

1 **Mixotrophic phytoplankton dynamics in a shallow Mediterranean water body: how to make a**
2 **virtue out of necessity**

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11
12 **Abstract**

13 Mixotrophy is a combination of photosynthesis and direct access to organic carbon sources, mainly
14 through osmotrophy or phagotrophy. This strategy is adopted by several, phylogenetically distinct,
15 phytoplankton groups and is commonly occurring in marine, brackish and freshwater ecosystems.
16 Traditionally, it has been put in relation to both scarcity of inorganic nutrients and poor light
17 conditions. However, we observed blooms of the mixotrophic, toxic haptophyte *Prymnesium*
18 *parvum* in different periods of the year, and under variable resources availability. The analysis of a
19 6.5-year data set of phytoplankton weekly records from a Sicilian shallow lake (Biviere di Gela,
20 south-eastern Sicily) allowed us to hypothesize that a depleted condition as regard inorganic
21 nutrients is not the main fuel to the growth of *Prymnesium parvum*, neither this is due to light
22 limitation. The results achieved show that an increased availability in suitable preys can stimulate
23 the growth of this phagotrophic photoautotroph. Contemporary, it was not found any clear
24 environmental patterns to explain species dominance and growth patterns as related to inorganic
25 nutrient availability. Moreover, it is shown that these organisms tend to monopolise resources when
26 these become available irrespectively of seasons, and under variable conditions as regard inorganic
27 nutrient availability.

28
29 **Keywords:** *Prymnesium parvum*, phagotrophy, BOD₅, nutrient availability, biological interactions

30 **Introduction**

31 The evolutionary pathways followed by protists in the last 2 billion years resulted in a legacy of
32 several, often amazing, nutritional modes, which witness the complexity of the polyphyletic origins
33 of these organisms (Zimmer, 2009; Simpson et al., 2017). By overturning this perspective, the wide
34 spectrum of nutritional modes exhibited by protists has been certainly contributing to ensure their
35 persistence and to improve their chance of survival (Yafremava et al., 2013).

36 Among these nutritional modes, the holophytic (photosynthetic) and holozoic (zoo-trophic,
37 ingestive or phagotrophic) are the most popularly renowned since they are ascribed to “plants” and
38 “animals”, respectively.

39 Eukaryotic phytoplankton form an ecological group of photosynthetic protists adapted to
40 live in suspension in the water column of aquatic ecosystems (Reynolds, 2006). Although, they
41 show a nutritional mode superficially similar to that of plants, phytoplankton encompass a broader
42 range of both phylogenetic diversity and life history strategies (Rengefors et al., 2017). This
43 diversity is mirrored by their broad spectrum of nutritional modes, and several phytoplankton taxa
44 can heterotrophically access energy and carbon.

45 One of the most important steps giving birth to photosynthetic eukaryotes, according to
46 endosymbiont theory, was an error recurrently occurring in phagotrophy, which has played a central
47 role in the evolution of photosynthetic eukaryotes (Raven, 1997). Actually, through a sequence of
48 complex endosymbiotic events involving engulfment (i.e. ingestion lacking digestion) of unicellular
49 photosynthetic prokaryotes and/or eukaryotes, the plastids, and other organelles, developed in algae
50 (Keeling, 2013). Kleptoplasty is a quite common phenomenon among holozoic organisms and it
51 supports the endosymbiotic origin of chloroplasts: several ciliate and dinoflagellate species
52 photosynthesize by sequestering chloroplasts from their photosynthetic preys, and by maintaining
53 these organelles in their cytoplasm for a variable amount of time (Johnson et al., 2016). This
54 strategy, adopted not only by protists but also by some marine molluscs (Händeler et al., 2008),

55 probably represented the first step in the evolutionary path that has led to actual photosynthetic
56 eukaryotes.

57 Since the recognition of the fundamental role exerted by the microbial loop in the
58 functioning of aquatic ecosystems (cf. Azam et al., 1983), a net distinction between
59 photoautotrophy and heterotrophy appeared insufficient to fully explain the pathway of carbon and
60 nutrient flux in aquatic ecosystems; therefore, the importance of a further nutritional mode,
61 mixotrophy, started receiving greater attention (Jones, 2000). Mixotrophy couples the
62 photosynthetic and the heterotrophic nutritional modes, and forms a continuum in the way in which
63 protists, but also bacteria (Eiler, 2006), can access different sources of energy and carbon (Jones,
64 1997; Stoecker, 1998). Recently, Mitra et al. (2016), proposed a functional grouping of planktonic
65 protists in an eco-physiological context which includes among constitutive mixotrophs (those
66 organisms bearing their own chloroplasts) also those ciliates able of kleptoplasty (non-constitutive
67 mixotrophs).

68 Several phytoplanktonic organisms, especially found among those bearing flagella, exhibit a
69 mixotrophic nutritional mode: they can therefore photosynthesize and contemporary, or
70 alternatively, can derive energy from the uptake of dissolved organic compounds (osmotrophy) or
71 from the ingestion of organic particles or other microorganisms (phagotrophy). In addition,
72 depending on resource availability, some species must employ mixotrophy obligately and some
73 others can do it facultatively (Ghyoot et al., 2017). Mixotrophy, along with species morphology
74 (Naselli-Flores & Barone, 2011), and nitrogen fixation, represents an important trait related to
75 resource acquisition by phytoplankton and therefore to their trophic niche (Litchman & Klausmeier,
76 2008). However, the existing literature reports contrasting information as regard the levels of
77 nutrients and light which can make mixotrophs more competitive than phototrophs in natural
78 environments. Both oligotrophic (e.g. Arenovski et al., 1995) and eutrophic conditions (e.g.
79 Burkholder et al., 2008) were reported as favourable for mixotrophic phytoplankton. According to
80 some authors, mixotrophy cannot allow phytoplankton to escape the need of photosynthesizing and

81 the availability of sufficient light remains a fundamental requisite for mixotrophic phytoplankton
82 (e.g. Caron et al., 1993). Nevertheless, there are papers suggesting that constitutive mixotrophy can
83 play a positive role when severe and prolonged light limitation occurs (e.g. Jones et al., 2009).

84 In this paper we present data on the phytoplankton of a natural, shallow lake in south-eastern
85 Sicily (Italy). The lake is largely dominated by constitutive mixotrophic phytoplankton and has
86 been regularly sampled since 2005. A 6.5-year dataset, constituted by weekly phytoplankton
87 samplings, was used to verify the hypothesis that the abundance of mixotrophic algae is not related
88 to the availability of inorganic nutrients, whether they are present in high or low concentrations.
89 Moreover, our aim is to show that these organisms tend to monopolise resources when these
90 become available irrespectively of seasons, and under variable environmental.

91

92 **Materials and Methods**

93 Study Site

94 The Biviere di Gela (BdG, 37°01'12" N, 14°20'28" E) is a natural coastal lake located in SE Sicily
95 (Italy) at 1.3 km from the Mediterranean Sea, 8 m a.s.l. The lake has no surface outflow and no
96 natural inflow and it is mainly fed by underground waters flowing from North to South. The lake
97 lies in a depression which intercepts the water table. The downgradient underground flow also
98 constitutes a barrier against seawater intrusion and avoid that the electrical conductivity of the lake
99 waters exceeds 2 mS cm⁻¹.

100 In 1988, the lake has been designated for the List of Wetlands of International Importance
101 and included in the ‘‘Ramsar Act’’. Nine years later, a Nature Reserve has been created by the
102 Sicilian Region to preserve the several rare and endangered vertebrate species (e.g., amphibians,
103 turtles, and birds) that inhabit the lake, and the lake itself which has an historical value, being
104 mentioned by Pliny the Elder in his *Naturalis Historia*. Nevertheless, the lake is surrounded by

105 greenhouses supporting intensive agriculture. These infrastructures compete with the lake for water
106 resources and exploit the same water table for their irrigation purposes.

107 As a result of overexploitation of groundwaters and climate change (Barone et al., 2010), the
108 lake risked to disappear at the end of the last century. This caused great concern, and an artificial
109 canal downstream of a dam reservoir (Lake Dirillo – 37°07'19" N, 14°41'54" E) was therefore
110 opened in 2004. Immediately after the BdG was quickly re-filled and the rich macrophyte belts
111 which characterised the littoral zone of the lake, and which were destroyed by the drop of lake level
112 and the concurrent increase in salinity, started recovering. Lake Dirillo is a eutrophic reservoir
113 receiving the untreated sewages from two urban centres (Vizzini – 6300 inhabitants, and Licodia
114 Eubea – 3100 inhabitants) located in its catchment. During the study period, inflows from Lake
115 Dirillo quite regularly reached the BdG through the canal, from 2011 to early spring 2013. This was
116 done to minimize the extent of water levels fluctuations in the BdG (also due to the overexploitation
117 of the water table caused by the irrigation needs in the nearby greenhouses) and to avoid the
118 vanishing of this Ramsar site. From spring 2013 to autumn 2016, also thanks to higher
119 precipitation, no significant water abstraction from Lake Dirillo was necessary to maintain the lake
120 levels. In November 2016, a quite rapid decrease in the lake level occurred and caused concern in
121 the Management of the Nature Reserve, which again moved waters from Lake Dirillo to refill the
122 Biviere di Gela.

123 Phytoplankton of Biviere di Gela has been regularly studied since 2005 and further
124 information on the lake is available in Barone et al. (2010), Naselli-Flores & Barone (2012),
125 Jeppesen et al. (2015) and Naselli-Flores et al. (2016).

126

127 Data collection and analysis

128

129 Phytoplankton data in the present study refers to weekly collections, from January, 2011 to May,
130 2017. Water samples for phytoplankton counting were collected at 10 am (\pm 30 min) in a station

131 located in the central part of the lake, approx. 20 cm below the surface, using a 100 ml dark-glass
132 bottle. These samples were preserved in Lugol's iodine solution immediately after collection.

133 Net samples were contemporary collected by towing vertically and horizontally a 40 μm
134 mesh size plankton net. Samples were immediately fixed in 4% buffered formaldehyde. However,
135 live samples were also taken for species identification. Taxa were identified at the lowest taxonomic
136 rank (species) in the majority of cases using the most up-to-date phycological literature.

137 A Zeiss Axiovert 100 microscope was used for cell counting which was performed in
138 accordance with the sedimentation method developed by Utermöhl; all the individuals (cells or
139 colonies) present in forty random fields at 400x magnification were counted.

140 The biovolume was computed approximating the shape of the cell or colony to simple
141 geometric shapes as shown in Hillebrand et al. (1999). To calculate the cell or colony volumes, 10
142 individuals of each recorded species were measured using a Zeiss AxioVision image analysis
143 system on the same microscope used for counting. The average value of each measurement was
144 used to calculate the average biovolume of species. Biovolumes were transformed in biomass
145 values assuming that phytoplankton has unit specific gravity ($10^9 \mu\text{m}^3 = 1 \text{ mg}$). Wet weight biomass
146 of each population was calculated multiplying the recorded abundance of populations by the
147 average biomass estimates of their individuals.

148 Net growth rates were computed from weekly cell abundance data using the equation:

149
$$r = \ln(N_1) - \ln(N_0) / (t_1 - t_0) \quad [\text{d}^{-1}]$$

150 where r is the net growth rate, and N_1 and N_0 are the number of phytoplankton cells in a unit volume
151 at time 1 (t_1) and time 0 (t_0), respectively.

152 In the same sampling station, water temperature was measured weekly, along vertical
153 profiles, using a YSI 556 MPS multiparametric probe. Water level data were supplied by the
154 Management Board of the Nature Reserve "Biviere di Gela".

155 Water samples for nutrients (Reactive Phosphorus – RP, and Dissolved Inorganic Nitrogen –
156 DIN) were collected sub-superficially, at monthly intervals. DIN refers to the sum of N-NO₃ and N-
157 NH₄. The chemical analyses were performed according to Tartari & Mosello (1997).

158 BOD₅ was measured as a surrogate of the degree of the organic pollution of the water
159 flowing from Lake Dirillo. The values were considered as proxies of bacterial abundance in the
160 waters. Samples for BOD₅ were taken every two weeks and additional samples were collected when
161 an evident flow in the artificial canal from Lake Dirillo occurred. The water was taken sub-
162 superficially and stored in 300 ml sterilized dark bottles. The analyses were performed in the lab of
163 the Nature Reserve, within a few minutes (15-30 minutes) from water collection. The analytical
164 method used (APAT, 2003), allows a direct determination of BOD₅ by measuring the concentration
165 of dissolved oxygen (Winkler method) before and after a five days incubation at 20 °C.

166

167 **Results**

168

169 Trends in selected environmental variables

170

171 Lake depth ranged between 5.5 and 7.9 m. Seasonal water level fluctuations (lower depth in
172 summer and higher in winter) between 0.5 and 1.5 m occurred in the lake during the studied period
173 (Fig. 1). Water temperature ranged between 12.5 (±0.4) °C (February) and 29.5 (±0.6) °C (July-
174 August). Due to the high summer water temperatures, the water column occasionally showed a
175 weak stratification in July and/or August in spite of the shallowness of the lake. These occasional
176 thermoclines were punctual, and they were seldom recorded in two consecutive sampling dates.

177 The monthly dynamics of nutrients showed a marked and repetitive seasonal pattern with
178 highest concentrations in winter (RP above 15 µg L⁻¹ and DIN above 200 µg L⁻¹) and lowest in
179 summer (RP below 10 µg L⁻¹ and DIN below 200 µg L⁻¹). In general, DIN showed a decreasing

180 tendency from the beginning of the study period and until summer 2016. Later on, the values
181 increased again (Fig. 2). Conversely, RP values were slowly growing throughout the study period.

182 Oxygen contents in the superficial lake waters were generally close to, or slightly higher
183 than, 100% of saturation. BOD₅ values were generally around 2.0 mg L⁻¹. However, significantly
184 higher values (up to 6.9 mg L⁻¹) were sporadically recorded in the period 2011-2013 and from the
185 end of 2016 onward (Fig. 1). Values above 3 mg L⁻¹ generally followed water inflows from Lake
186 Dirillo.

187

188 Phytoplankton structure and dynamics

189

190 Phytoplankton in the BdG was quite rich and diversified. A total number of 141 phytoplankton taxa
191 were identified in the lake during this study. These were ascribed to the following taxonomic
192 groups: cyanobacteria (8), haptophytes (3), cryptophytes (7), chlorophytes (91), bacillariophytes
193 (8), dinoflagellates (5), euglenophytes (12), xanthophytes (2), ochrophytes (3). “Picoplankton” and
194 “unidentified flagellates < 5 µm” were considered as monospecific taxa. Ochrophytes (two
195 unidentified species of *Mallomonas* and *Pedinella hexacostata* Vysotskij) appeared
196 idiosyncratically and punctually in a few winter samples but, when present, they were quite
197 abundant.

198 Phytoplankton total biomass varied between 2.5 and 25 mg L⁻¹ in the period January 2011 -
199 April 2013 and was characterised by frequent peaks of the haptophyte *Prymnesium parvum* Carter
200 (Fig. 1). This species contributed up to 97% to total haptophyte biomass (other haptophytes present
201 in the lake were *Hymenomonas roseola* Stein and *Chrysochromulina parva* Lackey) and up to 80%
202 to total phytoplankton biomass in spring 2011. It dominated the phytoplankton assemblage from
203 mid-October 2011 to the beginning of June 2012 with percentage values ranging between 50% and
204 95%. Virtually absent in July and August 2012, the species again started dominating in the
205 beginning of November 2012 showing relative biomass values higher than 60% until mid-April

206 2013. Since then, *Prymnesium parvum* showed relatively low values of biomass until October 2016,
207 when it started growing again and peaked at values above 35 mg L⁻¹ in the last week of November.
208 Later on, the species dominated the phytoplankton assemblage until mid-April 2017.

209 In the period May 2013 – September 2016, phytoplankton biomass ranged between 3.1 and
210 47 mg L⁻¹ and was characterised by summer peaks of Cyanobacteria [mainly *Cylindrospermopsis*
211 *raciborskii* (Wołoszyńska) Seenaya & Subba Raju and *Chrysochloris ovalisporum* (Forti)
212 E.Zapomelová, O.Skácelová, P.Pumann, R.Kopp & E.Janecek]. Their dominance frequently lasted
213 until mid-autumn with absolute biomass values peaking above 30 mg L⁻¹ at the end of October,
214 2014, in mid-June 2015, and at the end of September 2016 (Fig. 3).

215 When *Prymnesium parvum* and cyanobacteria biomass values were relatively lower, other
216 phytoplankton taxa could grow up and reached relatively higher biomass percentages, although
217 without showing a clear seasonal pattern. In particular, cryptophytes (*Cryptomonas* spp. and
218 *Plagioselmis* sp.) were generally more abundant or sub-dominant in winter, whereas
219 “chlorococcalean” chlorophytes and diatoms (*Cyclotella* spp. and *Diatoma tenuis* C.Agardh) tended
220 to be more abundant in spring and/or in summer (the trends of these taxa are shown in the
221 Electronic Supplementary Material). In these seasons, also euglenophytes and dinoflagellates were
222 relatively more abundant, but never exceeded 7% of total biomass.

223 Mixotrophic (which included, in order of abundance, haptophytes, cryptophytes,
224 dinoflagellates, euglenophytes and the ochrophyte *Pedinella hexacostata*) and phototrophic species
225 were alternatively present in the lake on a seasonal basis (Fig. 4). In winter mixotrophs were
226 relatively more abundant, whereas in summer autotrophs prevailed. This was particularly evident in
227 the periods 2011-2013 and 2016-2017, when mixotrophic species formed the bulk of biomass in the
228 BdG. A lower relative abundance of mixotrophs was recorded in 2014-2015, when cyanobacteria
229 showed summer blooms, and the alternation with autotrophs was less marked in that period.

230 No significant relationships were found among the different phytoplankton taxonomic
231 groups, nor between mixotrophs and phototrophs biomass values. Analogously, no significant

232 relationships were detected when correlating phytoplankton groups and environmental variables.
233 However, when the positive values of *Prymnesium parvum* growth rates were correlated to the
234 increase in water level which had occurred one week before, a significant correlation was clear (Fig.
235 5; n=89, r= 0.91, p<0.001). Figure 5 also shows the different seasonal contributions to the
236 relationship: 16 points refer to winter growth, 8 to spring growth, 6 to autumn growth and 4 to
237 summer growth. Although more frequent in winter, *Prymnesium parvum* also showed relatively
238 high growth rates in all the seasons, including summer. All these points were related to an increase
239 in depth higher than 10 cm, which corresponded to water inflows from Lake Dirillo. The remaining
240 55 points corresponded to “natural” water incomes and were related to growth rates lower than 0.05
241 d⁻¹.

242

243 **Discussion**

244 Environmental variability

245 Water level fluctuations in the BdG were not only due to the natural balance between inflows and
246 outflows. The increase in water level also occurred “upon request” from Lake Dirillo to refill the
247 water body and avoid the risks of excessive decrease in water level as already recorded in the past
248 years (Barone et al. 2010). The period of inflows from Lake Dirillo were therefore monitored by the
249 Nature Reserve Management Board and were easily discernible from inflows due to precipitation.

250 Biochemical Oxygen Demand (BOD) represents the quantity of oxygen consumed by
251 microorganisms to fuel the aerobic decomposition of organic materials. The BOD therefore has been
252 considered a reliable proxy of bacteria density in water samples (e.g. Kagalou et al., 2002). In
253 addition, Olutiola et al. (2010) found that it was also positively correlated to inorganic nitrogen
254 concentrations, which constitutes an end-product of organic matter degradation. The results of the
255 present study show that BOD₅ in the BdG showed frequent peaks in the period 2011-2013 and at
256 the end of 2016. In the same periods, DIN values higher than those recorded in 2014-2016 were
257 measured in the BdG. These periods corresponded to water inflows from Lake Dirillo, which

258 receives untreated sewage from the municipalities located in its catchment. Although no data are
259 available on the amounts of organic matter in Lake Dirillo, these repeated peaks make reasonable to
260 hypothesize that Lake Dirillo represents a source i) of bacteria and/or ii) of organic pollution which
261 triggers bacterial growth in the BdG.

262

263 Phytoplankton dynamics

264 Mixotrophy is considered an advantageous mode of nutrition when inorganic nutrients are limiting
265 and/or the light regime is poor (Pållson & Granéli, 2004). Lake BdG, due to its Mediterranean
266 location, is unlikely subjected to a poor light regime (Obrador & Pretus, 2008). Moreover,
267 planktonic organisms entrained in the turbulent water mixing, have similar chances to access light
268 when turbidity has an algal origin, especially when biomass is not dominated by buoyant
269 cyanobacteria (Reynolds, 2006; Naselli-Flores, 2014). In addition, dissolved nutrients in BdG
270 where generally above the limiting threshold, even though the biomass values reached by
271 phytoplankton in the lake were well above those attained in other Sicilian lakes showing a
272 comparable nutrients availability (Naselli-Flores & Barone, 1994; Naselli-Flores, 1999; 2000).

273 Mixotrophic phytoplankton largely dominated the phytoplankton of BdG during the periods
274 characterised by water inflows from Lake Dirillo. Among mixotrophic species, *Prymnesium*
275 *parvum* frequently bloomed during 2011-2013 and at the end of 2016. The species is well-known to
276 produce a variety of toxins (Eikrem et al., 2017) endangering the aquatic fauna (both invertebrates
277 and vertebrates). Moreover, it has been proven to exert an allelopathic effect addressed toward
278 outcompeting other phytoplankton species (Granéli & Johansson, 2003). These metabolites can in
279 some cases be used to directly kill potential preys, or, indirectly to promote bacteria growth through
280 the decomposition of killed organisms. In addition, as shown by Croft et al. (2006), many different
281 species of phytoplankton, including several haptophyte genera, cannot synthesize some vitamins
282 and get them from bacteria by phagotrophy. *Prymnesium parvum* is actually able to support
283 population growth through heterotrophy employing both osmotrophic and phagotrophic nutritional

284 modes (Roelke et al., 2016 and literature therein). In the studied lake, a significant positive
285 relationship was found between the instantaneous growth rates of *P. parvum* and the increment of
286 water level recorded in the lake one week before. These increments were due to water inflows from
287 Lake Dirillo, which caused an increase in the BOD₅ values measured in BdG, and occurred in any
288 season, when the intense use of water for irrigation was considered a menace for the water level
289 stability of the BdG. Accordingly, high growth rate of the haptophyte did not show a well-defined
290 seasonality, even though they were more frequent in winter. These repeated coincidences allow to
291 hypothesize that *Prymnesium parvum* benefited from an increased organic matter (and bacteria)
292 entering the lake through the inflows from Lake Dirillo. When the water inflows from Lake Dirillo
293 stopped (2014-2016), *P. parvum* biomass remained below 5 mg L⁻¹, and the species was virtually
294 absent for long periods. In the absence of *P. parvum*, also a decrease in inorganic nitrogen was
295 recorded. Filamentous nitrogen fixing cyanobacteria became very abundant, especially during
296 summer 2014, 2015 and 2016. This negative interaction between *P. parvum* and cyanobacteria had
297 been already observed immediately after the establishment of the water connection between Lake
298 Dirillo and the BdG (Barone et al, 2010). However, the dominance of the haptophyte was in that
299 case attributed to the decrease in conductivity caused by the inflows from Lake Dirillo (Jeppesen et
300 al., 2015). The present study offered an alternative explanation for *P. parvum* growth, which was
301 likely to be supported by the organic (and eventually bacteria) loads from Lake Dirillo.

302 Other supposed mixotrophic organisms in the phytoplankton of the studied lake belonged to
303 cryptomonads, euglenids and dinoflagellates.

304 Cryptomonads represented, according to their relative biomass, the second most important
305 group of mixotrophs in the lake. These organisms show ultrastructural details (a furrow-gullet
306 system surrounded by ejectosomes) suggesting an “attitude” to phagotrophy and several reports
307 indicate that these organisms may ingest bacteria (e.g. Tranvik et al., 1989). However, other more
308 recent studies refuse bacterivory (Hoef-Hemden & Archibald, 2017; Tanifuji & Onodera, 2017),
309 and a lack of consensus pervades the available literature. Although less abundant in summer, these

310 algae were persistent throughout all the seasons, as already observed in several Sicilian lakes
311 (Barone & Naselli-Flores, 2003). No relationships were found among Cryptophytes biomass and
312 growth rates, environmental variables and other phytoplankton groups in the studied lake.

313 Although belonging to a group of organisms largely considered as “heterotrophic”,
314 photoautotrophic euglenids have a feeding apparatus highly reduced and lost the predatory ability.
315 Some species are probably osmotrophic but, in general, their predatory impact on prokaryotes and
316 eukaryotes is negligible compared to that of heterotrophic euglenids (Leander et al., 2017).

317 Photosynthetic dinoflagellates are widely recognised as mixotrophic species, and are able to
318 display peculiar phagotrophic strategies (Saldarriaga & Taylor, 2017). According to Reynolds et al.
319 (2002), large thecate dinoflagellates are often associated to the coccalean cyanobacterium
320 *Microcystis* spp. This is probably due to the reduced light availability caused by the colonies of
321 *Microcystis* floating on the water surface, and to the possibility for these large mixotrophic
322 dinoflagellates to ingest single cells of the cyanobacterium as an additional source of carbon.

323 However, both euglenids and dinoflagellates were never important in the lake, even though
324 their representatives were present throughout the year. As for cryptophytes, no relationships were
325 found with other phytoplankton groups and environmental variables.

326 As regard strictly-phototrophic phytoplankton (i.e. “cholorococcalean” green algae and
327 diatoms) and cryptophytes, these were able to reach quite high biomass value any time there was a
328 decrease in the relative biomass of haptophytes. This would suggest that these algae may benefit
329 from the legacy of nutrients released in the water by haptophytes and deriving from their
330 heterotrophic mode of nutrition.

331 In conclusion, it can be stressed that *Prymnesium parvum*, thanks to its mixotrophic
332 behaviour and toxicity is able to monopolise resources and exert a negative effect on all the other
333 biological components of the ecosystem. However, to do that, the species needs a continuous
334 support in organic matter and bacteria. Lacking this “fuel”, which positively influences its growth,
335 the species is rapidly outcompeted by phototrophic organisms which can rapidly grow up thanks to

336 the legacy of nutrients left by *Prymnesium* itself. The alternation between mixotrophic and
337 phototrophic organisms is therefore a way to make a virtue out of necessity, and allow the entire
338 phytoplankton assemblage of the lake to quickly access both organic and inorganic nutrient
339 resources.

340

341 **References**

- 342 APAT, 2003. Metodi Analitici per le Acque. Manuali e Linee Guida 29/2003. Agenzia per la
343 Protezione dell’Ambiente e per i Servizi Tecnici, Roma. ISBN 88-448-0083-7. Available online
344 at [http://www.isprambiente.gov.it/it/pubblicazioni/manuali-e-linee-guida/metodi-analitici-per-](http://www.isprambiente.gov.it/it/pubblicazioni/manuali-e-linee-guida/metodi-analitici-per-le-acque)
345 [le-acque](http://www.isprambiente.gov.it/it/pubblicazioni/manuali-e-linee-guida/metodi-analitici-per-le-acque)
- 346 Arenovski, A.L., E.L. Lim & D.A. Caron, 1995. Mixotrophic nanoplankton in oligotrophic surface
347 waters of the Sargasso Sea may employ phagotrophy to obtain major nutrients. *Journal of*
348 *Plankton Research* 17: 801–820.
- 349 Azam, F., T. Fenchel, J.G. Field, J.S. Gray, L.A. Meyer-Reil & F. Thingstad, 1983. The ecological
350 role of water-column microbes in the sea. *Marine Ecology Progress Series* 10: 257–263.
- 351 Barone, R. & L. Naselli-Flores, 2003. Distribution and seasonal dynamics of Cryptomonads in
352 Sicilian water bodies. *Hydrobiologia* 502: 325-329.
- 353 Barone, R., G. Castelli & L. Naselli-Flores, 2010. Red sky at night cyanobacteria delight: the role of
354 climate in structuring phytoplankton assemblage in a shallow, Mediterranean lake (Biviere di
355 Gela, southeastern Sicily). *Hydrobiologia* 639: 43–53.
- 356 Burkholder, J.M., P.M. Glibert & H.M. Skelton, 2008. Mixotrophy, a major mode of nutrition for
357 harmful algal species in eutrophic waters. *Harmful Algae* 8: 77–93.
- 358 Caron, D.A., R.W. Sanders, E.L. Lim, C. Marrasé, L.A. Amaral, S. Whitney, R.B. Aoki & K.G.
359 Porter, 1993. Light-dependent phagotrophy in the freshwater mixotrophic crysophyte
360 *Dinobryon cylindricum*. *Microbial Ecology* 25: 93-111.

361 Croft, M.T., M.J. Warren & A.G. Smith, 2006. Algae need their vitamins. *Eukaryotic Cell* 5: 1175-
362 1183.

363 Eiler, A., 2006. Evidence for the ubiquity of mixotrophic bacteria in the upper ocean: implications
364 and consequences. *Applied and Environmental Microbiology* 72: 7431-7437.

365 Eikrem, W., L.K. Medlin, J. Henderiks, S. Rokitta, B. Rost, I. Probert, J. Throndsen & B.
366 Edvardsen, 2017. Haptophyta. In Archibald J.M., A.G.B. Simpson & C.H. Slamovits (eds),
367 *Handbook of the Protists*, 2nd Edition. Springer International Publishing AG, Cham: 893-953.

368 Ghyoot, C., K.J. Flynn, A. Mitra, C. Lancelot & N. Gypens, 2017. Modeling plankton mixotrophy:
369 a mechanistic model consistent with the Shuter-type biochemical approach. *Frontiers in*
370 *Ecology and Evolution* 5: Article 78.

371 Granéli, E. & N. Johansson, 2003. Increase in the production of allelopathic substances by
372 *Prymnesium parvum* cells grown under N- or P-deficient conditions. *Harmful Algae* 2: 135-
373 145.

374 Händeler, K., Y.P. Grzybowski, P.J. Krug & H. Wägele, 2009. Functional chloroplasts in
375 metazoan cells - a unique evolutionary strategy in animal life. *Frontiers in Zoology* 6: Article
376 28.

377 Hillebrand, H., C.-D. Dürselen, D. Kirschtel, U. Pollinger & T. Zohary, 1999. Biovolume
378 calculation for pelagic and benthic microalgae. *Journal of Phycology* 35: 403-424.

379 Hoef-Hemden, K. & J.M. Archibald, 2017. Cryptophyta (Cryptomonads). In Archibald J.M.,
380 A.G.B. Simpson & C.H. Slamovits (eds), *Handbook of the Protists*, 2nd Edition. Springer
381 International Publishing AG, Cham: 851-891

382 Jeppesen, E., S. Brucet, L. Naselli-Flores, E. Papastergiadou, K. Stefanidis, T. Nöges, P. Nöges, J.
383 L. Attayde, T. Zohary, J. Coppens, T. Bucak, R. F. Menezes, F. R. S. Freitas, M. Kernan, M.
384 Søndergaard & M. Beklioğlu, 2015. Ecological impacts of global warming and water
385 abstraction on lakes and reservoirs due to changes in water level and related changes in salinity.
386 *Hydrobiologia* 750: 201-227.

387 Jones, H.J.L., 1997. A classification of mixotrophic protists based on their behaviour. *Freshwater*
388 *Biology* 37: 35–43.

389 Jones, R.I., 2000. Mixotrophy in planktonic protists: an overview. *Freshwater Biology* 45: 219-226.

390 Jones, H., C.S. Cockell, C. Goodson, N. Price, A. Simpson & B. Thomas, 2009. Experiments on
391 mixotrophic protists and catastrophic darkness. *Astrobiology* 9: 563–571.

392 Johnson, M.D., D.J. Beaudoin, A. Laza-Martinez, S.T. Dyhrman, E. Fensin, S. Lin, A. Mercurief, S.
393 Nagai, M. Pompeu, O. Setälä & D.K. Stoecker, 2016. The Genetic Diversity of *Mesodinium*
394 and Associated Cryptophytes. *Frontiers in Microbiology* 7: Article 2017.

395 Kagalou, I., G. Tsimarakis & E. Bezirtzoglou, 2002. Inter-relationships between Bacteriological and
396 Chemical Variations in Lake Pamvotis – Greece. *Microbial Ecology in Health and Disease* 14:
397 37-41.

398 Keeling, P.J., 2013. The number, speed, and impact of plastid endosymbioses in eukaryotic
399 evolution. *Annual Review of Plant Biology* 64: 583-607.

400 Leander, B.S., G. Lax, A. Karnkowska & A.G.B. Simpson, 2017. Euglenida. In Archibald J.M.,
401 A.G.B. Simpson & C.H. Slamovits (eds), *Handbook of the Protists*, 2nd Edition. Springer
402 International Publishing AG, Cham: 1047-1088.

403 Litchman, E. & C.A. Klausmeier, 2008. Trait-based community ecology of phytoplankton. *Annual*
404 *Review of Ecology, Evolution, and Systematics* 39: 615-639.

405 Mitra, A., K.J. Flynn, U. Tillmann, J.A. Raven, D. Caron, D.K. Stoecker, F. Not, P.J. Hansen, G.
406 Hallegraeff, R. Sanders, S. Wilken, G. McManus, M. Johnson, P. Pitta, S. Våge, T. Berge, A.
407 Calbet, F. Thingstad, H.J. Jeong, J.-A. Burkholder, P.M. Glibert, E. Granéli & V. Lundgren,
408 2016. Defining Planktonic Protist Functional Groups on Mechanisms for Energy and Nutrient
409 Acquisition: Incorporation of Diverse Mixotrophic Strategies. *Protists* 167: 106-120.

410 Naselli-Flores, L., 1999. Limnological aspects of Sicilian reservoirs: a comparative ecosystemic
411 approach. In Tundisi, J. G. & M. Straškraba (eds), *Theoretical Reservoir Ecology and its*
412 *Applications*. Backhuys Publishers, Leiden: 283–311.

- 413 Naselli-Flores, L., 2000. Phytoplankton assemblages in twenty-one Sicilian reservoirs: relationships
414 between species composition and environmental factors. *Hydrobiologia* 424: 1-11.
- 415 Naselli-Flores, L., 2014. Morphological analysis of phytoplankton as a tool to assess ecological
416 state of aquatic ecosystems: the case of Lake Arancio, Sicily, Italy. *Inland Waters* 4: 15–26.
- 417 Naselli-Flores, L. & R. Barone, 1994. Relationship between trophic state and plankton community
418 structure in 21 Sicilian dam reservoirs. *Hydrobiologia* 275/276: 197-205.
- 419 Naselli-Flores, L. & R. Barone, 2011. Fight on plankton! Or, phytoplankton shape and size as
420 adaptive tools to get ahead in the struggle for life. *Cryptogamie Algologie* 32: 157–204.
- 421 Naselli-Flores, L. & R. Barone, 2012. Phytoplankton dynamics in permanent and temporary
422 Mediterranean waters: is the game hard to play because of hydrological disturbance?
423 *Hydrobiologia* 698: 147–159.
- 424 Naselli-Flores, L., R. Termine & R. Barone, 2016. Phytoplankton colonization patterns. Is species
425 richness depending on distance among freshwaters or on their connectivity? *Hydrobiologia* 764:
426 103-113.
- 427 Obrador, B. & J.L. Pretus, 2008. Light regime and components of turbidity in a Mediterranean
428 coastal lagoon. *Estuarine Coastal and Shelf Science* 77: 123-133.
- 429 Olutiola, P.O., K. O. Awojobi, O. Oyedeji, A.D.V. Ayansina & O.O. Cole, 2010. Relationship
430 between bacterial density and chemical composition of a tropical sewage oxidation pond.
431 *African Journal of Environmental Science and Technology* 4: 595-602.
- 432 Pállson, C. & W. Granéli, 2004. Nutrient limitation of autotrophic and mixotrophic phytoplankton
433 in a temperate and tropical humic lake gradient. *Journal of Plankton Research* 26: 1005-1014.
- 434 Raven, J.A., 1997. Phagotrophy in phototrophs. *Limnology and Oceanography* 42: 198–205.
- 435 Rengefors, K., A. Kremp, T.B.H. Reusch & M. Wood, 2017. Genetic diversity and evolution in
436 eukaryotic phytoplankton: revelations from population genetic studies. *Journal of Plankton*
437 *Research* 39: 165-179.
- 438 Reynolds, C.S., 2006. *The Ecology of Phytoplankton*. Cambridge University Press, Cambridge.

439 Reynolds, C.S., V. L. Huszar, C. Kruk, L. Naselli-Flores & S. Melo, 2002. Towards a functional
440 classification of the freshwater phytoplankton. *Journal of Plankton Research* 24: 417–428.

441 Roelke, D.L., A. Barkoh, B.W. Brooks, J.P. Grover, K.D. Hambright, J.W. LaClaire II, P.D.R.
442 Moeller & R. Patino, 2016. A chronicle of a killer alga in the west: ecology, assessment, and
443 management of *Prymnesium parvum* blooms. *Hydrobiologia* 764: 29-50.

444 Saldarriaga, J.F. & F.J.R.M. Taylor, 2017. Dinoflagellata. In Archibald J.M., A.G.B. Simpson &
445 C.H. Slamovits (eds), *Handbook of the Protists*, 2nd Edition. Springer International Publishing
446 AG, Cham: 625-678.

447 Simpson, A.G.B., C.H. Slamovits & J.M. Archibald, 2017. Protist Diversity and Eukaryote
448 Phylogeny. In Archibald J.M., A.G.B. Simpson & C.H. Slamovits (eds), *Handbook of the*
449 *Protists*, 2nd Edition. Springer International Publishing AG, Cham: 1-21.

450 Stoecker, D.K., 1998. Conceptual models of mixotrophy in planktonic protists and some ecological
451 and evolutionary implications. *European Journal of Protistology* 34: 281–290.

452 Tanifuji, G. & N.T. Onodera, 2017. Cryptomonads: A model organism sheds light on the
453 evolutionary history of genome reorganization in secondary endosymbiosis. *Advances in*
454 *Botanical Research* 84: 263-320.

455 Tartari, G. A. & R. Mosello, 1997. Metodologie analitiche e controlli di qualità nel laboratorio
456 chimico dell’Istituto Italiano di Idrobiologia. *Documenta dell’Istituto Italiano di Idrobiologia*
457 60: 1–160.

458 Tranvik, L.J., K.G. Porter & J.M. Sieburth, 1989. Occurrence of bacterivory in *Cryptomonas*, a
459 common fresh-water phytoplankter. *Oecologia*, 78: 473–476.

460 Yafremava, L., M. Wielgos, S. Thomas, A. Nasir, M. Wang, J.E. Mitthenthal & G. Caetano-
461 Anollés, 2013. A general framework of persistence strategies for biological systems helps
462 explain domains of life. *Frontiers in Genetics* 4: Article 16.

463 Zimmer, C., 2009. On the Origin of Eukaryotes. *Science* 325: 666-668.

464

Figure Captions

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467 Fig. 1. Seasonal trends of depth, Biochemical Oxygen Demand (BOD5) values, and *Prymnesium*
468 *parvum* biomass in the Biviere di Gela during the study period.

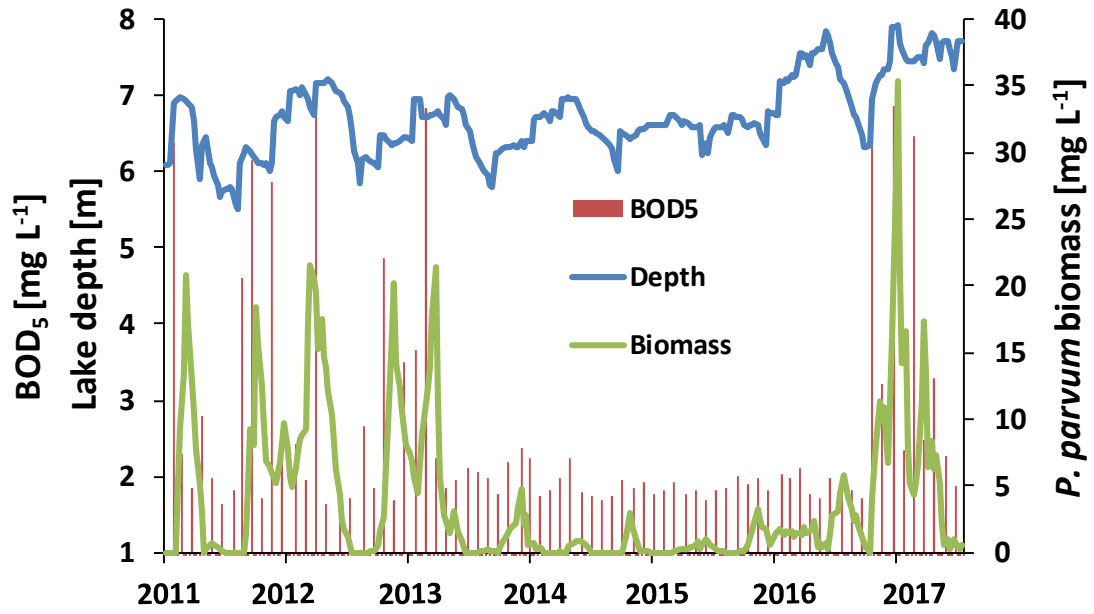
469 Fig. 2. Trends of inorganic nutrients (RP: reactive phosphorus; DIN: Dissolved Inorganic Nitrogen)
470 in the Biviere di Gela during the study period. The dotted lines represent the best fitting
471 polynomial line.

472 Fig. 3. Trends of phytoplankton biomass in the Biviere di Gela during the study period. HAP:
473 Haptophytes, CYA: Cyanobacteria.

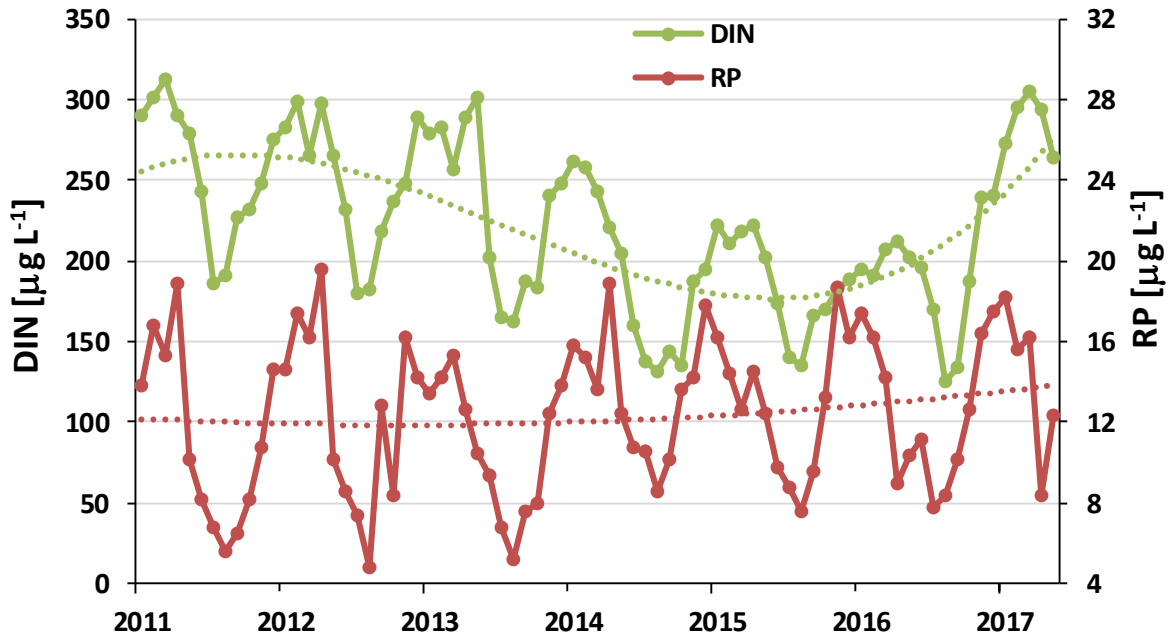
474 Fig. 4. Relative contribution of mixotrophic and phototrophic phytoplankton to total biomass in the
475 Biviere di Gela during the study period.

476 Fig. 5. Relationship between weekly *Prymnesium parvum* growth rates and the increment in depth
477 recorded in the Biviere di Gela one week earlier. Water incomes from Lake Dirillo are
478 seasonally differentiated by colours: red dots refer to winter growth, green dots refer to
479 spring growth, yellow dots refer to autumn growth and dark blue refer to summer growth.
480 Light blue dots correspond to “natural” water incomes and are related to growth rates lower
481 than 0.05 d^{-1} .

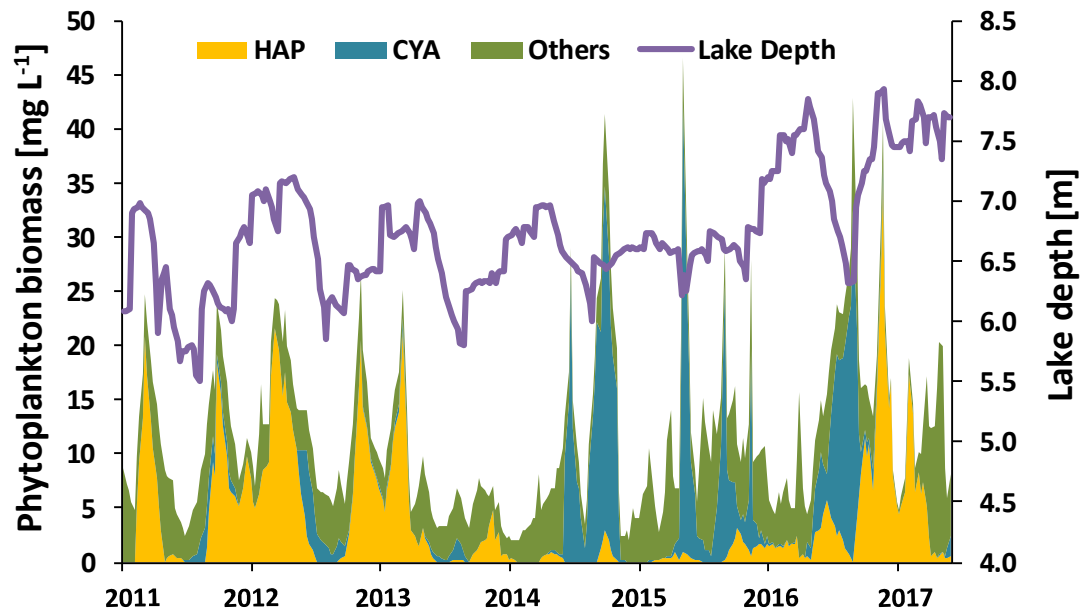
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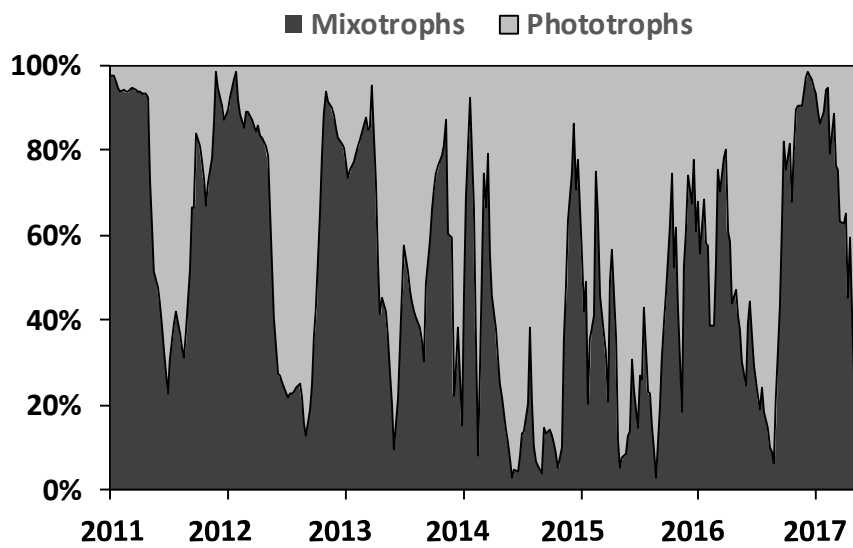
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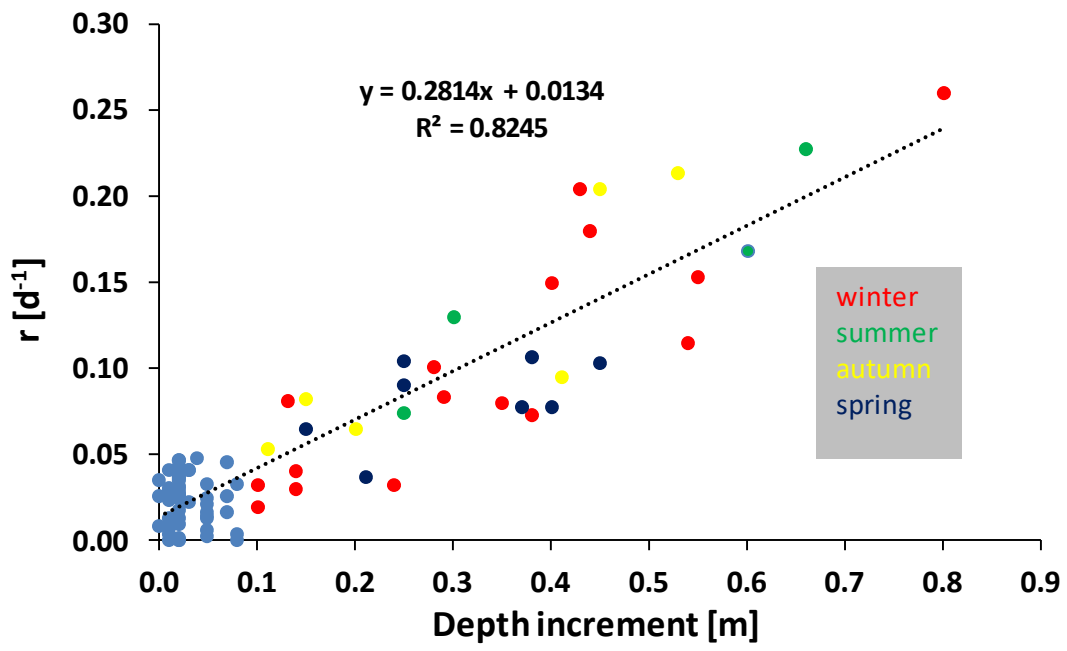


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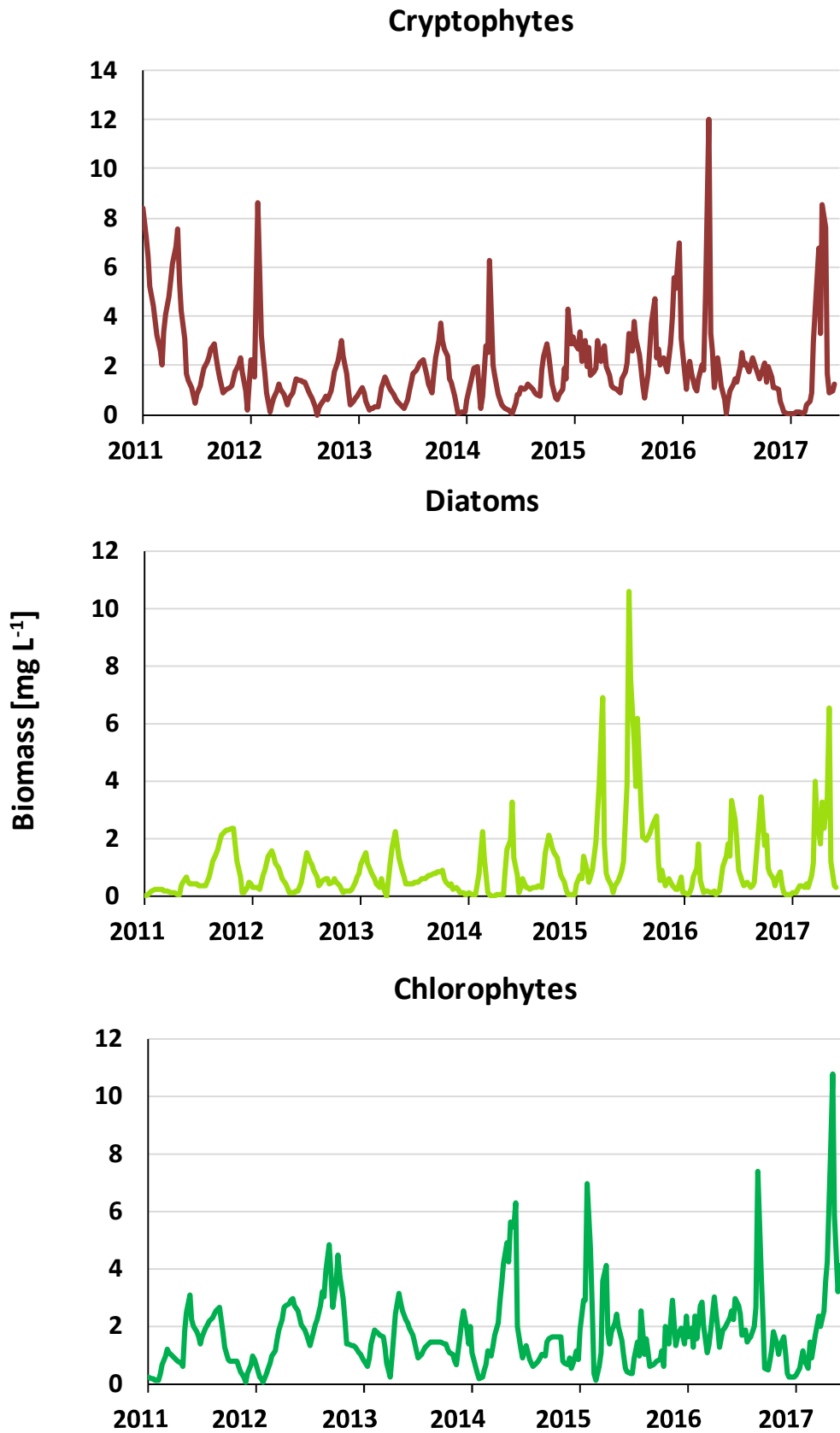
499 Fig. 5

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Electronic Supplementary Material

502 Biomass trends of the main phytoplankton groups in the Biviere di Gela during the study period.



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