.015 mol) was added. The reaction was heated on a steam bath for 1h, then KCN (3.39 g, 0.052 mol) in water (8.0 ml) was added and the mixture was heated for additional 5h. The solid formed upon cooling was filtered and purified by chromatography using DCM-ethyl acetate 9/1 or 8/2.In the case of derivatives **17c** and **17d**, the 1,2-phenylenediamine **2e** was first dissolved in DMF (15 ml) and then added to the reaction mixture. In the case of derivative **17m** the reaction was performed at rt during all reaction time.

6-(2-amino-4,5-dimethylphenyl)-6H-pyrrolo[3.4-b]pyridine-5-carbonitrile, 17a: m.p. 176.5-177.6°C; yield 10%; IR 3558, 3386 (NH₂), 2212 (CN) cm⁻¹; ¹H NMR DMSO (ppm): 2.13 (3H, s, CH₃), 2.19 (3H, s , CH₃), 4.95 (2H, bs, NH₂), 6.73 (1H, s, H-3'), 6.97 (1H, s, H-6'), 7.18 (1H, dd, J=3.7, 8.1 Hz, H-6), 7.84 (1H, s, H₃), 8.22 (1H, d, J= 8.1 Hz, H-7), 8.65 (1H, d, J= 2.2 Hz, H-5);

¹³C NMR DMSO-d6 (ppm): 18.03 (q), 19.34 (q), 95.18 (s), 113.35 (s), 117.16 (s), 117.31 (d), 118.36 (d), 120.30 (s), 121.27 (d), 123.68 (s), 128.03 (d), 130.21 (d), 138.95 (s), 141.61 (s), 145.79 (s), 150.81 (d).

6-(2-amino-4,5-dimethylphenyl)-6H-pyrrolo[3.4-b]pyridine-7-carbonitrile, 17b: m.p. 136.0-137.0°C; yield 10%; IR 3556, 3465 (NH₂), 2208 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 2.13 (3H, s, CH₃), 2.19 (3H, s, CH₃), 4.96 (2H, bs, NH₂), 6.72 (1H, s, H.3'), 6.97 (1H, s, H-6'), 7.29 (1H, dd, J=4.1, 8.6 Hz, H-5), 8.04 (1H, s, H-3), 8.16 (1H, d, J=8.5 Hz, H-4), 8.59 (1H, dd, J= 1.5, 4.1 Hz, H-6);

¹³C NMR DMSO-d6 (ppm): 17.99 (q), 19.31 (q), 92.93 (s), 113.26 (s), 117.23 (d),

120.16 (s), 120.58 (d), 122.49 (d), 123.57 (d), 124.11 (s), 126.19 (d), 127.99 (d), 138.93 (s), 139.93 (s), 141.58 (s), 149.43 (d).

6-(2-amino-4,5-dichlorophenyl)-6H-pyrrolo[3,4-b]pyridine-5-carbonitrile, 17c: m.p. 194.5-195.3 °C; yield 10%; IR 3558, 3480 (NH₂), 2210 (CN) cm⁻¹; ¹H NMR DMSO (ppm): 5.78 (2H, bs, NH₂), 7.12 (1H, s, H-3), 7.21 (1H, dd, J=4.0, 8.4 Hz, H-6), 7.64 (1H, s, H6²), 7.96 (1H, s, H-3), 8.25 (1H, d, J=8.4 Hz, H-7), 8.68 (1H, d, J=3.8 Hz, H-5);

¹³C NMR DMSO-d6 (ppm): 95.19 (s), 113.11 (s), 115.84 (s), 116.28 (d), 117.38 (s), 118.66 (d), 121.58 (d + s), 129.65 (d), 130.46 (d), 133.15 (s), 144.75 (s), 145.82 (s), 151.29 (d).

6-(2-amino-4,5-dichlorophenyl)-6H-pyrrolo[3,4-b]pyridine-7-carbonitrile, 17d: m.p. 348.6-350.4°C; yield 10%; IR 3396, 3330 (NH₂), 2206 (CN) cm⁻¹; ¹H NMR DMSO (ppm): 5.77 (1H, bs, NH2), 7.12 (1H, s, H-3'), 7.32 (1H, dd, J=4.0, 8.7 Hz, H-5), 7.64 (1H, s, H-6'), 8.17-8.21 (2H, m, H-3, H-4), 8.61 (1H, d, J=4, H-6);

¹³C NMR DMSO-d6 (ppm): 92.96 (s), 112.99 (s), 115.74 (s), 116.20 (d), 121.47 (s), 122.78 (d), 124.35 (s), 126.34 (d), 129.60 (d), 133.07 (s), 137.19 (s), 144.69 (s), 148.77 (d).

6-(2-amino-3,6-dimethoxyphenyl)-6H-pyrrolo[3,4-b]pyridine-5-carbonitrile, 17e: m.p. 214.2-214.7°C; yield 12%; IR 3284, 3189 (NH₂), 2214 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 3.62 (3H, s, OCH₃), 3.82 (3H, s, OCH₃), 4.84 (2H, bs, NH₂), 6.35 (1H, d, J=8.9 Hz, H-4'), 6.96 (1H, d, J=8.9 Hz, H-5'), 7.16 (1H, dd, J=4.1, 8.5 Hz, H-6), 7.77 (1H, s, H-3), 8.21 (1H, dd, J= 1.5, 8.5 Hz, H-7), 8.64 (1H, dd. J= 1.6, 4.2 Hz, H-5);

¹³C NMR DMSO-d6 (ppm): 55.72 (q), 56.06 (q), 95.39 (s), 97.05 (d), 111.54 (d), 111.65 (s), 113.28 (s), 117.31 (s), 118.21 (d), 121.66 (d), 130.32 (d), 135.73 (s), 141.10 (s), 145.61 (s), 149.31 (s), 150.81 (d).

6-(2-amino-3,6-dimethoxyphenyl)-6H-pyrrolo[3,4-b]pyridine-5-carbonitrile, 17f: m.p. 196.8-198.4°C; yield 15%; IR 3486, 3392 (NH₂), 2206 (CN) cm⁻¹; ¹H NMR DMSO (ppm):3.62 (3H, s, OCH₃), 3.82 (3H, s, OCH₃), 4.85 (2H, bs, NH₂), 6.36 (1H, d, J=8.9 Hz, H-4'), 6.96 (1H, d, J=9.1 Hz, H-5'), 7.28 (1H, dd, J=4.1, 8.6 Hz, H-5), 7.98 (1H, s, H-3), 8.15 (1H, dt, J=9.5 Hz, H-4), 8.57 (1H, dd, J=1.5, 4.1 Hz, H-6);

¹³C NMR DMSO-d6 (ppm): 55.73 (q), 56.05 (q), 93.28 (s), 97.07 (d), 99.50 (s), 111.54 (d), 113.25 (s), 120.59 (d), 122.88 (d), 121.10 (s), 126.34 (d), 135.74 (s), 139.25 (s), 141.10 (s), 148.39 (d), 149.27 (s).

6-(2-amino-4,5-dimethoxyphenyl)-6H-pyrrolo[3,4-b]pyridine-5-carbonitrile, 17g: m.p. 178.3-179.2°C; yield 15%; IR: 3218,3116 (NH₂), 2208 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 3.65 (3H, s, OCH₃), 3.78 (3H, s, OCH₃), 4.85 (2H, bs, NH₂). 6.58 (1H, s, H-3'), 6.89 (1H, s, H-6'), 7.18 (1H, dd, J= 4.17, 8.5 Hz, H-6), 7.86 (1H, s, H-3), 8.22 (1H, dd, J=1.55, 8.5 Hz, H-7), 8.65 (1H, dd, J=4.54, 4.15 Hz, H-5);

¹³C NMR DMSO-d6 (ppm): 55.39 (q), 56.41 (q), 95.53 (s), 100.23 (d), 112.49 (d), 113.43 (s), 113.88 (s), 117.18 (s), 118.35 (d), 121.61 (d), 130.20 (d), 138.52 (s), 139.68 (s), 145.70 (s), 150.82 (d), 150.97 (s).

6-(2-amino-4,5-dimethoxyphenyl)-6H-pyrrolo[3,4-b]pyridine-7-carbonitrile, 17h: m.p. 216.2-217.3°C; yield 15%; IR 3320, 3207 (NH2), 2202 (CN) cm⁻¹; ¹H NMR DMSO (ppm): 3.65 (3H, s, OCH₃), 3.76 (3H, s, OCH₃), 4.85 (2H, bs, NH₂), 6.57 (1H, s, H-3'), 6.89 (1H, s, H-6'), 7.29 (1H, dd, J= 4.1, 8.6 Hz, H-5), 8.05 (1H, s, H-3), 8.14 (1H, d, J=8.5 Hz, H-4), 8.6 (1H, dd, J=1.3, 4.02 Hz, H-6);

¹³C NMR DMSO-d6 (ppm): 55.40 (q), 56.40 (q), 95.35 (s), 100.25 (d), 112.50 (d), 113.38 (s), 113.85 (s), 120.64 (d), 122.84 (d), 124.11 (s), 124.21 (d), 138.50 (s), 139.13 (s), 139.69 (s), 148.49 (d), 150.99 (s).

6-(2-amino-3,6-dimethoxy-4,5-dimethylphenyl)-6H-pyrrolo[3,4-b]pyridine-5-

carbonitrile, 17i: m.p. 186.0-187.0°C, yield 11%; IR 3457, 3436 (NH₂), 2212 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 2.08 (3H, s, CH₃), 2.19 (3H, s, CH₃), 3.28 (3H, s, OCH₃), 3.65 (3H, s, OCH₃), 4.81 (2H, s, NH₂), 7.19 (1H, dd, J=4.1, 7.5 Hz, H-6), 7.88 (1H, s, H-3), 8.24 (1H, dd, J=1.6, 8.6 Hz, H-7), 8.66 (1H, dd, J=1.6, 4.2 Hz, H-5);

¹³C NMR DMSO-d6 (ppm):11.79 (q), 11.64 (q), 61.07 (q), 95.66 (s), 113.33 (s), 115.08 (s), 115.96 (s), 117.17 (s), 118.35 (d), 122.19 (d), 130.39 (d), 132.10 (s), 136.49 (s), 141.16 (s), 145.64 (s), 149.87 (s), 150.95 (d).

6-(2-amino-3,6-dimethoxy-4,5-dimethylphenyl)-6H-pyrrolo[3,4-b]pyridine-7-

carbonitrile, 17j: m.p.180.0°C; yield 26 %; IR 3475, 3374 (NH₂), 2204 (CN) cm⁻¹; ¹H NMR DMSO (ppm): 2.08 (3H, s, CH₃), 2.19 (3H, s, CH₃), 3.28 (3H, s, OCH₃), 3.65 (3H, s, OCH₃), 4.80 (2H, s, NH₂), 7.30 (1H, dd, J=4.1, 8.6 Hz, H-5), 8.09 (1H, s, H-3), 8.18 (1H, d, J= 8.4 Hz, H-4), 8.60 (1H, dd, J=1.4, 4.1 Hz, H-6);

¹³C NMR DMSO-d6 (ppm):11.79 (q), 13.63 (q), 59.23 (q), 61.11 (q), 93.57 (s), 113.29 (s), 115.07 (s), 115.98 (s), 120.70 (d), 123.38 (d), 124.12 (s), 126.38 (d), 132.11 (s), 136.48 (s), 139.16 (s), 141.17 (s), 148.93 (d), 149.86 (s).

6-(2-aminophenyl)-6H-pyrrolo[3,4-b]pyridine-5-carbonitrile, 17k: m.p. 164.7-165.9°C; Yield 30%; IR 3423, 3311 (NH₂); 2200 (CN) cm⁻¹; ¹H NMR DMSO (ppm):5.28 (2H, bs, NH₂), 6.70 (1H, t, J=7.5 Hz, H-5'), 6.92 (1H, d. J=8.3 Hz, H-6), 7.19-7.23 (2H, m, H-3', H-6'), 7.27 (1H, t, J=7.5 Hz, H-7), 8.67 (1H, d, J=3.9 Hz, H-5); ¹³C NMR DMSO-d6 (ppm): 95.12 (s), 113.34 (s), 115.73 (d), 116.18 (d), 117.29 (s), 118.48 (d), 121.34 (d), 122.40 (s), 127.91 (d), 130.34 (d), 130.74 (d), 144.21 (s), 145.88 (s), 150.97 (d).

6-(2-aminophenyl)-6H-pyrrolo[3,4-b]pyridine-7-carbonitrile, 171: m.p. 180.3-181.4°C; yield 40%; IR 3478, 3392 (NH₂), 2208 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 5.28 (2H, s, NH2), 6.70 (1H, dt, J=1.2, 7.5 Hz, H5'), 6.92 (1H, dd, J=1.2, 8.5 Hz, H-3'), 7.19-7.34 (3H, m, H-4', H-6', H-5), 8.12 (1H, s, H-3), 8.15-8.20 (1H, dt, J= 1.3, 8.5 Hz, H-4), 8.6 (1H, dd, J=1.5, 4.1 Hz, H-6);

¹³C NMR DMSO-d6 (ppm): 92.84 (s), 113.18 (s), 115.60 (d), 116.07 (d), 120.66 (d), 122.26 (s), 122.50 (d), 124.20 (s), 126.24 (d), 127.81 (d), 130.64 (d), 139.20 (s), 144.11 (s), 148.50 (d).

6-(2-amino-4-methoxyphenyl)-6H-pyrrolo[3,4-b]pyridine-5-carbonitrile, 17m: m.p. 185.0-186°C; yield 35%; 3308, 3415 (NH₂), 2255 (CN) IR cm⁻¹ ¹H NMR DMSO (ppm): 3.76 (3H, s, OCH₃), 5.27 (2H, bs; NH₂), 6.27 (1H, dd, J=2.6, 8.7 Hz, H-4'), 6.45 (1H, d, J=2.6 Hz, H-2'), 7.12 (1H, d, J=8.7 Hz, H-5'). 7.20 (1H, t, J=4.2 Hz, H-6), 7.84 (1H, s, H-3), 8.22 (1H, dd, J=1.3, 8.5 Hz, H-7), 8.65 (1H, dd, J=1.4, 4.1 Hz, H-5);

¹³C NMR DMSO-d6 (ppm): 55.06 (q), 95.47 (s), 100.04 (d), 102.16 (d), 113.39 (s), 116.14 (s), 117.20 (s), 118.36 (d), 121.66 (d), 128.80 (d), 130.25 (d), 145.33 (s), 145.75 (s), 150.82 (d), 160.95 (s).

Synthesis of 7-azaisoindole[2,1-a]quinoxalin-6(5H)-one 19:

A solution of the intermediates 17a-m (3 mmol) in acetic acid (10 ml) was heated under reflux for 30 min. After cooling the mixture is poured onto ice and the resulting precipitate filtered off. It was crystallized in ethanol. In case of derivatives 17i and 17j the reflux was maintained for 1h and 30 min and 3h respectively. In case of derivatives 17e and 17f the reflux was maintained for 8h and 3h respectively.

2,3-dimethyl-7-azaisoindole[2,1-a]quinoxalin-6(5H)-one, 19a: m.p. 340.0-341.0°C; yield quant.; IR 3079 (NH), 1656 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 2.36 (3H, s,

CH₃), 2.39 (3H, s, CH₃); 7.49 (1H, dt, J=6.5, H-8), 8.37 (1H, s, NH), 8.52 (1H, dd, J= 1.2, 8.5 Hz, H-7), 8.87 (1H, dd, J= 1.2, 4,2 H-9), 9.14 (1H, s, H-6).

2,3-dimethyl-4-azaisoindole[2,1-a]quinoxalin-6(5H)-one, 19b: m.p. >410.0 °C; yield 40%; IR 3558 (NH), 1643 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 2.38 (3H, s, CH₃), 2.42 (3H, s, CH₃), 7.47-7.52 (2H, m, H-5, H-9), 8.45 (1H, s, H-2), 8.81 (1H, d, J= 3.42 Hz, H-10), 9.05 (1H, d, J=8.73 Hz, H-8), 9.39 (1H, s, H-6), 13.65 (1H, bs, NH).

2,3-dichloro-7-azaidoindole[2,1-a]quinoxalin-6(5H)-one, 19c: m.p. 322.0-323.0°C; yield 99%; IR 3343 (NH), 1664 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 7.51 (1H, dd, J=4.28, 8.56 Hz, H-8), 8.03 (1H, s, H-2), 8.54 (1H, dd, J=1.38, 10.0 Hz, H-7), 8.91 (1H, dd, J=1.38, 4.27 Hz, H-9), 9.01 (1H, s, H-5), 9.30 (1H, s, H-6), 13.60 (1H, bs, NH).

2,3-dichloro-4-azaidoindole[2,1-a]quinoxalin-6(5H)-one, 19d: mp. 348.0-349.0°C; yield 71%; IR 3089 (NH), 1650 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 7.30-7.36 (2H, m, H-5, H-8), 7.73 (1H, s, H-2), 8.72 (1H, d, J=4.3 Hz, H-7), 8.91 (1H, d, J=2.9 Hz, H-9), 9.25 (1H, s, H-6), 13.80 (1H, bs, NH).

2,3-dimethoxy-4-azaisoindole[2,1-a]quinoxalin-6(5H)-one, 19e: m.p. 284.0-285.0°C; yield 90%; IR 3143 (NH), 1646 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 3.92 (3H, s, OCH3), 4.00 (3H, s, OCH3), 7.30 (1H, s, H-2), 7.55 (1H, dd, J=4.0, 8.3 Hz, H-9), 8.15 (1H, s, H-5), 8.85 (1H, d, J=3.8 Hz, H-10), 9.10 (1H, d, J=8.8 Hz, H-8), 9.63 (1H, s, H-6), 13.70 (1H, bs, NH).

2,3-dimethoxy-7-azaisoindole[2,1-a]quinoxalin-6(5H)-one, 19f: m.p. 272.0-273.0°C; yield 80%; IR 3305 (NH), 1660 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 3.91 (3H, s, OCH3), 3.99 (3H, s, OCH3), 7.32 (1H, s, H-2), 7.50 (1H, dd, J=4.3, 8.5 Hz, H-8), 8.09 (1H, s, H-5), 8.53 (1H, dd, J=1.3, 8.5 Hz, H-7), 8.87 (1H, dd, J=1.3, 4.3 Hz, H-9), 9.26 (1H, s, H-6), 13.29 (1H, bs, NH).

1,4-dimethoxy-7-azaisoindole[2,1-a]quinoxalin-6(5H)-one, 19g: m.p. 287-288°C; yield 40%; IR 3018 (NH), 1627 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 3.87 (3H, s, OCH3), 4.07 (3H, s, OCH3), 6.93 (1H, d, J=9.0 Hz, H-3), 7.04 (1H, d, J=4.2, 8.5 Hz, H-8), 8.40 (1H, d, J=7.5 Hz, H-7), 8.72 (1H, dd, J=4.14, H-9), 9.25 (1H, s, H-6), 13.50 (1H, bs, NH).

1,4-dimethoxy-4-azaisoindole[2,1-a]quinoxalin-6(5H)-one, 19h: m.p. 268-269°C; yield 70%; IR 3288 (NH), 1646 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 4.01 (3H, s, OCH3), 4.13 (3H, s, OCH3), 7.19 (1H, d, J=9.1 Hz, H-3), 7.31 (1H, dd, J=4.1, 8.6 Hz, H-9), 8.86 (1H, d, J = 3.8 Hz, H-10), 9.03 (1H, d, J= 8.6 Hz, H-8), 13.65 (1H, bs, NH).

1,4-dimethoxy-2,3-dimethyl-7-azaisoindolo[2,1-a]quinoxalin-6(5H)-one, 19i: m.p. 285-286°C; yield 41%; IR 3324 (NH), 1658 (CN) cm⁻¹ ¹H NMR DMSO (ppm): 2.34 (3H, s, CH3), 2.36 (3H, s, CH3), 2.86 (3H, s, OCH3), 2.87 (2H, s, OCH3), 7.52 (1H, dd, J= 4.3, 8.5 Hz, H-9), 8.58 (1H, dd, J= 1.2, 8.5 Hz, H-10), 8.93 (1H, dd, J= 1.3, 4.3 Hz, H-8), 9.38 (1H, s, H-6), 12.47 (1H, bs, NH).

1,4-dimethoxy-2,3-dimethyl-4-azaisoindolo[2,1-a]quinoxalin-6(5H)-one, 19j: m.p. 246-247°C; yield 90%; IR 3018 (NH), 1710 (CN) cm⁻¹ ¹H NMR DMSO (ppm):2.35 (3H, s, CH3), 2.37 (3H, s, CH3), 3.85 (3H, s, OCH3), 3.88 (3H, s, OCH3), 7.54-7.61 (1H, m, H-8), 8.87 (1H, d, J=3.6 Hz, H-7), 9.03 (1H, d, J=8.7, H-9), 9.50 (1H, s, H-6), 12.70 (1H, bs, NH).

4-azaisoindole[2.1-a]quinoxalin-6(5H)-one, 19k: m.p. 330.0-331.0°C; yield 60%; IR 3322 (NH), 1656 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 7.53-7.79 (3H, m, H-2, H-3, H-4), 8.65-8.69 (2H, m, H-9, H-5), 8.87 (1H, d, J=3.7 Hz, H-10), 9.11 (1H, d, J=8.5 Hz, H-8), 9.61 (1H, s, H-6), 13.50 (1H, bs, NH).

7-azaisoindole[2.1-a]quinoxalin-6(5H)-one, 191: m.p. 322.0-323.0 °C; yield 80%; IR 3073(NH), 1664 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 7.51-7.71 (3H, m, H-3, H-4, H-8), 7.81 (1H, d, J=7.3 Hz, H-2), 8.53-8.62 (2H, m, H-5, H-7), 8.91 (1H, d, J=3.3 Hz, H-9), 9.29 (1H, s, H-6), 12.90 (1H, bs, NH).

3-methoxy-4-azaisoindole[2.1-a]quinoxalin-6(5H)-one, 19m: m.p. 267.0-268.0°C; yield 80%; IR 3382 (NH), 1659 (CO) cm⁻¹ ¹H NMR DMSO (ppm): 3.90 (3H, s, OCH3), 7.19 (1H, dd, J=2.6, 9.1 Hz, H-5), 7.29 (1H, d, J=2.6 Hz, H-2), 7.50 (1H, dd, J=4.3, 8.5 Hz, H-8), 8.52 (2H, d, J=9.5 Hz, H-5, H-7), 8.88 (1H, dd, J=1.2, 4.3 Hz, H-9), 9.20 (1H, s, H-6), 13.75 (1H, bs, NH).

Synthesis of 6-amino-3-methoxy-7-azaisoindole[2,1-a]quinoxaline, 18a

A solution of the intermediates 17 m (3 mmol) in acetic acid (10 ml) was heated under reflux for 30 min. After cooling the resulting precipitate filtered off. It was crystallized in ethanol. In case of derivatives 17i and 17j the reflux was maintained for 1h and 30 min and 3h respectively. In case of derivatives 17e and 17f the reflux was maintained for 8h and 3h respectively.

6-amino-3-methoxy-7-azaisoindole[2,1-a]quinoxaline, 18a: m.p. 238.0-239.0°C; yield 60%; IR 3399, 3000 (NH₂) cm⁻¹ ¹H NMR DMSO (ppm): 3.87 (3H, s, OCH₃),

6.96 (1H, dd, J=2.8, 8.9 Hz, H-5), 7.08 (1H, d, J=2.7 Hz, H-2), 7.30-7.36 (4H, m, H-8, H-4, NH2), 8.34 (1H, dd, J=2.1, 8.8 Hz, H-7), 8.68 (1H, dd, J=1.4, 4.2 Hz, H-9), 8.77 (1H, s, H-6).

¹³C NMR DMSO-d6 (ppm): 55.36 (q), 104.47 (d), 106.33 (s), 108.01 (d), 110.71 (d), 116.72 (d), 117.61 (s), 118.47 (s), 118.73 (d), 128.03 (d), 135.75 (s), 140.79 (d), 147.25 (s), 151.59 (s), 158.75 (s).

Synthesis of 2-(2-aminopyridin-3-yl)-2H-isoindole-1-carbonitrile 23:

To a solution of sodium hydrogen sulfite (1.56 g, 0.015 mol) in water (3.8 ml), phtalaldehyde **1a**, **1c** or **1e** (0.015 mol). The mixture was stirred until the solid was dissolved, and 2,3-diaminopyridine (0.015 mol, 1.64 g) was added. The reaction was heated on a steam bath for 1h, then KCN (3.39 g, 0.052 mol) in water (8.0 ml) was added and the mixture was heated for additional 6h and then cooled to rt it was stirred at rt overnight. The solid formed upon cooling was filtered and purified by chromatography using DCM-ethyl acetate 9/1 as eluent. In the case of derivative 2**3b**, it was purified in column using DCM-ethyl acetate 84/16 as eluent.

2-(2-aminopyridin-3-yl)-2H-isoindole-1-carbonitrile, 23a: : m.p. 284.0-285.0°C ;yield 38%; IR 3500, 3397 (NH2), 2200 (CN); 1H NMR DMSO (ppm): 6.16 (2H, s, NH2), 6.74 (1H, dd, J=4.9,7.6, H-5'), 7.17 (1H, t, J=6.8 Hz, H-3), 7.33 (1H, t, J=6.8, H-4), 7.59-7.69 (2H, m, H-3, H-5), 7.79 (1H, d, J=8.5 Hz, H-4'), 7.94 (1H, s, H-6), 8.16 (1H, dd, J=1.7, 4.9 Hz, H-6'); ¹³C NMR DMSO-d6 (ppm): 111.91 (d), 113.96 (s), 117.58 (d), 117.83 (s), 121.62 (d), 122.47 (d), 122.55 (d), 124.30 (s), 125.78 (d), 131.38 (s), 136.23 (d), 149.76 (d), 155.13 (s).

2-(2-aminopyridin-3-yl)-5,6-dimethyl-2H-isoindole-1-carbonitrile, 23b: m.p. 175.0-176.0°C; yield 15%; IR 3381, 3313 (NH₂), 2198 (CN) cm⁻¹; ¹H NMR DMSO (ppm): 6.07 (2H, bs, NH₂), 6.72 (1H, dd, J=4.9, 7.6 Hz, H-5'), 7.42 (1H, s, H-4), 7.51 (1H, s, H-7), 7.57 (1H, dd, J=1.7, 7.6 Hz, H-4'), 7.74 (1H, s, H-3), 8.12 (1H, dd, J=1.7, 4.9 Hz, H-6');

3C NMR DMSO-d6 (ppm): 20.02 (q), 20.33 (q), 99.50 (s), 111.92 (d), 114.31 (s), 116.39 (d), 117.96 (s), 120.07 (d), 121.35 (d), 123.71 (s), 131.05 (s), 132.17 (s), 136.00 (s), 136.12 (d), 149.55 (d), 155.19 (s).

70

2-(2-aminopyridin-3-yl)-5,6-dimethoxy-2H-isoindole-1-carbonitrile, 23c: m.p. >410°C; yield 10%; IR 3323, 3321 (NH₂), 2256 (CN) cm-1; ¹H NMR DMSO (ppm): 3.80 (3H, s, OCH₃), 3.87 (3H, s, OCH₃), 6.06 (2H, bs, NH₂), 6.71 (1H, dd, J=4.9, 7.6 Hz, H-4'), 7.64 (1H, s, H-3), 8.13 (1H, dd, J=1.6, 4.8 Hz, H-6');

¹³C NMR DMSO-d6 (ppm): 55.37 (q), 55.61 (q), 99.29 (d), 99.55 (d), 112.02 (d), 114.55 (s), 118.13 (s), 119.55 (s), 121.10 (d), 127.86 (s), 136.19 (d), 148.23 (s), 149.40 (d), 151.07 (s), 155.38 (s).

Synthesis of pyrido[2',3':5,6]pyrazino[2,1-a]isoindol-6(5H)-one 25

To a solution of the intermediates 23a-c (2.1 mmol) in acetic acid (7.5 ml) DMF (1ml) was added. The mixture was heated under reflux for 3h. After cooling the mixture is poured onto ice and the resulting precipitate filtered off. It was crystallized in ethanol.

Pyrido[2',3':5,6]pyrazino[2,1-a]isoindol-6(5H)-one, 25a: m.p. 352.5-354.0°C; yield quant.; IR 3079 (NH), 1635 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 7.43-7.66 (3H, m, H-5', H-4, H-5), 8.04 (1H, d, J=8.2, H-3, H-6), 8.60 (1H, m, H-4'), 9.02 (1H, d, J=7.1 Hz, H-6'), 9.42 (1H, s, H-2), 13.30 (1H, bs, NH);

8,6-dimethylpyrido[2',3':5,6]pyrazino[2,1-a]isoindol-6(5H)-one, 25b m.p. >410°C; yield 95%; IR 3092 (NH), 1634 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 2.46 (3H, s, CH₃), 2.50 (3H, s, CH₃), 7.75 (1H, s, H-2), 7.58 (1H, m, H-5'), 8.37 (1H, s, H-6), 8.60 (1H, d, J=5.3 Hz, H-4'), 8.95 (1H, d, J=7.7 Hz, H-6'), 9.21 (1H, s, H-2), 13.10 (1H, bs, NH);

8,6-dimethoxypyrido[2',3':5,6]pyrazino[2,1-a]isoindol-6(5H)-one, 25c: m.p. 275.5-277.0°C; yield quant.; IR 3350 (NH), 1630 (CO) cm⁻¹; ¹H NMR DMSO (ppm): 3.90 (3H, s, OCH₃), 3.99 (3H, s, OCH₃), 7.31 (1H, s, H-6), 7.56 (1H, dd, J=4.9, 8.2 Hz, H-5'), 7.81 (1H, s, H-3), 8.52 (1H, d, J=5.1 Hz, H-4'), 8.90 (1H, d, J=7.4 Hz, H-6'), 9.13 (1H, s, H-2) 13.25 (1H, bs, NH);

Synthesis of 5,6-dihydropyrido[2',3':5,6]pyrazino[2,1-a]isoindol-6-amine, 24

To a solution of the intermediates 23a-c (2.1 mmol) in acetic acid (7.5 ml) DMF (1ml) was added. The mixture was heated under reflux for 3h. After cooling the solvent was evaporated under reduced pressure and it was crystallized in ethanol.

5,6-dihydropyrido[**2',3':5,6]pyrazino**[**2,1-a]isoindol-6-amine**, **24a**: m.p. 306.2-307.1°C; yield 70%; IR 3405, 3493 (NH₂) cm⁻¹; ¹H NMR DMSO (ppm): 7.27-7.35 (5H, m, NH₂, H-3, H-4, H-6), 7.87-7.91 (1H, m, H-5), 8.46-8.51 (1H, m, H-5'), 8.58 (1H, dd, J= 1.6, 5.0 Hz, H-4'), 8.78 (1H, dd, J=1.6, 8.1 Hz, H-6'), 9.01 (1H, s, H-2).

¹³C NMR DMSO-d6 (ppm): 99.55 (s), 107.62 (s), 109.43 (d), 117.09 (d), 119.10 (d), 119.63 (d), 119.74 (s), 120.12 (d), 123.37 (d), 123.84 (d), 126.16 (s), 148.27 (d), 150.00 (s), 153.59 (s).

8,6-dimethyl-5,6-dihydropyrido[2',3':5,6]pyrazino[2,1-a]isoindol-6-amine, 24b: m.p. >410.0°C; yield 70%; IR 3423, 3320 (NH₂) cm⁻¹; ¹H NMR DMSO (ppm): 7017 (2H, bs, NH₂), 7.26 (1H, m, H-5'), 7.61 (1H, s, H-3), 8.27 (1H, s, H-6), 8.53 (1H, dd, J=1.5, 4.6 Hz, H-4'), 8.70 (1H, dd, J=1.5, 8.1 Hz, H-6'), 8.82 (1H, s, H-2);

¹³C NMR DMSO-d6 (ppm): 20.22 (q), 20.40 (q), 106.80 (d), 107.38 (s), 108.48 (d), 116.84 (d), 118.38 (d), 118.91 (d), 119.22 (s), 119.26 (s), 123.42 (d), 125.65 (s), 132.94 (s), 133.30 (s), 147.87 (s), 153.43 (s).

8,6-dimethoxy-5,6-dihydropyrido[2',3':5,6]pyrazino[2,1-a]isoindol-6-amine, 24c: m.p. 233.0 234.0°C; yield 80%; IR 3380, 3445 (NH₂) cm⁻¹; ¹H NMR DMSO (ppm): 3.87 (3H, s, OCH₃), 3.95 (3H, s, OCH₃), 7.16-7.26 (4H, m, NH₂, H-6, H-5'), 7.67 (1H, s, H-3), 8.48 (1H, dd, J=7.6, 4.6 Hz, H-4'), 8.62 (1H, dd, J=1.6, 8.1, H-6'), 8.72 (1H, s, H-2);

¹³C NMR DMSO-d6 (ppm): 55.19 (q), 55.86 (q), 98.80 (d), 99.43 (d), 107.28 (s), 108.28 (d), 115.29 (s), 116.17 (d), 119.42 (s), 121.95 (s), 122.64 (d), 147.01 (d), 148.57 (s), 149.03 (s), 149.69 (s), 153.32 (s).

Molecular modeling

These studies were carried out using GLIDE program from Schrodinger software.

Docking on tubuline

The structure PDB 1SA0 was used for tubulin docking in several works present in literature.

As ISQO compounds inhibit tubulin polymerization in a way similar to colchicine this X-ray structure was used for docking.

The protein was prepared using Protein Preparation Wizard. The stathmin-like domain and the C and D subunits, Mg ions, were removed.

Prime refinement was used to complete residue side chains with missing atoms.

Docking was carried out using Glide program.

To validate the Glide docking protocol, colchicine, the co-crystallized ligand, was docked into the active site. The docked structure was compared with the crystal structure showing that this protocol successfully reproduces the crystal tubulin-colchicine complex.

Docking studies on Topoisomerase 1

The X-ray structure with PDB code 1K4T was used for docking.

The protein was prepared using Protein Preparation Wizard. The TTC coligand, Hg ions and water molecules with a distance longer than 5 A were removed.

Glide program was used to carry out the docking. To validate the Glide protocol, Topotecan (TTG), the co-crystallized coligand, was docked into the active site. The docked structure was compared with the crystal structure showing that this protocol successfully reproduces the crystal tubulin- colchicine complex.

Synthesis of Ventilatone A in collaboration with The University of Nottingham (May-September 2011)

Ventilatone A was isolated from the root bark of Ventilago calyculata in 1985 and belongs to the pyranonaphthoquinone family of natural products [91JCS327], [85P2669]. Naphtho[2,3-c]pyran-5,10-dione is the most common ring system arrangement for pyranonaphthoquinones, however the naphtha[2,3-c]pyran-6,9-dione systems are also known. This configuration is much less common in nature and only a handful of natural products of this type have been synthesized [99NPR267], [00T1937], [08NPR376].

Ventilatone A belongs to this second group and no synthesis of it has yet been reported. Its configuration in not very common in nature and only a handful of natural products of this type have been synthesized [90JOC1466], [92AJC2025], [04THL939], [07THL1545]. An optical rotation is given for it but the absolute configuration at its stereogenic center is not known.

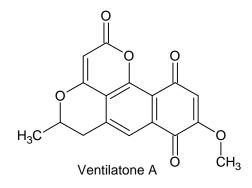


Fig.30

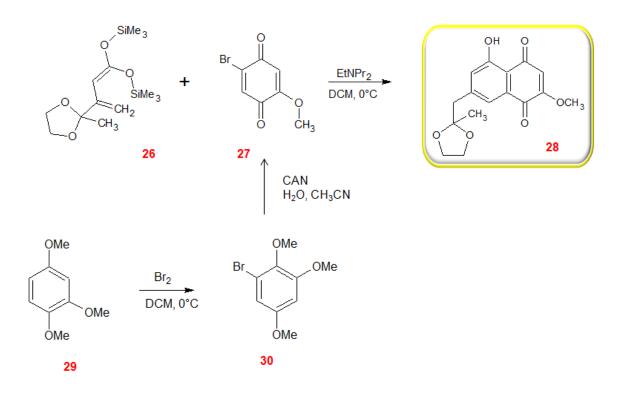
Results and discussion

To synthesize Ventilatone A whose structure is shown in fig.30, we imagine to break down the molecule in order to obtain the smaller unit easier to synthesize.

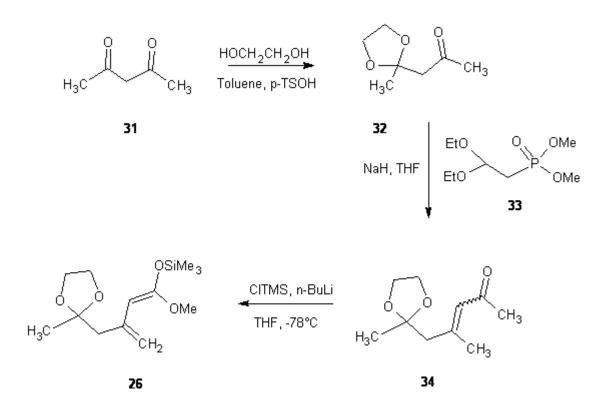
We planned to start our Ventilatone A synthesis from the naphtoquinone intermediate **28**.

Diels-Alder reaction between benzoquinone **27** and diene **26** can give the intermediate **28**. Benzoquinone **27** was obtained by bromination of trimethoxybenzene **29** and then subsequent oxidation of 1-bromo-2,4,5-trimethoxybenzene **30** with cerium ammonium nitrate (CAN).

The position of the bromine in the benzoquinone dienophile dictates the regiochemistry of the halogen directing Diels-Alder reaction with diene **26**.



The diene **26** was synthesized starting from the diketone **31** in which one carbonyl function was protected to give compound **32**. Reaction of this latter with methyl diethylphosphonoacetate **33** afforded compound **34** in a mixture of the two isomers that was directly used for the sequent step with chloro-trimethylsilane and n-BuLi.



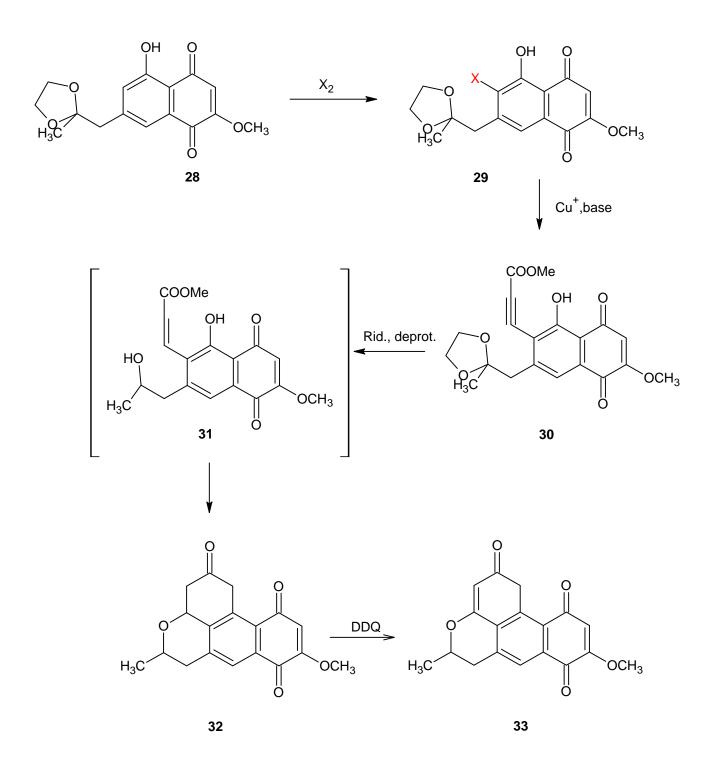
Once we synthesized the naphoquinone intermediate we planned the synthetic route for Ventilatone A starting from this compound.

What we though to do was first try the introduction of an halogen atom into the orto phenolic position to obtain compound **29** that can be useful for Sanogashima coupling and to obtain compound **30**.

Reduction of the triple bond and of the aldehyde function, and then deprotection of the ketone give compound **31** that probably will evolve to compound **32** spontaneously. Finally, oxidation of compound **32** with DDQ allows to complete the synthesis of

Ventilatone A.

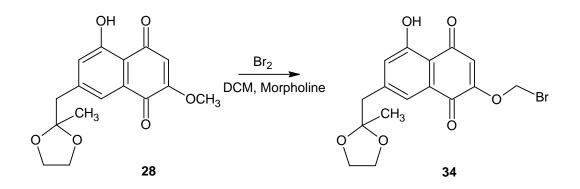
We start to find the best method to introduce the halogen atom on the orto-phenolic position of naphtoquinone intermediate.

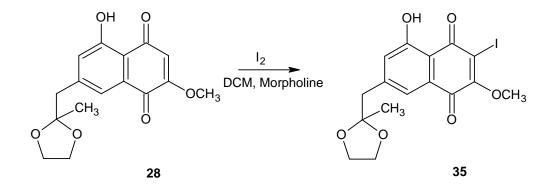


Our first attempt was the halogenation with NBS or NIS with different ratio equivalents and in the presence of diisopropylamine. From this reaction was possible isolate only starting material.

The iodination using I₂ and HIO₃ in water and CH₃CN gave starting material.

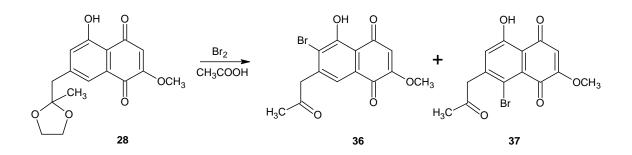
The halogenation of compound **28** in DCM and morpholine in presence of Br_2 gave compound **34**; instead when we used I_2 was isolated the corresponding iodinated compound in good yield but not on quinine portion.





Then we try halogenation in acetic acid with Br_2 and I_2 .

In case of Br_2 , using 1:1 ratio of quivalentes ($Br_2:28$), we collected just starting material. Using a ratio 2:1 ($Br_2:28$) we obtained a mixture of two isomers 36 and 37 from which was very difficult to isolate the pure compoundsify.



Using I_2 we were finally able to obtain the desired compound although the yield should be improved.

Actually this project is still in process in Nottingham.

EXPERIMENTAL DATA SYNTHESIS VENTILATONE A

Synthesis of 1-bromo-2,4,5-trimethoxybenzene 30

Bromine (3,2 ml, 63.1 mmol) in DCM (50 ml) was added dropwise over 1h to a solution of 1,2,4-trimethoxybenzene (10.1 g, 60.1 mmol) in DCM (200 ml) at 0°C. After addition had completed the resulting brown solution was stirred for 5 min then washed with saturated aqueous sodium thiosulfate solution (200ml), saturated aqueous sodium hydrogen carbonate solution (100 ml), water (100 ml) and brine (100 ml). The organic phase was then dried and concentrated .

Yield 75%; mp 52-53°C; IR 3011, 2938, 2843, 1586, 1506, 1465, 1438, 1379, 1166, 1030; CDCl₃ HNMR 7.04 (1H, s, ArH), 6.56 (1H, s, ArH), 3.89 (3H, s, OCH₃), 3.87 (3H, s, OCH₃), 3.83 (3H, s, OCH₃); CNMR 150.3 (C), 149.1 (C), 143.8 (C), 116.4 (CH), 101.1 (C), 98.9 (CH), 57.2 (CH₃), 56.6 (CH₃)

Synthesis of 2-bromo-5-methoxy-1,4-benzoquinone 27

CAN (83.2 g, 151.8 mmol) in water (200 ml) was added to 1-bromo-2,4,5trimethoxybenzene (15 g, 60.7 mmol) in aceto nitrile (200 ml). The dark brown solution was stirred in an open flask at rt for 1h, becoming bright orange. The reaction mixture was diluted with water (500 ml) and filtered to give a yellow solid compound. Yield 90%; IR 3045, 2941, 2850, 1680, 1661, 1624, 1588, 1192, 1171, 1006; CDCl₃ HNMR 7.25 (1H, s, ArH), 6.13 (1H, s, OCH3); CNMR 179.2 (C), 160.0 (C), 139.2 (C), 135.8 (CH), 106.9 (CH), 56.7 (CH₃)

Synthesis of acetylacetone-2,2-(ethylene glycol)monoketal 32

P-toluenesulfonic acid (11 mg, 0.006 mmol) was added to a solution of acetylacetone (5g, 50 mmol) and ethylene glicol (3.15 g, 50mmol) in toluene. The mixture was warmed to reflux overnight with azeotropic removal of water using a Dean-Stark trap. Toluene was removed under reduced pressure and the remaining crude was distillated

twice under reduced pressure.

Yield 70%; bp 115-118 °C; IR CDCl₃ 2989, 2888, 1709, 1790, 1361, 1241, 1054 cm⁻¹; CDCl₃ HMNR 3.97-3.94 (4H, m, -OCH₂), 2.75 (2H, s, CH₂), 2.20 (3H, s, OCH₃), 1.39 (3H, s, CH₃); CNMR 205.9 (C), 107.8 (C), 64.6 (CH₂), 64.2(CH₂), 52.5 (CH₂), 31.6 (CH₃), 24.3 (CH₃)

Synthesis of 3-methyl-4-(2-methyl-1,3-dioxolan-2-yl)but-2-enoate 34

Sodium hydrade (60% in mineral oil; 8.32 g, 208.1 mmol) was washed with dry npentane (30 ml), dried under vacuum, suspended in THF (250 ml) and cooled to 0°C.

Methyl diethylphosphonoacetate (38.2 ml, 208.1 mmol) was added dropwise and the reaction mixture was warmed to reflux over 30 min. Acetylacetone-2,2-(ethylene glycol)monoketal (20g,138.7 mmol) in THF was added dropwise and the suspension stirred for 6 h then cooled to rt. Ether and brine were added and the aqueous phase was extracted with ether three times. The combined ethereal extracts were washed with water and dried under reduced pressure.

The crude product was purified in column eluting with petroleum ether-ethyl acetate 8:2 Yield 65%; IR 2989, 2952, 2888, 1712, 1646, 1437, 1380, 1155, 1055 cm⁻¹; CDCl₃ HNMR 5.77 (1H, d, J 0.7, CH), 3.93-3.91 (4H, m, OCH₂OCH₂), 3.69 (3H, s, OCH₃), 2.45 (2H, s, CH₂), 2.07 (3H, s, CH=C-CH₃), 1.35 (3H, s, OCH₃); CNMR 166.8 (C), 154.9 (C), 118.8 (CH), 109.9 (C), 64.5 (CH₂), 50.8 (CH₃), 41.1 (CH₂), 26.5 (CH₃), 24.0 (CH₃)

<u>Synthesis of 1-methoxy-3-[(2-methyl-1,3-dioxolan-2-yl)methyl]buta-1,3-</u> <u>dienyloxytrimethylsilane 26</u>

n-Butyllithium (2.5 in hexane; 0.52 ml, 1.3mmol) was added to a 0°C solution of diidopropylamine (0.17 ml,1.30 mmol) in THF (6 ml). The solution was warmed to rt, stirred for 5 min then cooled to -78°C. Chlorotrimethylsilane (0.20 ml, 1.60 mmol) was added, followed by dropwise addition of methyl 3 –methyl-4-(2-methyl-1,3-dioxolan-2-yl)but-2-enoate (200mg, 1 mmol) in THF (3 ml). The resulting solution was stirred at - 78°C for 90 min then warmed to rt and concentrated without heating in vacuo. The residue was diluted with dry n-pentane (10 ml) and filtrated and concentrated. Yield 75%; IR 3011, 2962, 2888, 1707, 1646, 1598, 1254, 1097, 1050, 676 cm⁻¹; CDCl₃

HNMR 5.26 (1H, d, J 2.5, CH), 4.81 (1H, d, J 2.5, H₂C=C), 4.24 (1H, s, H₂C=C), 3.97-3.95 (4H, m, OCH₂OCH₂), 2.50 (2H, s, CH₂), 1.37 (3H, s, CH₃), 0.26 (9H, s, OSiCH₃); CNMR 157.7 (C), 138.3 (C), 110.9 (CH2), 110.2 (C), 79.2 (CH), 64.4 (CH₂), 55.0 (CH), 47.3 (CH₂), 23.8 (CH₃), 20.5 (CH₃)

Synthesis of 5-hydroxy-2-mehoxy-7-[(2-methyl-1,3-dioxolan-2-yl)-methyl]-1,4naphtoquinone 28

1-methoxy-3-[(2-methyl-1,3-dioxolan-2-yl)methyl]buta-1,3-dienyloxytrimethylsilane (1.88 g, 6.91 mmol) in DCM (25 ml) was added dropwise to a 0°C solution of 2-bromo-5-methoxy-1,4-benzoquinone (500 mg, 2.30 mmol) and diisopropylethylamine (0.48 ml, 2.76 mmol) in DCM (25 ml). The resulting green solution was stirred for 2 h then silica gel (10g) was added and the mixture warmed to rt and stirred for 24h. The mixture was filtrated to remove silica and the filtrate concentrated without heating. The crude was purified in column eluiting with petroleum ether- ethyl acetate 6:4 Yield 90%; mp 173-174 °C; 3694, 2987,1685, 1635, 1605, 1380, 1346, 1275, 1242, 1115 cm-1; CDCl₃ HNMR 12.17 (1H, s, OH), 7.65 (1H, d, J 1.5, ArH), 7.25 (1H, d, J 1.5, ArH), 6.11 (1H, s), 3.99-3.95 (2H, m, OCH₂), 3.94 (3H, s, OCH₃), 3.82-3.86 (2H, m, OCH₂), 3.01 (2H, s, CH₂), 1.36 (3H, s, CH₃); CNMR 190.5 (C), 179.6 (C), 161.1(C), 160.8 (C), 130.5 (C), 126.7 (CH), 122.3 (CH), 112.7 (C), 109.5 (CH), 109.1 (C), 64.9 (CH₂), 56.7 (CH₃), 45.5 (CH₂), 24.5 (CH₃)

Synthesis of 2-(bromomethoxy)-5-hydroxy-7-((2-methyl-1,3-dioxolan-2yl)methyl)naphthalene-1,4-dione 34

To a stirred solution of 5-hydroxy-2-methoxy-7-(2-methyl-[1,3]dioxolan-2-ylmethyl)-[1,4]naphthoquinone (100 mg, 0.3 mmol) in DCM (10 ml) bromine (110 mg, 0.66 mmol) was added at rt followed by morpholine (catalytic amount). The reaction mixture was stirred at rt for 2h and then quenched with $Na_2S_2O_3$ saturated solution. The aqueous layer was extracted with ethyl acetate (10 ml x2) and evaporated under reduced pressure. The crude was purified in column eluting with petroleum ether – ethyl acetate 8:2

Yield 30%; IR 3690, 3607, 2360, 1626, 1674, 1569, 1488, 1239, 1017; mp 172-173°C; HNMR CDCl3: 12.51 (1H, OH), 7.46 (1H, d, ArH), 7.16 (1H, d, ArH), 5.89 (1H, s, ArH), 3.97-3.94 (2H, m), 3.88 (2H, s), 3.86-3.82 (2H, m), 2.97 (2H, s), 1.34 (3H, s); CNMR 188.86 (C), 182.53 (C), 160.29 (C), 154.24 (C), 144.79 (C), 132.21 (C), 126.10 (CH), 121.90 (CH), 112.91 (C), 110.22 (CH), 109.22 (C), 66.41 (CH₂), 64.91 (CH₂),

45.15 (CH₂), 45.44 (CH₂), 24.41 (CH₃)

Synthesis of 5-hydroxy-3-iodo-2-methoxy-7-((2-methyl-1,3-dioxolan-2yl)methyl)naphthalene-1,4-dione 35

To a stirred solution of 5-hydroxy-2-methoxy-7-(2-methyl-[1,3]dioxolan-2-ylmethyl)-[1,4]naphthoquinone (100 mg, 0.3 mmol) in DCM (10 ml) iodine (167 mg, 0.66 mmol) was added at rt followed by morpholine (catalytic amount). The reaction mixture was stirred for 2h and then quenched with $Na_2S_2O_3$ saturated solution (10 ml). The acqueous layer was estracted with ethyl acetate (10 ml x2) and evaporated under reduced pressure. The crude was purified in column eluiting with petroleum ether ethyl acetate 8:2.

Yield 70%; IR: 3682, 3011, 1627, 1520, 1425, 1239, 1103; mp165-166°C; CDCl₃ HNMR: 12.20 (1H,s, ArOH), 7.48 (1H, d, J 1.6, ArH); 7.13 (1H, d, J 1.6, ArH), 3.88 (3H, s, OCH₃), 3.94-3.84 (2H, m, OCH₂), 3.81-3.78 (2H, m, OCH₂), 2.94 (2H, s, CH₂), 1.34 (3H, s, CH₃); CNMR 184.64 (C), 180.43 (C), 160.57 (C), 158.47 (C), 145.73 (C), 130.89 (C), 125.97 (CH), 122.62 (CH), 111.39 (C), 110.47 (C), 109.15 (C), 67.48 (CH₂), 64.93(CH₂), 56.23 (CH₃), 45.44(CH₂), 24.44 (CH₃)

Synthesis bromo-5-hydroxy-7-((E)-2-hydroxyprop-1-enyl)-2-methoxynaphthalene-1,4-dione 36, 37

A solution of hydroxy-2-methoxy-7-(2-methyl-[1,3]dioxolan-2-ylmethyl)-[1,4]naphthoquinone (0.33 mmol, 100 mg) in acetic acid (3ml) was treated at rt with bromine (0.33mmol, 53 mg) and stirred overnight. The cooled reaction mixture was poured into ice and the precipitate filtered and washed with ethanol.

6-bromo-5-hydroxy-7-((E)-2-hydroxyprop-1-enyl)-2-methoxynaphthalene-1,4dione 36: Yield 35%; IR 3691, 3045, 1636, 1603, 1241, 1114; HNMR CDCl₃ 12.82 (1H, s, OH), 7.83 (1H, s, ArH), 5.93 (1H, s), 4.41 (3H, CH₃), 2.50 (1H, s), 2.45 (3H, s); CNMR 183.55 (C), 183.52 (C), 177.44 (C), 159.72 (C), 153.83 (C), 143.33 (C), 129.48 (C), 121.41 (C), 120.42 (CH), 119.46 (C), 113.16 (CH), 63.35 (=CH), 62.45 (OCH₃), 27.49 (CH₃)

8-bromo-5-hydroxy-7-((E)-2-hydroxyprop-1-enyl)-2-methoxynaphthalene-1,4dione 37: Yield 25%; IR 3607, 2999, 1724, 1686, 1274; HNMR CDCl₃ 12.82 (1H, s, OH), 7.89 (1H, s, ArH), 6.03 (1H, s, ArH), 4.37 (3H, s), 2.54 (1H, s), 2.50 (3H, s); CNMR 183.54 (C), 183.20 (C), 161.00 (C), 157.78 (C), 143.28 (C), 129.34 (C), 122.17 (CH), 121.28 (C), 119.44 (C), 115.93 (C), 113.00 (CH), 62.30 (OCH₃), 52.37 (=CH), 28.07 (OCH₃)

Synthesis of 5-hydroxy-6-iodo-2-methoxy-7-(2-oxopropyl)naphthalene-1,4-dione 38

A solution of hydroxy-2-methoxy-7-(2-methyl-[1,3]dioxolan-2-ylmethyl)-[1,4]naphthoquinone (0.33 mmol, 100 mg) in acetic acid (3ml) was treated at rt with iodine (0.33mmol, 84 mg) and stirred overnight. The cooled reaction mixture was poured into ice and the precipitate filtered and washed with ethanol.

Yield 30%; IR 3607, 1724, 1686, 1636, 1274; HNMR CDCl₃ 13.03 (1H, s, OH), 7.53 (1H, s, ArH), 6.15 (1H, s), 4.06 (2H, s, CH₂), 3.96 (1H, s, OCH₃), 2.33 (1H, s, CH₃); CNMR 203.18 (C), 190.10 (C),179.65 (C), 161.02 (C), 158.08 (C), 143.17 (C), 130.25 (C), 121.66 (CH), 113.42 (C), 109.23 (CH),105.00 (C), 56.86 (CH₃), 51.30 (CH₂), 30.18 (CH₃)

BIBLIOGRAPHY

| [85P2669] | Hanumaiah T., Marshall D.S., Rao B.K., Phytochemistry, 24,2669- |
|---------------|---|
| | 2672, 1985 |
| [90JOC1466] | M.Blouin, M.C. Beland, P. Brassard, J. Org. Chem., 55, 1466- |
| | 1471, 1990 |
| [91JCSR327] | Cowe H. J., Cox P.J., R.A. Howie, J.Crystallogr. Spectrosc.Res, 21, |
| | 327-332, 1991 |
| [92AJC2025] | D.W.Cameron, I.T. Crosby, G.I. Feutrill, Aust. J. Chem., 45, 2025- |
| | 2035, 1992 |
| [95ACDD277] | C.H. Ngueyen, Anti-Cancer Drug Design, 10, 277, 1995. |
| [98ADP352] | A.H. Abadi, Arciv. der Pharmazie, 331, 352, 1998. |
| [98SCI1504] | M.R. Redimbo, L. Stewart, P. Kuhn, J.J. Champoux, W.G.J.Hol, |
| | Science, 279, 1504, 1998 |
| [99BMC1591] | P. Barraja, P. Diana, A. Lauria, A. Passannanti, A. M. Almerico, |
| | C. Minnei, S.Longu, D. Congiu, C. Musiu, L. Pisano, P. La Colla, |
| | Bioorg. Med. Chem., 7, 1591, 1999. |
| [99JMC2561] | G. Cirrincione, A.M. Almerico, P.Barraja, P. Diana, A. Lauria, A. |
| | Passannanti, A. Pani, E. Pinna, F. Scintu, C. Musiu, L. Pisano, P. |
| | La Colla, J. Med. Chem., 37, 267, 1999. |
| [99NPR267] | M.A. Brimble, L.J. Duncalf, M.R. Nairn, Nat. Prod. Rep., 16, 267- |
| | 281, 1999 |
| [00T1937] | M.A. Brimble, M.R. Nairn, H. Prabaharan, Tetrahedron, 56, 1937- |
| | 1992, 2000 |
| [01JMC594] | D.S. Laurence, J. Med. Chem., 44, 594, 2001. |
| [02BMC3153] | P.Chen, Bioorg. Med. Chem., 12, 3153, 2002. |
| [02EJMC267] | P. Diana, P. Barraja, A. Lauria, A.M. Almerico, G. Dattolo, G. |
| | Cirrincione, Eur. J. Med. Chem., 37, 267, 2002. |
| [02PNAS15387] | B.Staker, K.Hjerrild, M.D.Feese, C.A.Behnke, A.B.Burgin, |
| | L.Stewart, Proc. Natl. Acad. Sci. U.S.A. 99, 15387, 2002. |
| [03BMC2371] | P. Diana, P. Barraja, A. Lauria, A. Montalbano, A.M. Almerico, |

| | G. Dattolo, G. Cirrincione Bioorg. Med. Chem., 11, 2371, 2003. |
|--------------|---|
| [04ACR3775] | P.Barraja et al. Anticanc. Res. 24, 3775, 2004 |
| [04NAT198] | R.B. Ravelli, B.Gigant, P.A.Curmi, I.Joudain, S.Lachkar, |
| | A.Sobel, M.Knossow, Nature 428, 198, 2004. |
| [04TL939] | A. Shimbashi, Y. Ishikawa, S. Nishiyama, Tetrahedron Lett., |
| | 45,939-941, 2004 |
| [05BMCL295] | P.Barraja er al. Bioorg. Med. Chem.Lett., 13, 295, 2005. |
| [05JMC6107] | T.L.Nguyen, C.McGrath, A.R.Hermone, J.C.Burnett, |
| | D.W.Zaharevitz, B.W.Day, P.Wipf, E.Hamel, R.Gussio J. Med. |
| | <i>Chem.</i> 48, 6107-6116, 2005 . |
| [06IND343] | T.S. Bekaii-Saab, Inv. New Drugs, 24, 343, 2006. |
| [07IND147] | AM. Alousi, Inv. New Drugs, 25, 147, 2007 |
| [07TL1545] | A. Shimbashi, S. Nishiyama, Tetrahedron Lett., 45, 939-941, 2007 |
| [08EJC1684] | S. D. Undevia, Eur. J. Cancer 44, 1684, 2008. |
| [08IND331] | W. Stock, Inv. New Drugs, 26, 331, 2008. |
| [08JMC2387] | P. Diana, A. Martorana, P. Barraja, A. Montalbano, G. Dattolo, G. |
| | Cirrincione, F. Dall'Acqua, A. Salvador, D. Vedaldi, G. Basso, |
| | G. Viola J. Med. Chem. 51, 2387, 2008. |
| [08JMC4609] | M.A.Cinelli, A.Morrell, T.S.Dexheimer, E.S.Scher, Y.Pommier, |
| | M.Cushman J. Med. Chem 51, 4609, 2008 |
| [08JMC4620] | G.M.Chinigo, M.Paaige, S.Grindrod, E.Hamel, |
| | S.Dakshanamurthy, M.Chruszcz, W.Minor, M.L. Brow J. Med. |
| | <i>Chem.</i> 51, 4620-4631, 2008. |
| [08NPR376] | J.Sperry, P.Bachu, M.A. Brimble, Nat. Prod. Rep., 25, 376-400, |
| | 2008 |
| [08WO041264] | G. Cirrincione, P. Diana International Patent: PCT WO |
| | 2008/041264 A1. |
| [09BMC6279] | T.Liu, X.Dong, N.Xue, R.Wu, Q.He, B.Yang, Y.Hua, Bioorg. |
| | Med. Chem., 17, 6279-6285, 2009. |
| [09JMC7512] | La Regina et al. J. Med. Chem, 52, 7512-7527, 2009 |