

Life Sciences

Effects of Oleuropein on tumor cell growth and bone remodeling: Potential implications for the prevention and treatment of malignant bone diseases --Manuscript Draft--

Manuscript Number:	LFS-D-20-04499R2
Article Type:	Review article
Keywords:	Bone; Cancer; Chemoprevention; Metastasis; Oleuropein; Polyphenols; Tumor progression
Corresponding Author:	G Leto ITALY
First Author:	Gaetano Leto
Order of Authors:	Gaetano Leto Carla Flandina Marilena Crescimanno Marco Giammanco Maria Vittoria Sepporta
Manuscript Region of Origin:	ITALY
Abstract:	<p>Oleuropein (Ole) is the main bioactive phenolic compound present in olive leaves, fruits and olive oil. This molecule has been shown to exert beneficial effects on several human pathological conditions. In particular, recent preclinical and observational studies have provided evidence that Ole exhibits chemo-preventive effects on different types of human tumors. Studies undertaken to elucidate the specific mechanisms underlying these effects have shown that this molecule may thwart several key steps of malignant progression, including tumor cell proliferation, survival, angiogenesis, invasion and metastasis, by modulating the expression and activity of several growth factors, cytokines, adhesion molecules and enzymes involved in these processes. Interestingly, experimental observations have highlighted the fact that most of these signalling molecules also appear to be actively involved in the homing and growth of disseminating cancer cells in bones and, ultimately, in the development of metastatic bone diseases. These findings, and the experimental and clinical data reporting the preventive activity of Ole on various pathological conditions associated with a bone loss, are indicative of a potential therapeutic role of this molecule in the prevention and treatment of cancer-related bone diseases. This paper provides a current overview regarding the molecular mechanisms and the experimental findings underpinning a possible clinical role of Ole in the prevention and development of cancer-related bone diseases.</p>



Elsevier
Radarweg 29
Amsterdam 1043 NX,
elsevier.com

Empowering Knowledge

Life Sciences

Conflict of Interest Policy

Article Title:

Effects of Oleuropein on tumor cell growth and bone remodeling. Potential implication for the prevention and treatment of malignant bone diseases

Author Names:

Gaetano Leto

Declarations

Life Sciences require that the **corresponding author**, signs on behalf of all authors, a declaration of conflicting interests. If you have nothing to declare in any of these categories then this should be stated.

Conflict of Interest

A conflicting interest exists when professional judgment concerning a primary interest (such as patient's welfare or the validity of research) may be influenced by a secondary interest (such as financial gain or personal rivalry). It may arise for the authors when they have financial interest that may influence their interpretation of their results or those of others. Examples of potential conflicts of interest include employment, consultancies, stock ownership, honoraria, paid expert testimony, patent applications/registrations, and grants or other funding.

Please state any competing interests:

All authors of this paper declare that there is no conflict of interest related to the content of this manuscript

Funding Source

All sources of funding should also be acknowledged and you should declare any involvement of study sponsors in the study design; collection, analysis and interpretation of data; the writing of the manuscript; the decision to submit the manuscript for publication. If the study sponsors had no such involvement, this should be stated.

Please state any sources of funding for your research:

No funds



Elsevier
Radarweg 29
Amsterdam 1043 NX,
elsevier.com

Empowering Knowledge

Author Contribution to Study

All authors listed on your paper must have made significant contributions to the study. To ensure clarity, you are required to enter the specific details of each author's contribution, which must substantiate the inclusion of each person on the manuscript. Please detail this information below (submit additional sheets as necessary):

Author Name	Author Email	Specific Role in Study
Gaetano Leto	gaetano.letto@unipa.it	conception, design, acquisition and interpretation of data
Carla Flandina	carla.flandina@unipa.it	design acquisition of the data, critical revision of the content. Final approval of the MS
Marilena Crescimanno	marilena.crescimanno@unipa.it	acquisition of the data and final approval of the MS
Marco Giammanco	marco.giammanco@unipa.it	acquisition and critical revision of the data; final approval of the MS
Maria Vittoria Sepporta	vittoriasepporta@gmail.com	Conception, design, acquisition of the data; inal approval of the MS

Signature

Print name

G a e t a n o L e t t o _____

Gaetano Leto _____

Prof. L.E. Wold
Editor-in -Chief
Life Sciences

Palermo 10/19/2020

Dear Professor Wold,

Enclosed, please find the last revised versions of our review article “***Effects of Oleuropein on tumor cell growth and bone remodelling...***” (Your ref Ms.No. LFS-D-20-04499R1) which is being submitted for consideration for publication in Life Sciences.

Enclosed also find :

1. An additional file of the revised paper (“marked version”) reporting the changes made throughout the text, and which are highlighted in yellow according to the editorial and reviews’ suggestions.
2. A file containing the Response to Reviewers

As suggested by the Publisher, we have included the names of up to 5 chemical compounds which recur in the article.

Please let me know if you need any further assistance.

Thank you for your time and consideration in this matter.

Best regards

Sincerely Yours

Gaetano Leto

Response to Reviewers

REVIEWERS' COMMENTS:

Reviewer #1

The authors made the corrections, which increased the clarity of the manuscript. The tables and Figure 3 integrate the information about Ole's studies very well.

The observations are minimal to the format:

Q. Correctly write the name of the species: it says *Lactobacillus Plantarum* must be *Lactobacillus plantarum*)
Section 4:

a) ... tumor cell in the bone tissue [20,103,104] (Fig. 3) .. On the other hand ...

There is a point left

b) emodelling processes (Fig. 2, Fig. 3). the point is in bold

Format:

4.2. Oleuropein and Epithelial to Mesenchymal transition (EMT).

Use the same format for the arrows in the tables (1, 2 and 3)

In table 1:

In vivo instead of *In vivo* (in italics)

Says *OLe* instead of *Ole*

Explain the symbology (arrows)

In table 2:

In vitro instead of *In vitro* (in italics)

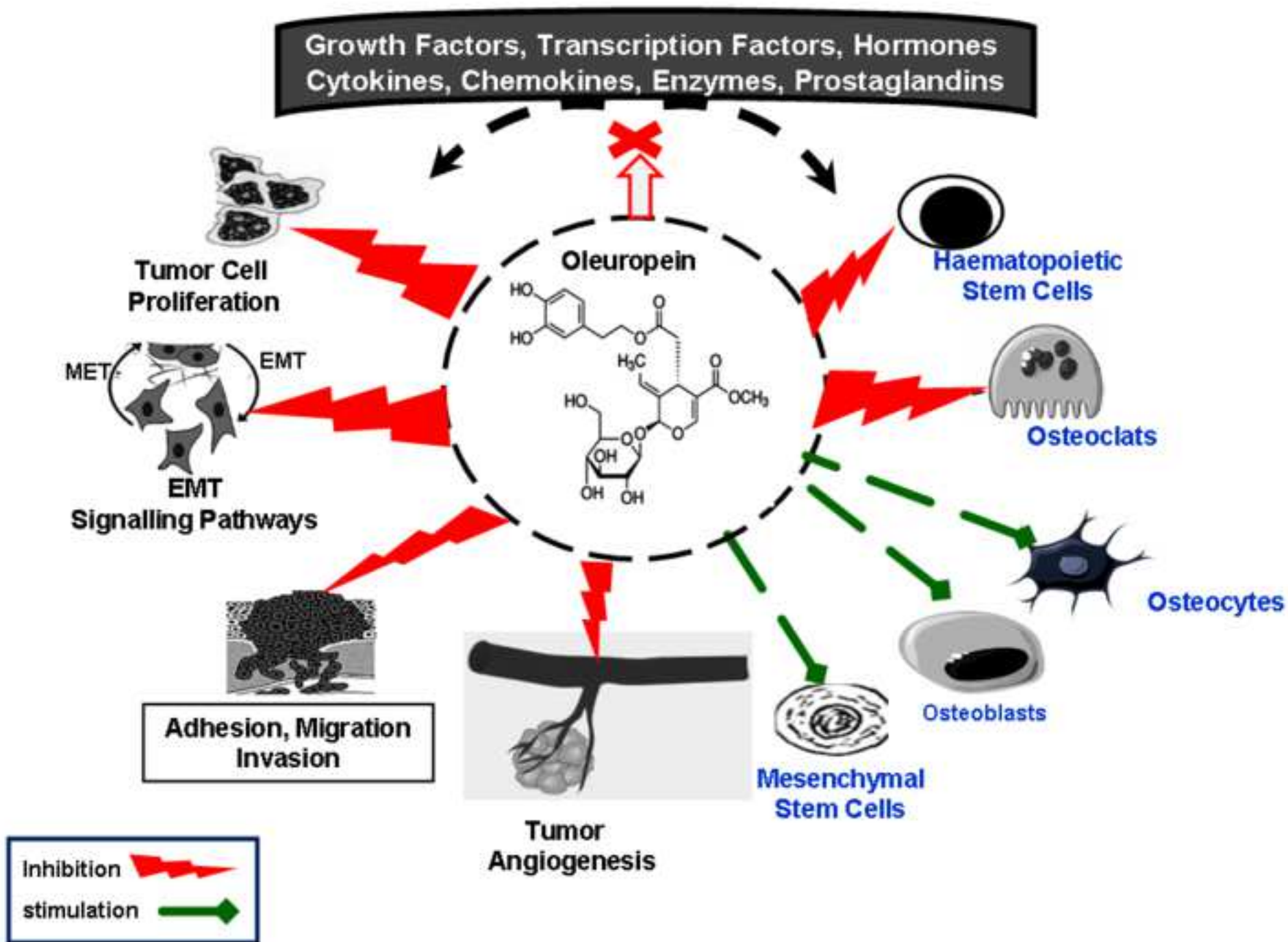
Some references are in red letters

Explain the symbology (arrows)

R. We provided to correct typo and to make the appropriate changes indicated by the review where required. These corrections have been highlighted in yellow in the marked version.

Tables and figures have been revised accordingly

As suggested by the Publisher, we have enriched the text by including the names of up to 5 chemical compounds (highlighted in yellow in the marked version) which recur in the article



Word count 10967 ; Figures 3; Tables 3.;

Effects of Oleuropein on tumor cell growth and bone remodeling: Potential clinical implications for the prevention and treatment of malignant bone diseases

Gaetano Leto^{a*}, Carla Flandina^a, Marilena Crescimanno^a, Marco Giammanco^b and Maria Vittoria Sepporta^c

^a*Laboratory of Experimental Pharmacology, Department of Health Sciences, University of Palermo, 90127 Palermo, Italy.*

^b*Department of Surgical, Oncological and Oral Sciences University of Palermo, 90127 Palermo, Italy.*

^c*Pediatric Unit, Department Women-Mother-Children, Pediatric Hematology-Oncology Research Laboratory, Lausanne University Hospital, Lausanne, Switzerland.*

*Corresponding author: Gaetano Leto, Laboratory of Experimental Pharmacology, Department of Health Sciences, University of Palermo, 90127 Palermo, Italy. e-mail, gaetano.letto@unipa.it; Orcid:0000-0002-0900-1259

Abstract

Oleuropein (Ole) is the main bioactive phenolic compound present in olive leaves, fruits and olive oil. This molecule has been shown to exert beneficial effects on several human pathological conditions. In particular, recent preclinical and observational studies have provided evidence that Ole exhibits chemo-preventive effects on different types of human tumors. Studies undertaken to elucidate the specific mechanisms underlying these effects have shown that this molecule may thwart several key steps of malignant progression, including tumor cell proliferation, survival, angiogenesis, invasion and metastasis, by modulating the expression and activity of several growth factors, cytokines, adhesion molecules and enzymes involved in these processes. Interestingly, experimental observations have highlighted the fact that most of these signalling molecules also appear to be actively involved in the homing and growth of disseminating cancer cells in bones and, ultimately, in the development of metastatic bone diseases. These findings, and the experimental and clinical data reporting the preventive activity of Ole on various pathological conditions associated with a bone loss, are indicative of a potential therapeutic role of this molecule in the prevention and treatment of cancer-related bone diseases. This paper provides a current overview regarding the molecular mechanisms and the experimental findings underpinning a possible clinical role of Ole in the prevention and development of cancer-related bone diseases.

Keywords: Bone; Cancer; Chemoprevention; Metastasis; Oleuropein; Polyphenols; Tumor progression

Chemical compounds: Elenolic Acid (PubChem CID: 169607); Hydroxytyrosol (PubChem CID: 82755); Oleuropein (PubChem CID: 5281544); Oleuropein aglycone (Pubchem CID: 56842347); Tyrosol (PubChem CID: 10393)

Abbreviations

BCa, Breast Cancer

BMSCs, Bone Marrow Mesenchymal Stem Cells

BMP, Bone Morphogenic Protein

BPH, Benign prostate hyperplasia epithelial cells

COX-2, Cyclooxygenase-2

CSC, Cancer Stem Cells

CXCRs, CXC Chemokine receptors

Elac, Elenolic Acid

ELAM-1 endothelial-leukocyte adhesion molecule 1

ECM, Extracellular matrix

EMT, Epithelial Mesenchymal Transition

EVOO, Extra Virgin Olive Oil

γ -GCS, γ -Glutamylcysteine Synthetase

Glo2, Mitochondrial Glyoxalase 2

HDAC2, Histone deacetylase 2

HDAC3, Histone deacetylase 3

HIF, Hypoxia Inducible Factor

HO-1, Heme Oxygenase-1

Hyt, Hydroxytyrosol

ICAM-1, Intercellular Adhesion Molecule-1

i-NOS, Inducible Form Of Nitric Oxide Synthase

IL-1 β , Interleukin-1 β

IL-6, Interleukin-6

IL-8, Interleukin-8

MAPK, Mitogen Activated Protein-Kinase

MCP-1, Monocyte Chemo-Attractant Protein-1

MMPS, Matrix Metalloproteinases

MMP-2, Matrix Metalloproteinases

MMP-9, Matrix Metalloproteinases

MM, Multiple Myeloma

mTOR, Mammalian Target Of Rapamycin

NF-kB, Nuclear Factor kB

NSLC, Non Small Cell Lung Cancer

OLE, Oleuropein.

OLE-A, Oleuropein Aglycone

OB, Osteoblast

OCL, Osteoclast

OPG/RANKL, Osteoprotegerin/Receptor Activator Of Nuclear Factor Kappa-B Ligand Expression Ratio

OS, Osteosarcoma

PARP, Poly (ADP-Ribose) Polymerase
 PCa, Prostate Cancer
 PTPB1, Protein Tyrosine Phosphatase B1
 PTHrP, Parathyroid hormone-related protein
 PGs, Prostaglandins
 PI3K/AKT, Phosphatidylinositol-3 Kinase/ Protein Kinase B
 PPAR γ , Peroxisomal Proliferator-Activated Receptor
 RANK-L, Receptor Activator of NF- κ B Ligand
 RNI, Reactive Nitrogen Intermediates
 ROS, Reactive Oxygen Species
 RUNX2, Runt-Related Transcription Factor 2
 SOD2, Superoxide Dismutase 2
 STAT3, Signal Transducer and Activators of Transcription 3
 TIMP, Tissue Inhibitor of Metallproteinase
 TNF- α , Tumor Necrosis Factor- α
 TGF- β , Transforming Growth Factor β ,
 TRAP, Tartrate-Resistant Acid Phosphatase
 VEGF, Vascular Endothelial Growth Factor;
 VEGFR, VEGF Receptor
 VGSCs blocking voltage-gated sodium channels
 V-CAM-1, Vascular Cellular Adhesion Molecule-1
 Wnt/ β -Catenin, Wingless-Related Integration Site/ B-Catenin

1. Introduction

The skeleton is the third most common site of metastatic disease after lung and liver [1]. Many types solid cancers, in particular, breast, prostate, thyroid, lung, and kidney tumors preferentially metastasize to bone [1,2]. The incidence of bone metastasis by tumor type, reaches 65–90% in prostate cancer (PCa), about 65–75% in breast cancer (BCa), 60% in thyroid cancer, 30-40% in lung cancer, 40% in bladder, cancer, 20-25% in renal cell carcinoma. Skeletal involvement is less frequent in other malignancies such as melanoma (11-17%) and colorectal tumors (10%) [2,3]. The median-survival from diagnosis of bone metastasis from breast cancer, prostate cancer, thyroid cancer are 27 months, 25 months, and 23 months, respectively while, in patients with renal, bladder, lung, and melanoma median survival is lower (12 months, 8 months, 7-9 months and 6 months respectively) [2-4]. Furthermore, in some haematological malignancies such as in multiple myeloma (MM) the rate of incidence reaches 70–95% [4,5]. Interestingly, MM patients diagnosed after 2010 experienced improved clinical outcomes, the 2-year survival rate having increased from 69.9% in 2006 to 87.1% in 2012 [5,6]. On the other hand, primary malignant bone tumors are relatively rare, occurring at a rate of about one to 100,000 [4,5]. In adults, over 40% of primary bone cancers are chondrosarcomas. This is

followed by osteosarcomas (28%), chordomas (10%), Ewing tumors (8%), and malignant fibrous histiocytoma and fibrosarcomas (4%). The remainder of cases are several rare types of bone cancer [4,5]. In children and teenagers (those younger than 20 year), osteosarcoma (56%) and Ewing's sarcoma (34%) are much more common than chondrosarcoma (6%) [4,5]. The clinical treatment of primary malignant bone tumors or bone metastasis is currently based on therapeutic approaches encompassing systemic treatments, including chemotherapy, hormone therapy, immunotherapy, bone modifying agents, radiopharmaceuticals, and local treatments such as radiation therapy, radiofrequency ablation and surgery [7,8]. Unfortunately, to date, none of these therapeutic options have shown to exert a positive clinical impact on patients' survival [7,8]. Furthermore, the systemic administration of antitumor drugs and/or bone modifying agents and/or radiopharmaceuticals may negatively affect the normal metabolic turnover of bone tissue with detrimental consequences for cancer patients [9,10]. Hence, the need to identify new compounds that may be effective in preventing the growth and dissemination of malignant cells in the bone and, at the same time, are endowed with low toxicity and/or limited side effects [7,8,11]. In recent years an increasing number of preclinical studies have been directed towards the discovery and the development of new drugs based on natural products from plants environment [12-17]. The advantages of using natural products for therapeutic purposes in cancer treatment are manifold, as these compounds appear, in general, to be *i*) readily available, *ii*) non-toxic to normal human cells, *iii*) may act as multi-target agents since they may affect different signalling pathways that control cancer progression [12-18]. Among the natural compounds, phytochemicals such as polyphenols, terpenoids, alkaloids, phytosterols, and organosulfur compounds have been reported to produce beneficial effects on human health and, in particular, in pathological conditions such as inflammation and cancer [12-14]. In this setting, there is growing experimental evidence highlighting the fact that several extra virgin olive oil (EVOO)-derived phenolic compounds are endowed with anti-proliferative, anti-invasive and anti-metastatic properties [14,17-20]. Therefore, these molecules seem to be promising as potential chemo-preventive agents. In particular, oleuropein (OLE) (Fig.1), one of the main bioactive phenolic compounds present in olive leaves (*Olea europaea* L., Oleaceae), in unprocessed olive drupes and, in the aglycone form, in olive-oil exhibits a broad range of pharmacological properties [17,20-22], which may account for the therapeutic effects of this molecule observed in various human pathological conditions [14,18,20-22] including chronic inflammatory diseases associated with bone loss [20-22] and human cancers [17-19,23-25]. The finding that OLE may thwart the dissemination of cancer cells to distant organs [18,21-23] and exercises protective effects against the onset of various pathological conditions associated with bone loss [20-22] suggests the potential clinical usefulness of this molecule in the prevention and treatment of cancer-related bone diseases. In this review we discuss and critically appraise, in particular, the results from emergent data suggesting a potential role for OLE in the chemoprevention and clinical management of malignant bone tumors.

2. Oleuropein in human tumors

An increasing number of *vitro* studies have shown that OLe may inhibit the growth and survival of cancer cells from different human solid tumors including breast cancer [18,20,22-25] colorectal cancer [20,23-25], genito-urinary cancer [20,23-26], lung cancer [19,23-25], central nervous system tumors [19,20,23-25] melanoma [19,27,28], and haematological malignancies such as leukemias [19,20,29,30]. In line with these observations, *in vivo* studies have highlighted the fact that the administration of OLe to experimental animals may hinder the development of tumors such as skin cancer [31], soft tissue sarcoma [32], melanoma [27,28] or breast cancer [33-36] (Table 1) In particular, the administration of OLe to mice transplanted with B16F10 melanoma or MCF-7 human breast cancer cells, resulted in a significant inhibition of lymph-node or lung metastasis respectively [33-36] (Table 1). The numerous studies carried out in order to elucidate the specific mechanisms underlying the antitumor effects of OLe have highlighted the fact that this molecule, besides its inhibiting activity on tumor cell proliferation and survival [23-26], may also hinder other key steps of malignant progression such as cell migration [16,19,20,24,32,37-40], invasion[19,20,24,32,39,40,43], angiogenesis [20,24, 32,33,44,45] and dissemination of malignant cells to distant organs [15,20,24-26,28,32-36,46,47]. (Fig. 2) In this setting, molecular studies have revealed that OLe can modulate the expression and activity of several growth factors, hormones, cytokines, adhesion molecules and enzymes, which regulate different cellular processes involved in cancer progression including apoptosis [20,24,33,39,42,43,49,50], cell cycle progression [20,26,43,44,50], cell differentiation [20,24,30,51] and epithelial-to-mesenchymal transition (EMT) [20,37,52-56] (Fig. 2). These findings indicate the therapeutic potential of OLe as a chemo-preventive drug that might thwart early events in the metastatic process [24-28,35,36,44-48] (Fig. 2). However, at least *in vivo*, the probable contribution of a few active metabolites of OLe, endowed with anticancer activity such as hydroxytyrosol (Hyt), and OLe-aglycone (Ole A) (Fig. 1) may further account for the anticancer activity of this molecule [19,20,23-25,51,57]. In fact, experimental and clinical pharmacokinetic studies have shown that following its absorption in the intestinal tract, Ole undergoes phase I and phase II metabolic processes that generate several metabolites endowed with various pharmacological effects including OLe-A and Hyt [58-61]. In this setting, Mosele et al. [62] have provided evidence on the involvement of some lactic acid bacteria which can be found in human gastrointestinal tract (in particular *Lactobacillus plantarum*) in the metabolic conversion of OLe in OLe-A and then into Hyt.

3. Oleuropein and the bone microenvironment

Experimental and observational studies have provided evidence on the effectiveness of OLe in the prevention and treatment of various pathological conditions associated with bone loss [36,63-72]. These effects appear to be related to

the modulating activity of this phenolic compound on multiple common signalling molecules underlying the pathogenesis of malignant and non-malignant bone diseases [7,47,68,71]. On the basis of these observations it appears conceivable to speculate about a potential clinical role of OLe as a drug that might prevent the growth and dissemination of cancer cells in the bone [18,20,38,45-47,54,55,63] (Fig 2). In this context, compelling evidence shows that the protective effects exhibited by OLe on bone health appear to be due to the antioxidant [20,21,57-60] anti-inflammatory and immune-modulating properties of this molecule [20-22,67-71]. The antioxidant effects of OLe have been attributed to its ability to prevent the production of reactive oxygen species (ROS) and/or to act as radical chain breakers, and/or as a metal ion chelator [20,21,72]. Instead, the anti-inflammatory and immune-modulatory activities of OLe recognize multiple mechanisms and involve the capability of this compound to modulate the expression and activity of signalling molecules, such as growth factors [20,41,66,67,72-75], transcription factors [18,20,38,71,75-80], hormones [76,81,81], cytokines, chemokines [20,21,55,69,71,73,75] and enzymes [20,43,44,71,75,77], which, in physiological conditions, regulate the normal metabolic turnover of bone tissue while their expression proves to be deregulated in various pathological conditions associated with altered bone remodelling processes [20,44,45,71,82-84]. In particular, preclinical studies have highlighted the fact that the anti-inflammatory and immunomodulatory effects of OLe are the result of its inhibitory activity on nuclear factor- κ B (NF- κ B) and mitogen activated protein-kinase (MAPk) signalling pathways [20,66,71,79,84,85]. These phenomena eventuate in an attenuated production of various downstream molecular effectors modulating the immune-inflammatory response, such as tumor necrosis factor- α (TNF- α) [20,68,79,80,85,86], interleukin-1 β (IL-1 β) [20-22,71,85,86], interleukin-6 (IL-6) [20-22,71,86-89], interleukin-8 (IL-8) [8,20,21,47,68,72,88], monocyte chemo-attractant protein-1 (MCP-1) [89], prostaglandins (PGs), in particular prostaglandin E₂ (PGE₂), and enzymes such as cyclooxygenase-2 (COX-2) [20,24,44,70,71,85,86,90], matrix metalloproteinases (MMPs) [20,21,44,45,51,71] and the inducible form of nitric oxide synthase (iNOS) [20,65,71,80,85]. On the other hand, these findings are in line with the results from experimental studies indicating that the bone-protective effects of OLe appear to be related mainly to its modulating activity on inflammatory signalling pathways rather than directly on bone metabolism [63-68]. Nevertheless, recent findings have shown that OLe may facilitate osteoblast (OB) proliferation and differentiation [66,82,91-93], whilst it suppresses osteoclastogenesis [64,65,82,89-91]. These phenomena were associated with an increase in the expression levels of osteogenic transcription factors such as runt-related transcription factor 2 (Runx2) and osterix, with an increased osteoprotegerin/receptor activator of nuclear factor kappa-B ligand (OPG/RANKL) expression ratio [91,94] and the up-regulation of other osteoblastic markers such as type I collagen, osteocalcin or alkaline phosphatase [67,77,94,95]. In particular, OLe elicits osteoprotective effects by fostering the differentiation of human bone marrow mesenchymal stem cells into osteoblasts [91,96] and by reducing tartrate-resistant acid phosphatase (TRAP) activity in osteoclasts

formed from mouse spleen cells [65]. Furthermore, recent *in vitro* studies have shown that OLe modulates osteoblastogenesis by suppressing the expression and activity of perioxosome proliferator-activated receptor γ (PPAR γ), which functions as a transcriptional regulator of adipocyte differentiation and inhibitor of OB [91,96-98].

4. Oleuropein and malignant bone disease progression

Most of the molecular pathways involved in pathological conditions associated with disorders of bone remodeling and which may be targeted by Ole, have also been shown to contribute to the homing and growth of cancer cells in the bone [20,75,99-103] (Fig 3). The findings that various chronic inflammatory bone diseases and bone-related cancers share common molecular mechanisms underlying their pathogenesis [20,75,83,99-103] may, in part, explain the reason why chronic inflammation is actually considered as one of the main pathological processes that may foster the homing and growth of tumor cell in the bone tissue [20,103,104] (Fig. 3). On the other hand, these observations are consistent with the results from several experimental studies showing that chronic inflammation may function as a promoting factor in the early stages of a malignant progression [101, 103-107] (Fig. 3). These data may also, partially, explain the findings that disturbances of the bone microenvironment, induced by osteoporosis and/or other immunoinflammatory bone diseases, appear to facilitate the growth and dissemination of some types of cancer cells in the bone tissue [106-113]. Ultimately, these observations indicate that OLe might well exert its potential chemopreventive and therapeutic effects on cancer-related bone disease through multiple mechanisms involved in early events of cancer progression and in the regulation of bone remodelling processes (Fig. 2, Fig. 3).

4.1. Oleuropein and tumor cell proliferation and survival

A consistent body of *in vitro* studies have demonstrated that OLe may hinder the proliferation, survival and migration of malignant cells from human primary bone tumor, such as osteosarcoma [22,38,63,114], and from human tumors which preferentially metastasize to the bone, such as breast cancer [20,23-25,38-40,45,49,115], prostate cancer [20,23-25,116,117] and lung cancer [23-25,117-120]. The mechanisms underlying the anti-proliferative effects of OLe on osteosarcoma cells still need to be fully understood [37,63,114,116]. Conversely, accumulating evidence shows that OLe can inhibit the growth and survival of human breast, prostate or lung cancer cell lines by *i*)delaying or halting different phases of cell cycle progression [20,26,119-123], *ii*)by activating different pro-apoptotic molecular signalling pathways [20,26,40,45,76,113,116-118,123-127] and/or *iii*)by inhibiting the expression of the transcription factor NF-kB which regulates many genes driving cancer growth and progression (Table 2).

4.2. Oleuropein and Epithelial to Mesenchymal transition (EMT) .

OLE has been demonstrated to modulate *in vitro* multiple signalling pathways associated with the different types of EMT, including type 3 EMT [36,46,51-54,119], which has been recognized as an initial, critical event for metastasis formation by individually-invading carcinoma cells [119,122,124,128-131]. Almost all the key regulators of EMT are expressed in the bone microenvironment where their cross-talk contributes to favouring the homing and growth of cancer cells [37,47,51-54,129-131] (Fig. 2, Table 3). In particular, these studies have highlighted the promoting role of signalling pathways such as TGF- β /BMP [52,74,132-134], MAP/ERK [52,120,125,134-136], PI3K/AKT [52,105,137], Wnt/ β -catenin [54-56,74,125-131], NF- κ B [128,133,137,140,141], HIF [36,142], and PTHrP [142,143] on the growth and dissemination of cancer cells in the bone (Table 3). OLE has been reported to preserve bone health by interfering with various cellular processes triggered by these molecular pathways [20,66,68,71,85,87,92-94,135]. On the basis of these findings, numerous preclinical investigations have been undertaken to assess the therapeutic effectiveness of this molecule, alone or in association with antitumor drugs in the prevention and treatment of several human solid tumors [13,15,43,45,50-53,71,111-113,134].

4.3. Oleuropein and tumor angiogenesis.

Tumor angiogenesis is a crucial event for the growth and dissemination of circulating tumor cells to distant organs including the bones [128-130]. Several experimental observations highlight the fact that OLE appears to thwart tumor neovascularization by decreasing the expression levels of various signaling molecules involved in this process [32,33,36,44, 89,118,123-129,144] (Fig. 2, Table 3). In particular, these investigations have shown that the treatment of endothelial cells [75,76,89,90,143,144] and/or human cancer cells from different tumors [34,35,76,124,125,143,145], with OLE results in a decrease in the expression level of specific growth factors and receptors such as VEGF and VEGFR2 [31,33,36,75,90,118,129], pro-inflammatory cytokines such as IL-6, IL-8, IL-1 β TNF- α [19,20,22,48,80,90,143-146], adhesion molecules such as vascular cellular adhesion molecule-1 (VCAM-1) intercellular adhesion molecule-1 (ICAM-1) and E-selectin [20,22,39,52,71,75,145,146], inflammatory and proteolytic enzymes such as COX-2 [31,39,44,71,75,98] and MMPs [31,37,44,45,52,71,75,118,130] (Table 3). Most of these effects can also be attributed to the direct up-stream inhibiting activity of OLE on NF- κ B, a transcription factor that modulates the expression of various genes and related signalling pathways involved in almost all the hallmarks of malignant progression [20,39,79,131,132, 143,147].

4.4. Oleuropein and tumor cell migration, adhesion and invasion.

Besides tumor neo-vascularization, some of the molecules mentioned above, such as pro-inflammatory cytokines, adhesion molecules or COX-2 and MMPs, also modulate the adhesion, migration and invasion of tumor cells [44,147-153] (Table 3). For instance, *in vitro* studies have demonstrated that adhesion molecules such as VCAM-1, ICAM-1 and E-selectin contribute to the escaping of tumor cells from dormancy and bone metastasis formation [146,150,152-154]. Moreover, emerging evidence has shown that OLe may indirectly affect the adhesive properties of tumor cells by inhibiting protein tyrosine phosphatase B1 (PTPB1), a regulator of cell adhesion and migration in normal and cancer cells [155,156]. Furthermore, MMP-2 and MMP-9 have been reported to actively contribute to the migratory activity of tumor cells, to the degradation of the extracellular matrix (ECM) and to regulating the expression of cytoskeletal proteins, growth factors, cytokines and chemokines, which favour the preferential adhesion of disseminating cancer cells to the ECM underlying epithelial cells and vascular endothelial cells [44,134,135,150,152]. Consistent with these observations, recent *in vitro* studies demonstrated that the and anti-invasive activity induced by OLe on tumor cells paralleled the down-regulation of MMP-9 and MMP-2 expression and the up-regulation of their intracellular inhibitors TIMP-1,-3,-4 [23,31,44,45,55,71,132,143,152] (Table 3)

4.5. Oleuropein and tumor-bone niche

Most of the signalling molecules mentioned above have also been shown to contribute to the formation of a permissive microenvironment, the so-called “(pre)metastatic niche”, which supports the homing, colonization and growth of cancer cells in the bone and, their subsequent survival, dormancy and resistance to clinical treatments [157-159] (Fig. 2, Fig.3, Table 3). Therefore, the targeting of these signalling molecules may be regarded as a novel potential therapeutic tool in the prevention and, eventually, eradication of the establishment of malignant cells in the bone. In this regard, preclinical evidence has suggested the possibility that OLe might thwart the formation of the (pre)metastatic bone niche (Fig. 2). For instance, pro-inflammatory interleukins (ILs) such as IL-6, IL-8, IL-1 β , TNF- α have been shown actively to contribute to the homing of malignant cells in the bone [19,20,90,143,147,160-163] (Fig. 3) Ole can down-regulate the expression of these cytokines in several pathological conditions associated with an altered bone resorption and in early events of the metastatic process, namely EMT transition [37,52-55,130,147] and angiogenesis [20,36,75,143,148] (Fig. 3, Table 3). Moreover, proteolytic enzymes such as MMP-2 and MMP-9 have also been shown to contribute to the formation of the “(pre)metastatic niche” by modifying the ECM structure and by promoting the release growth factors and chemokines which fuel the “vicious cycle” of bone metastasis [44,90,130-133,147,152] (Table 3). As described above, OLe has been proven to inhibit the expression levels of MMP-2 and MMP-9 in human metastatic tumor cells while it may increase the expression of intracellular inhibitors of these

enzymes, namely TIMP-1, TIMP-2 and TIMP-3 [45,51,54,75,132,152]. Moreover, OLe has been reported to indirectly affect the expression and activity of these proteolytic enzymes through the up-stream inhibition of NF- κ B [39,51,54,71,148,149,165] (Fig. 3). Furthermore, experimental evidence has shown that the osteo-protective effects of OLe also appear to be due to its inhibiting activity on RANKL-induced NF- κ B activation, a phenomenon which facilitates osteoclast differentiation and bone remodeling [68,70,93,94,161,166]. On the other hand, *in vitro* studies have highlighted the fact that the RANKL/RANK/OPG system appears to exert an active role in the formation of the pre-metastatic niche and in fostering the metastatic spread to bone [7,100,149,153,157,165,166]. On the basis of these observations it is conceivable to hypothesize a potential clinical role of OLe as a novel drug that may thwart the formation of a permissive microenvironment enabling tumor cells to grow and disseminate in the bone [160,165].

5. Discussion

Despite the therapeutic advances registered in recent years, the current treatments for bone-related cancers based on the systemic administration of anti-resorptive drugs and/or anti-tumour agents and/or radiopharmaceutical, still experience several drawbacks, including the negative effects elicited by these agents on the normal bone metabolism. These effects may result in detrimental consequences for cancer patients, such as increased risk of osteoporosis and fractures [7,8,9]. Furthermore, none of the currently available clinical treatments have been shown to exert a positive impact on patients' survival [7,8,9]. Hence, the need to develop new molecules endowed with both antitumor activity and low toxicity and/or limited side effects [12-17,24]. In this setting, an increasing number of preclinical studies are currently aimed at identifying and/or developing new molecules based on natural products from plants [12-17]. In particular, a notable number of preclinical investigations have provided convincing evidence on the chemo-preventive effects of several extra virgin olive oil (EVOO)-derived phenolic compounds, including oleuropein, in combatting the growth of several human solid tumors [17-20,22,23,35,36]. Furthermore, experimental and observational studies have shown that OLe exhibits protective effects on bone health due to its property of modulating various molecular signalling pathways involved in the regulation of bone resorption in normal and pathological conditions, including bone-related tumors [7,16,65-68,75,107-109,143]. Therefore, the results emerging from these studies make this molecule an attractive candidate as a novel potential agent in the prevention and treatment of bone-related cancers [17-20,54,45,53,65,66,143]. Interestingly, as OLe appears to be endowed with both anti-tumor [19,20,23,24,25,35,53,142] and bone-anabolic effects [20,65,66,81,82,91-94,98,143], this molecule might be regarded as a potential, additional therapeutic option, in the treatment of specific tumors such as multiple myeloma (MM), which is known to cause severe osteolytic bone loss due to hyper-activation of osteoclasts and suppression of the differentiation of bone marrow mesenchymal stem cells (BMSCs) into functional osteoblasts [91,96,97,167,168] (Table 3). Furthermore these findings

also indicate the administration of OLe may be effective in the prevention of bone loss induced by the systemic administration of antitumor drugs and/or bone modifying agents and/or radiopharmaceuticals [8,9,10,19]. Although, the data from *in vitro* studies are promising in this regard, unfortunately, there is still a lack of *in vivo* studies with the specific aim to evaluate the therapeutic effectiveness of this compound on primary bone tumors or metastatic bone diseases by using suitable experimental animal tumor models. Nevertheless, the accumulating *in vitro* evidence on the capability of OLe to target different molecular pathways involved in the pathogenesis of cancer-related bone diseases, prompts the need for extensive future experimental and clinical investigations.

6. Conclusions

Oleuropein has been reported to target several signaling pathways involved in the modulation of bone remodelling processes in normal and pathological conditions including malignant bone diseases [45,52,53,68,144,161]. These observations suggest the potential usefulness of OLe, administered alone and/or in association with cytotoxic drugs and/or inhibitors of bone metabolism and/or radiopharmaceuticals to prevent the growth and spreading of cancer cells in the bone [19,20,29,37,38,168]. These drug associations might also result in improved therapeutic activity, in a reduction of side effects and in a decreasing probability of developing drug resistance [19,20,24,28,37,76,169,170]. However, there are several drawbacks in transferring these experimental in clinic data. One of the major drawbacks regards the bioavailability and, more in general, the pharmacokinetic properties of OLe [171-174]. The rate of absorption and bioavailability of this molecule appears to be influenced by several factors, such as the route and form of administration, interaction with food, age, sex, different extraction and analytical methods used [171-174]. Experimental and clinical studies have shown that the oral administration of OLe results in rapid absorption, metabolism and renal clearance [117,171-174]. Furthermore, following absorption this compound undergoes extensive phase I and phase II metabolic processes [56,169,173,174]. The potential therapeutic effectiveness of OLe is closely related to the possibility that this molecule may reach in adequate concentrations its specific molecular targets in human tissues [20,143]. Therefore, these phenomena may affect the bioavailability of OLe and its systemic transfer at adequate concentrations to the target tissues, ultimately blunting the therapeutic effectiveness of this compound. However, in this context, data on the effective organ distribution of OLe in human tissues including the bone are currently lacking. Nevertheless, a promising strategy to overcome these hurdles may rely on the development of new semi-synthetic OLe derivatives endowed with better bioavailability and possibly, improved biological activity and spectrum of activity [19,20,160,175-178] and by the use of novel drug delivery systems based on nanotechnology [179-184]. In this latter case, preclinical *in vivo* studies have reported that the nano-encapsulation of OLe could prolong circulation, improve localization, enhance efficacy and reduce the chances of multidrug resistance [117,183-

185]. In conclusion, the increasing number of preclinical *in vitro* studies suggesting a potential chemo-preventive role of OLe in cancer-related bone disease, warrant future studies to assess the impact of this phenolic compound on the clinical management and outcome of patients with primary tumors that reside in, or form metastasis in the bone [13,15,36,43,54,51,67,134,151,156].

Acknowledgements

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflict of Interest statement

The authors declare that there are no conflicts of interest.

References

- [1] R.E. Coleman, Clinical features of metastatic bone disease and risk of skeletal morbidity, *Clin. Cancer Res.* 12(2006) 6243s–6249s, doi: 10.1158/1078-0432.CCR-06-0931.
- [2] J.F. Huang, J. Shen, X. Li, R. Rengan, N. Silvestris, M. Wang, L. Derosa, X. Zheng, A. Belli, X. L. Zhang, Y.M. Li, A. Wu, Incidence of patients with bone metastases at diagnosis of solid tumors in adults: a large population-based study, *Ann. Transl. Med.* 8(7) (2020): 482. doi:10.21037/atm.2020.03.55.
- [3] R.K. Hernandez, S.W. Wade, A. Reich, M. Pirolli, A. Liede, G.H. Lyman, Incidence of bone metastases in patients with solid tumors: analysis of oncology electronic medical records in the United States, *BMC Cancer.* 18(1)(2018) : 44. doi: [10.1186/s12885-017-3922-0](https://doi.org/10.1186/s12885-017-3922-0).
- [4] R.L. Siegel, K.D. Miller, A. Jemal, Cancer statistics, 2019, *CA Cancer J. Clin.* 69(1)(2019):7-34. doi:10.3322/caac.21551.
- [5] SEER Cancer Stat Facts: Bone and Joint Cancer. National Cancer Institute. Bethesda, MD, [HYPERLINK "https://seer.cancer.gov/statfacts/html/bones.html"](https://seer.cancer.gov/statfacts/html/bones.html) . <https://seer.cancer.gov/statfacts/html/bones.html>.
- [6] R. Fonseca, S. Abouzaid, M. Bonafede, Q. Cai, K. Parikh, L. Cosler, P. Richardson, Trends in overall survival and costs of multiple myeloma, 2000-2014, *Leukemia.* 31(9)(2017)1915-1921. doi:10.1038/leu.2016.380.
- [7] S. D'Oronzo, R. Coleman, J. Brown, F. Silvestris, Metastatic bone disease: Pathogenesis and therapeutic options. Up-date on bone metastasis management, *J. Bone Oncol.* 15(2019)4, <https://doi.org/10.1016/j.jbo.2018.10.004>. eCollection 2019 Apr.
- [8] I. Makhoul, C.O. Montgomery, D. Gaddy, L. J. Suva, The best of both worlds - managing the cancer, saving the bone, *Nat. Rev. Endocrinol.* 12 (2016) 29–42, <https://doi.org/10.1038/nrendo.2015.185>.

- [9] M.K. Skjødt, M. Frost, B. Abrahamsen, Side effects of drugs for osteoporosis and metastatic bone disease, *Br. J. Clin. Pharmacol.* 85 (2019) 1063–1071, <https://doi.org/10.1111/bcp.13759>.
- [10] P.G. Turner, J.M. O'Sullivan, ^{223}Ra and other bone-targeting radiopharmaceuticals—the translation of radiation biology into clinical practice, *Br. J. Radiol.* 88 (2015) 20140752, <https://doi.org/10.1259/bjr.20140752>.
- [11] G. Leto, Current status and future directions in the treatment of bone metastases from breast cancer, *Clin. Exp. Pharmacol. Physiol.* 46 (2019) 968–971, <https://doi.org/10.1111/1440-1681.13139>.
- [12] L. Sun, W. Zhou, H. Zhang, Q. Guo, W. Yang, B. Li, Z. Sun, S. Gao, R. Cui, Modulation of Multiple Signaling Pathways of the Plant-Derived Natural Products in Cancer. *Front Oncol* 9(2019). 1153. <https://doi.org/10.3389/fonc.2019.01153>.
- [13] S. Sur, R.B. Ray, Bitter Melon (*Momordica Charantia*), a Nutraceutical Approach for Cancer Prevention and Therapy. *Cancers (Basel)*. 2020;12(8):E2064. Published 2020 Jul 27. doi:10.3390/cancers12082064.
- [14] B. Salehi, P. Zucca, M. Sharifi-Rad, R. Pezzani, S. Rajabi, W.N. Setzer, E.M. Varoni, M. Iriti, F. Kobarfard, J. Sharifi-Rad, Phytotherapeutics in cancer invasion and metastasis, *Phytother. Res.* 32 (2018) 1425–1449, <https://doi.org/10.1002/ptr.6087>.
- [15] P. Juárez, Plant-derived anticancer agents: a promising treatment for bone metastasis, *Bonekey Rep.* 3 (2014) 599, <https://doi.org/10.1038/bonekey.2014.94>.
- [16] A. Kapinova, P. Kubatka, A. Liskova, D. Baranenko, P. Kruzliak, M. Matta, D. Busselberg, B. Malicherova, A. Zulli, T.K. Kwon, E. Jezkova, D. Blahutova, P. Zubor, J. Danko, Controlling metastatic cancer: the role of phytochemicals in cell signaling, *J. Cancer Res. Clin. Oncol.* 145 (2019) 1087–1109, <https://doi.org/10.1007/s00432-019-02892-5.h>.
- [17] A. Maruca, R. Catalano, D. Bagetta, F. Mesiti, F.A. Ambrosio, I. Romeo, F. Moraca, R. Rocca, F. Ortuso, A. Artese, G. Costa, S. Alcaro, A. Lupia, The Mediterranean Diet as source of bioactive compounds with multi-targeting anti-cancer profile., *Eur. J. Med. Chem.* 181 (2019) 111579, doi: 10.1016/j.ejmech.2019.111579. Epub 2019 Jul 3.
- [18] I. Casaburi, F. Puoci, A. Chimento, R. Sirianni, C. Ruggiero, P. Avena, V. Pezzi, Potential of olive oil phenols as chemopreventive and therapeutic agents against cancer: a review of in vitro studies, *Mol. Nutr. Food Res.* 57 (2013) 71–83, <https://doi.org/10.1002/mnfr.201200503>.
- [19] J. Torić, A.K. Marković, C.J. Brala, M. Barbarić, Anticancer effects of olive oil polyphenols and their combinations with anticancer drugs, *Acta Pharm.* 69 (2019) 461–482, <https://doi.org/10.2478/acph-2019-0052>.
- [20] M.L. Castejón, T. Montoya, C. Alarcón-de-la-Lastra, M. Sánchez-Hidalgo, Potential Protective Role Exerted by Secoiridoids from *Olea europaea* L. in Cancer, Cardiovascular, Neurodegenerative, Aging-Related, and Immunoinflammatory Disease, *Antioxidants (Basel)*. 9 (2) (2020) 149, <https://doi.org/10.3390/antiox9020149>.
- [21] I. Hassen, H. Casabianca, K. Hosni, Biological activities of the natural antioxidant oleuropein: Exceeding the expectation-A mini-review, *J. Funct. Foods.* 18 (2015) 926–940, <https://doi.org/10.1016/j.jff.2014.09.001>.

- [22] S. Rigacci, M. Stefani, Nutraceutical Properties of Olive Oil Polyphenols. An Itinerary from Cultured Cells through Animal Models to Humans, *Int. J. Mol. Sci.* 17 (2016), <https://doi.org/10.3390/ijms17060843>.
- [23] H. Shamshoum, F. Vlaveciski, E. Tsiani, Anticancer effects of oleuropein, *Biofactors*. 43 (2017) 517–528, <https://doi.org/10.1002/biof.1366>.
- [24] M. Imran, M. Nadeem, S.A. Gilani, S. Khan, M.W. Sajid, R.M. Amir, Antitumor Perspectives of Oleuropein and Its Metabolite Hydroxytyrosol: Recent Updates, *J. Food Sci.* 83(2018) 1781–1791, <https://doi.org/10.1111/1750-3841.14198>.
- [25] A. Ahmad Farooqi, S. Fayyaz, A.S. Silva, A. Sureda, S.F. Nabavi, A. Mocan, S.M. Nabavi, A. Bishayee, Oleuropein and Cancer Chemoprevention: The Link is Hot, *Molecules*. 22(2017), <https://doi.org/10.3390/molecules22050705>.
- [26] R. Acquaviva, C. Di Giacomo, V. Sorrenti, F. Galvano, R. Santangelo, V. Cardile, S. Gangia, N. D’Orazio, N.G. Abraham, L. Vanella, Antiproliferative effect of oleuropein in prostate cell lines, *Int. J. Oncol.* 41 (2012) 31–38, <https://doi.org/10.3892/ijo.2012.1428>.
- [27] S.A. Mijatovic, G.S. Timotijevic, D.M. Miljkovic, J.M. Radovic, D.D. Maksimovic-Ivanic, D.P. Dekanski, S.D. Stosic-Grujicic, Multiple antimelanoma potential of dry olive leaf extract, *Int. J. Cancer*. 128 (2011) 1955–1965, <https://doi.org/10.1002/ijc.25526>.
- [28] J. Ruzzolini, S. Peppicelli, E. Andreucci, F. Bianchini, A. Scardigli, A. Romani, G. la Marca, C. Nediani, L. Calorini, Oleuropein, the Main Polyphenol of *Olea europaea* Leaf Extract, Has an Anti-Cancer Effect on Human BRAF Melanoma Cells and Potentiates the Cytotoxicity of Current Chemotherapies. *Nutrients*. 10 (2018), <https://doi.org/10.3390/nu10121950>.
- [29] M. Crescimanno, M.V. Sepporta, E. Tripoli, C. Flandina, M. Giammanco, F.M. Tumminello, D. Di Majo, M. Tolomeo, M. La Guardia, G. Leto, Effects of extra virgin olive oil phenols on HL60 cell lines sensitive and resistant to anthracyclines, *J. Biol. Res. - Bollettino della Società Italiana di Biologia Sperimentale*. 82, 1 (Jan. 2009), <https://doi.org/10.4081/jbr.2009.4729>.
- [30] S. Fagan, N. Fulco, J. Gilbertson, A.R. Van Dyke, S.A. Phelan, Anti-cancer effects of oleuropein olive leaf extract in K562 leukemia cells, *Cancer Res.* 78 (13 Suppl.) (2018) 4388, <https://doi.org/10.1158/1538-7445.am2018-4388>.
- [31] Y. Kimura, M. Sumiyoshi, Olive Leaf Extract and Its Main Component Oleuropein Prevent Chronic Ultraviolet B Radiation-Induced Skin Damage and Carcinogenesis in Hairless Mice, *J. Nutr.* 139 (2009) 2079–2086, <https://doi.org/10.3945/jn.109.104992>.
- [32] H.K. Hamdi, R. Castellon, Oleuropein, a non-toxic olive iridoid, is an anti-tumor agent and cytoskeleton disruptor, *Biochem. Biophys. Res. Commun.* 334 (2005) 769–778, <https://doi.org/10.1016/j.bbrc.2005.06.161>.

- [33] R. Liman, F.K. Coban, I.H. Cigerci, I. Bulduk, S. Bozkurt, Antiangiogenic and Apoptotic Effects of Oleuropein on Breast Cancer Cells, *Br. J. Pharm. Res.* 16 (2017) 1-10, <https://doi.org/10.9734/bjpr/2017/33403>.
- [34] M.H. Elamin, A.B. Elmahi, M.H. Daghestani, E.M. Al-Olayan, R.A Al-Ajmi, A.F. Alkhuriji, S. S.Hamed, M. .F Elkhadragy, Synergistic Anti-Breast-Cancer Effects of Combined Treatment With Oleuropein and Doxorubicin In Vivo, *Altern. Ther. Health Med.* 25(3) (2019) 17-24.
- [35] M.V. Sepporta, R. Fuccelli, P. Rosignoli, G. Ricci, M. Servili, G. Morozzi, R. Fabiani, Oleuropein inhibits tumour growth and metastases dissemination in ovariectomised nude mice with MCF-7 human breast tumour xenografts, *J. Funct. Foods.* 8 (2014) 269–273, <https://doi.org/10.1016/j.jff.2014.03.027>.
- [36] H. Song, D.Y. Lim, J.I. Jung, H.J. Cho, S.Y. Park, G.T. Kwon, Y.H. Kang, K.W. Lee, M.S. Choi, and J. Park, Dietary oleuropein inhibits tumor angiogenesis and lymphangiogenesis in the B16F10 melanoma allograft model: a mechanism for the suppression of high-fat diet-induced solid tumor growth and lymph node metastasis, *Oncotarget.* 8(19) (2017) 32027- 32042, doi:10.18632/oncotarget.16757.
- [37] J. Choupani, M.R. Alivand, S. M Derakhshan, M. Zaeifzadeh, M.S. Khaniani, Oleuropein inhibits migration ability through suppression of epithelial-mesenchymal transition and synergistically enhances doxorubicin-mediated apoptosis in MCF-7 cells, *J. Cell Physiol.* 234 (2019) 9093–9104, <https://doi.org/10.1002/jcp.27586>.
- [38] P. Przychodzen, R. Wyszowska, M. Gorzynik-Debicka, T. Kostrzewa, A. Kuban-Jankowska, M. Gorska-Ponikowska, Anticancer Potential of Oleuropein, the Polyphenol of Olive Oil, With 2-Methoxyestradiol, Separately or in Combination, in Human Osteosarcoma Cells, *Anticancer Res.* 39 (2019) 1243–1251, <https://doi.org/10.21873/anticancer.13234>.
- [39] L. Liu, K.S. Ahn, M.K. Shanmugam, H. Wang, H. Shen, F. Arfuso, A. Chinnathambi, S.A. Alharbi, Y. Chang, G. Sethi, F.R. Tang, Oleuropein induces apoptosis via abrogating NF- κ B activation cascade in estrogen receptor-negative breast cancer cells, *J. Cell Biochem.* 120 (2019) 4504–4513, <https://doi.org/10.1002/jcb.27738>.
- [40] H.Y. Lu, J.S. Zhu, Z. Zhang, W.J. Shen, S. Jiang, Y.F. Long, B. Wu, T. Ding, F. Huan, S.L. Wang, Hydroxytyrosol and Oleuropein Inhibit Migration and Invasion of MDA-MB-231 Triple-Negative Breast Cancer Cell via Induction of Autophagy, *Anticancer Agents Med. Chem.* 19 (2019) 1983–1990, <https://doi.org/10.2174/1871520619666190722101207>.
- [41] G. Aktas and H. Ayan, Oleuropein: A Potential Inhibitor for Prostate Cancer Cell Motility by Blocking Voltage-Gated Sodium Channels, *Nutr. Cancer* (2020), doi: 10.1080/01635581.2020.1807575.
- [42] M. Abtin, M.R. Alivand, M.S. Khaniani, M. Bastami, M. Zaeifzadeh, S.M. Derakhshan, Simultaneous downregulation of miR-21 and miR-155 through oleuropein for breast cancer prevention and therapy, *J. Cell Biochem.* 119 (2018) 7151–7165, <https://doi.org/10.1002/jcb.26754>.

- [43] M. Liu, J. Wang, B. Huang, A. Chen, X. Li, Oleuropein inhibits the proliferation and invasion of glioma cells via suppression of the AKT signaling pathway, *Oncol. Rep.* 36 (2016) 2009–2016, <https://doi.org/10.3892/or.2016.4978>.
- [44] M. Seçme, C. Eroğlu, Y. Dodurga, G. Bağcı, Investigation of anticancer mechanism of oleuropein via cell cycle and apoptotic pathways in SH-SY5Y neuroblastoma cells, *Gene.* 585 (2016) 93–99, <https://doi.org/10.1016/j.gene.2016.03.038>.
- [45] E. Scoditti, N. Calabriso, M. Massaro, M. Pellegrino, C. Storelli, G. Martines, R. De Caterina, M.A. Carluccio, Mediterranean diet polyphenols reduce inflammatory angiogenesis through MMP-9 and COX-2 inhibition in human vascular endothelial cells: a potentially protective mechanism in atherosclerotic vascular disease and cancer, *Arch. Biochem. Biophys.* 527 (2012) 81–89, <https://doi.org/10.1016/j.abb.2012.05.003>.
- [46] Z.K. Hassan, M.H. Elamin, M.H. Daghestani, S.A. Omer, E.M. Al-Olayan, M.A. Elobeid, P. Virk, O.B. Mohammed, Oleuropein induces anti-metastatic effects in breast cancer, *Asian Pac. J. Cancer Prev.* 13 (2012) 4555–4559, <https://doi.org/10.7314/apjcp.2012.13.9.4555>.
- [47] Q. Zhou, L.L. Bennett, S. Zhou, Multifaceted ability of naturally occurring polyphenols against metastatic cancer, *Clin. Exp. Pharmacol. Physiol.* 43 (2016) 394–409, <https://doi.org/10.1111/1440-1681.12546>.
- [48] S. Fayyaz, T. Aydin, A. Cakir, M.L. Gasparri, P.B. Panici, A.A. Farooqi, Oleuropein Mediated Targeting of Signaling Network in Cancer, *Curr. Topics Med. Chem.* 16 (2016) 2477–248, <https://doi.org/10.2174/1568026616666160212123706>.
- [49] C.D. Goldsmith, D.R. Bond, H.Jankowski, J. Weidenhofer, C.E. Stathopoulos, P.D. Roach, C.J. Scarlett CJ, The Olive Biophenols Oleuropein and Hydroxytyrosol Selectively Reduce Proliferation, Influence the Cell Cycle, and Induce Apoptosis in Pancreatic Cancer Cells. *Int. J. Mol. Sci.* 19(7) (2018) 1937, doi:10.3390/ijms19071937.
- [50] S. Bayat, S.M. Derakhshan, N.M. Derakhshan, M.S. Khaniani, M.R. Alivand, Downregulation of HDAC2 and HDAC3 via oleuropein as a potent prevention and therapeutic agent in MCF-7 breast cancer cells, *J. Cell. Biochem.* 120 (2019) 9172–9180, <https://doi.org/10.1002/jcb.28193>.
- [51] I. Samet, J. Han, L. Jlaiel, S. Sayadi, H. Isoda, Olive (*Olea europaea*) Leaf Extract Induces Apoptosis and Monocyte/Macrophage Differentiation in Human Chronic Myelogenous Leukemia K562 Cells: Insight into the Underlying Mechanism, *Oxid. Med. Cell. Longev.* (2014) 927619, doi: 10.1155/2014/927619.
- [52] M. Dell'Agli, R. Fagnani, G. V. Galli, O. Maschi, F. Gilardi, S. Bellosta, M. Crestani, E. Bosisio, E. De Fabiani, D. Caruso, Olive Oil Phenols Modulate the Expression of Metalloproteinase 9 in THP-1 Cells by Acting on Nuclear Factor-kappa B Signaling, *J. Agric. Food Chem.* 58 (2010) 2246–2252, <https://doi.org/10.1021/jf9042503>.

- [53] A. Vazquez-Martin, S. Fernandez-Arroyo, S. Cufi, C. Oliveras-Ferraros, J. Lozano-Sanchez, L. Vellon, V. Micol, J. Joven, A. Segura-Carretero, J.A. Menendez, Phenolic Secoiridoids in Extra Virgin Olive Oil Impede Fibrogenic and Oncogenic Epithelial-to-Mesenchymal Transition: Extra Virgin Olive Oil As a Source of Novel Antiaging Phytochemicals, *Rejuvenation Res.* 15 (2012) 3–21, <https://doi.org/10.1089/rej.2011.1203>.
- [54] R.A. Razali, Y. Lokanathan, M.D. Yazid, A.S. Ansari, A. Bin Saim, R.B.H. Idrus, Modulation of Epithelial to Mesenchymal Transition Signaling Pathways by *Olea Europaea* and Its Active Compounds, *Int. J. Mol. Sci.* 20(14) (2019) 3492, <https://doi.org/10.3390/ijms20143492>.
- [55] Y.Q. Ci, J.P. Qiao, M. Han, Molecular Mechanisms and Metabolomics of Natural Polyphenols Interfering with Breast Cancer Metastasis, *Molecules.* 21 (2016) 1634, <https://doi.org/10.3390/molecules21121634>.
- [56] H. Amawi, C.R. Ashby, T. Samuel, R. Peraman, A.K. Tiwari, Polyphenolic Nutrients in Cancer Chemoprevention and Metastasis: Role of the Epithelial-to-Mesenchymal (EMT) Pathway, *Nutrients.* 9 (2017), <https://doi.org/10.3390/nu9080911>.
- [57] F. Xu, Y. Li, M. Zheng, X. Xi, X. Zhang, C. Han, Structure Properties, Acquisition Protocols, and Biological Activities of Oleuropein Aglycone, *Front. Chem.* 6 (2018) 239. doi: 10.3389/fchem.2018.00239. eCollection 2018.
- [58] M.P. Carrera-González, M.J. Ramírez-Expósito, M.D. Mayas, J. M. Martínez-Martos, Protective role of oleuropein and its metabolite hydroxytyrosol on cancer, *Trends Food Sci. Technol.* 31 (2013) 92-99. doi: 10.1016/j.tifs.2013.03.003.
- [59] H.K. Obied, P.D. Prenzler, S.H. Omar, R. Ismael, M. Servili, S. Esposito, A. Taticchi, R. Selvaggini, S. Urbani, Pharmacology of Olive Biophenols, in: J.C. Fishbein, J.M. Heilman (Eds), *Adv. in Mol. Toxicol.* Elsevier: Amsterdam, The Netherlands, 2012; Vol. 6, pp. 195–223.
- [60] M.N. Vissers, P.L. Zock, A.J. Roodenburg, R. Leenen, M.B. Katan, Olive oil phenols are absorbed in humans, *J. Nutr.* 132(3) (2002) 409-417, doi:10.1093/jn/132.3.409.
- [61] R. García-Villalba, M. Larrosa, S. Possemiers, F. A. Tomás-Barberán, J. C. Espín, Bioavailability of phenolics from an oleuropein-rich olive (*Olea europaea*) leaf extract and its acute effect on plasma antioxidant status: comparison between pre- and postmenopausal women, *Eur. J. Nutr.* 53 (2014) 1015–1027, <https://doi.org/10.1007/s00394-013-0604-9>.
- [62] J.I. Mosele, S. Martín-Peláez, A. Macià, M. Farràs, R.-M. Valls, U. Catalán, M.-J. Motilva, Faecal microbial metabolism of olive oil phenolic compounds: In vitro and in vivo approaches, *Mol. Nutr. Food Res.* 58 (2014) 1809–1819. doi: 10.1002/mnfr.20140012.

- [63] J.M. Morana, O. Leal-Hernande, M.L. Canal-Macías, R. Roncero-Martin, R. Guerrero-Bonmatty, I. Aliaga, J.D. Zamorano, Antiproliferative Properties of Oleuropein in Human Osteosarcoma Cells, *Nat. Prod. Commun.* 11 (2016) 491–492, <https://www.ncbi.nlm.nih.gov/pubmed/27396201>.
- [64] Puel, J. Mathey, A. Agalias, S. Kati-Coulibaly, J. Mardon, C. Obled, M.J. Davicco, P. Lebecque, M.N. Horcajada, A.L. Skaltsounis, V. Coxam, Dose-response study of effect of oleuropein, an olive oil polyphenol, in an ovariectomy/inflammation experimental model of bone loss in the rat, *Clin. Nutr.* 25 (2006) 859–868, <https://doi.org/10.1016/j.clnu.2006.03.009>.
- [65] K. Hagiwara, T. Goto, M. Araki, H. Miyazaki, H. Hagiwara, Olive polyphenol hydroxytyrosol prevents bone loss, *Eur. J. Pharmacol.* 662 (2011) 78–84, <https://doi.org/10.1016/j.ejphar.2011.04.023>.
- [66] K.Y. Chin, S. Ima-Nirwana, Olives and Bone: A Green Osteoporosis Prevention Option, *Int. J. Environ. Res. Public Health.* 13 (2016). <https://doi.org/10.3390/ijerph13080755>.
- [67] K.Y. Chin, K.L. Pang, Therapeutic Effects of Olive and Its Derivatives on Osteoarthritis: From Bench to Bedside, *Nutrients.* 9 (2017) <https://doi.org/10.3390/nu9101060>.
- [68] V. Nicolin, N. De Tommasi, S.L. Nori, F. Costantinides, F. Berton, R. Di Lenarda, Modulatory Effects of Plant Polyphenols on Bone Remodeling: A Prospective View From the Bench to Bedside, *Front. Endocrinol. (Lausanne).* 10 (2019) 494, <https://doi.org/10.3389/fendo.2019.00494>.
- [69] E. Impellizzeri, E. Esposito, E. Mazzon, I. Paterniti, R. Di Paola, V.M. Morittu, A. Procopio, D. Britti, S. Cuzzocrea, Oleuropein aglycone, an olive oil compound, ameliorates development of arthritis caused by injection of collagen type II in mice, *J. Pharmacol. Exp. Ther.* 339 (2011) 859–869, <https://doi.org/10.1124/jpet.111.182808>.
- [70] M.N. Horcajada, C. Sanchez, F. Membrez Scalfò, P. Drion, F. Comblain, S. Taralla, A.F. Donneau, E.A. Offord, Y. Henrotin, Oleuropein or rutin consumption decreases the spontaneous development of osteoarthritis in the Hartley guinea pig, *Osteoarthritis Cartilage.* 23 (2015) 94–102, <https://doi.org/10.1016/j.joca.2014.08.016>.
- [71] Z. Feng, X. Li, J. Lin, W. Zheng, Z. Hu, J. Xuan, W. Ni, X. Pan, Oleuropein inhibits the IL-1 β -induced expression of inflammatory mediators by suppressing the activation of NF- κ B and MAPKs in human osteoarthritis chondrocytes, *Food Funct.* 8 (2017) 3737–3744, <https://doi.org/10.1039/c7fo00823f>.
- [72] S. Bulotta, M. Oliverio, D. Russo, A. Procopio, Biological Activity of Oleuropein and its Derivatives. In: *Natural Products*, Ramawat K., Mérillon JM. (eds), Chapter: 156, Publisher: Springer-Verlag Berlin Heidelberg, (2013) 3605-3638, doi 10.1007/978-3-642-22144-6_156.

- [73] A Cabarkapa, L. Zivkovic, S. Borozan, M. Zlatkovic-Svenda, D. Dekanski, I. Jancic, M. Radak-Perovic, V. Bajic, B. Spremo-Potparevic, Dry Olive Leaf Extract in Combination with Methotrexate Reduces Cell Damage in Early Rheumatoid Arthritis Patients. A Pilot Study, *Phytother. Res.* 30 (2016) 1615–1623, <https://doi.org/10.1002/ptr.5662>.
- [74] M. Cruz-Lozano, A. González-González, J.A. Marchal, E. Muñoz-Muela, M.P. Molina, F.E. Cara, A.M. Brown, G. García-Rivas, C. Hernández-Brenes, J.A. Lorente, P. Sanchez-Rovira, J.C. Chang, S. Granados-Principal, Hydroxytyrosol inhibits cancer stem cells and the metastatic capacity of triple-negative breast cancer cell lines by the simultaneous targeting of epithelial-to-mesenchymal transition, Wnt/ β -catenin and TGF β signaling pathways, *Eur. J. Nutr.* 58(8) (2019) 3207-3219, <https://doi.org/10.1007/s00394-018-1864-1>.
- [75] F. Margheri, B. Menicacci, A. Laurenzana, M. Del Rosso, G. Fibbi, M.G. Cipolleschi, J. Ruzzolini, C. Nediani, A. Mocali, L. Giovannelli, Oleuropein aglycone attenuates the pro-angiogenic phenotype of senescent fibroblasts: A functional study in endothelial cells, *J. Funct. Foods.* 53 (2019) 219–226, <https://doi.org/10.1016/j.jff.2018.12.026>.
- [76] A. Boss, K.S. Bishop, G. Marlow, M.P.G. Barnett, L.R. Ferguson, Evidence to Support the Anti-Cancer Effect of Olive Leaf Extract and Future Directions, *Nutrients.* 8(8) (2016) 513, <https://doi.org/10.3390/nu8080513>.
- [77] C. Santangelo, R. Vari, B. Scazzocchio, P. De Sanctis, C. Giovannini, M. D'Archivio, R. Masella, Anti-inflammatory Activity of Extra Virgin Olive Oil Polyphenols: Which Role in the Prevention and Treatment of Immune-Mediated Inflammatory Diseases?, *Endocr. Metab. Immune Disord. Drug Targets.* 18 (2018) 36–50, <https://doi.org/10.2174/1871530317666171114114321>.
- [78] M. Rosillo, C. Alarcón-de-la-Lastra, M.L. Castejón, T. Montoya, M. Cejudo-Guillén, M. Sánchez-Hidalgo, Polyphenolic extract from extra virgin olive oil inhibits the inflammatory response in IL-1 β -activated synovial fibroblasts, *Br. J. Nutr.* 121 (2019) 55–62, <https://doi.org/10.1017/S0007114518002829>.
- [79] I. Peluso, T. Magrone, D. Villaño Valencia, C.O. Chen, M. Palmery, Antioxidant, Anti-Inflammatory, and Microbial-Modulating Activities of Nutraceuticals and Functional Foods, *Oxid. Med. Cell Longev.* (2017) 7658617, <https://doi.org/10.1155/2017/7658617>.
- [80] K. Qabaha, F. Al-Rimawi, A. Qasem, S.A. Naser, Oleuropein Is Responsible for the Major Anti-Inflammatory Effects of Olive Leaf Extract, *J. Med. Food.* 21 (2018) 302–305, <https://doi.org/10.1089/jmf.2017.0070>.
- [81] J.M. Fernández-Real, M. Bulló, J.M. Moreno-Navarrete, W. Ricart, E. Ros, R. Estruch, J. Salas-Salvadó, A Mediterranean diet enriched with olive oil is associated with higher serum total osteocalcin levels in elderly men at high cardiovascular risk, *J. Clin. Endocrinol. Metab.* 97 (2012) 3792–3798, <https://doi.org/10.1210/jc.2012-2221>.

- [82] E. Torre, Molecular signaling mechanisms behind polyphenol-induced bone anabolism, *Phytochem. Rev.* 16 (2017), 1183–1226, <https://doi.org/10.1007/s11101-017-9529-x>.
- [83] X. Feng, J.M. McDonald, Disorders of Bone Remodeling, *Ann. Rev. Pathol.* 6 (2011) 121–145, <https://doi.org/10.1146/annurev-pathol-011110-130203>.
- [84] K.B.S. Paiva, J.M. Granjeiro, Matrix Metalloproteinases in Bone Resorption, Remodeling, and Repair, in: *Matrix Metalloproteinases and Tissue Remodeling in Health and Disease*, R.A. Khalil (Ed.), Target Tissues and Therapy, (2017) 203–303, <https://doi.org/10.1016/bs.pmbts.2017.05.001>.
- [85] M.L. Castejón, M. Rosillo, T. Montoya, A. González-Benjumea, J.G. Fernández-Bolaños, C. Alarcón-de-la-Lastra, Oleuropein down-regulated IL-1 β -induced inflammation and oxidative stress in human synovial fibroblast cell line SW982, *Food Funct.* 8 (2017), 1890–1898. <https://doi.org/10.1039/c7fo00210f>.
- [86] X. Mao, B. Xia, M. Zheng, Z. Zou, Assessment of the anti-inflammatory, analgesic and sedative effects of oleuropein from *Olea europaea* L., *Cell. Mol. Biol. (Noisy-le-grand)* 65(1) (2019)52-55.
- [87] M.M. Taskan, H. Balci Yuce, O. Karatas, F. Gevrek, H. Toker, Evaluation of the effect of oleuropein on alveolar bone loss, inflammation, and apoptosis in experimental periodontitis, *J. Periodontal Res.* 54(6) (2019) 624-632, <https://doi.org/10.1111/jre.12662>.
- [88] B. Menicacci, C. Cipriani, F. Margheri, A. Mocali, L. Giovannelli, Modulation of the Senescence-Associated Inflammatory Phenotype in Human Fibroblasts by Olive Phenols, *Int. J. Mol. Sci.* 18 (2017) <https://doi.org/10.3390/ijms18112275>.
- [89] G. Sindona, A. Caruso, A. Cozza, S. Fiorentini, B. Lorusso, E. Marini, M. Nardi, A. Procopio, S. Zicari, Anti-inflammatory effect of 3,4-DHPEA-EDA [2-(3,4 -hydroxyphenyl) ethyl (3S, 4E)-4-formyl-3-(2-oxoethyl)hex-4-enoate] on primary human vascular endothelial cells, *Curr. Med. Chem.* 19 (2012) 4006–4013, <https://doi.org/10.2174/092986712802002536>.
- [90] S. Lamy, A. Ben Saad, A. Zgheib, B. Annabi, Olive oil compounds inhibit the paracrine regulation of TNF- α -induced endothelial cell migration through reduced glioblastoma cell cyclooxygenase-2 expression, *J. Nutr. Biochem.* 27 (2016) 136–145, <https://doi.org/10.1016/j.jnutbio.2015.08.026>.
- [91] O. García-Martínez, E. De Luna-Bertos, J. Ramos-Torrecillas, C. Ruiz, E. Milia, M.L. Lorenzo, B. Jimenez, A. Sánchez-Ortiz, A. Rivas, Phenolic Compounds in Extra Virgin Olive Oil Stimulate Human Osteoblastic Cell Proliferation, *PLoS One.* 11 (2016) e0150045, <https://doi.org/10.1371/journal.pone.0150045>.
- [92] R. Santiago-Mora, A. Casado-Díaz, M.D. De Castro, J.M. Quesada-Gómez, Oleuropein enhances osteoblastogenesis and inhibits adipogenesis: the effect on differentiation in stem cells derived from bone marrow, *Osteoporosis Int.* 22 (2011) 675–684, <https://doi.org/10.1007/s00198-010-1270-x>.

- [93] R.B.H Idrus, A.B. Saim, *Olea europaea* and Its Constituents Promote Bone Health by Enhancing Osteoblast Differentiation and Proliferation: A Review, *Pharmacognosy J.* 11(1) (2019) 1–7, doi: 10.5530/pj.2019.1.1.
- [94] L. Dudaric, A. Fuzinac-Smojver, D. Muhvic, J. Giacometti, The role of polyphenols on bone metabolism in osteoporosis, *Food Res. Int.* 77 (2015) 290–298, <https://doi.org/10.1016/j.foodres.2015.10.017>.
- [95] R. Filip, S. Possemiers, A. Heyerick, I. Pinheiro, G. Raszewski, M.J. Davicco, V. Coxam, Twelve-month consumption of a polyphenol extract from olive (*Olea europaea*) in a double blind, randomized trial increases serum total osteocalcin levels and improves serum lipid profiles in postmenopausal women with osteopenia, *J. Nutr. Health Aging.* 19 (2015) 77–86, <https://doi.org/10.1007/s12603-014-0480-x>.
- [96] M. Svobodova, I. Andreadou, A.L. Skaltsounis, J. Kopecky, P. Flachs, Oleuropein as an inhibitor of peroxisome proliferator-activated receptor gamma, *Genes Nutr.* 9 (2014) 376, <https://doi.org/10.1007/s12263-013-0376-0>.
- [97] Casado-Díaz, J. Anter, S. Müller, P. Winter, J.M. Quesada-Gómez, G. Dorado, Transcriptomic analyses of the anti-adipogenic effects of oleuropein in human mesenchymal stem cells, *Food Funct.* 8 (2017) 1254–1270, <https://doi.org/10.1039/c7fo00045f>.
- [98] Z.C. Xiong, P. Luo, J. Zhou, M.S. Tan, 15-Deoxy-Delta(12,14)-prostaglandin J(2) as a potential regulator of bone metabolism via PPAR gamma-dependent and independent pathways: a review, *Drug Des. Develop. and Ther.* 13 (2019) 1879–1888, <https://doi.org/10.2147/dddt.s206695>.
- [99] J. Fornetti, A.L. Welm, S.A. Stewart, Understanding the Bone in Cancer Metastasis, *J. Bone Miner. Res.* 33 (2018) 2099–2113, <https://doi.org/10.1002/jbmr.3618>.
- [100] M. Infante, A. Fabi, F. Cognetti, S. Gorini, M. Caprio, A. Fabbri, RANKL/RANK/OPG system beyond bone remodeling: involvement in breast cancer and clinical perspectives, *J. Exp. Clin. Cancer Res.* 38 (2019) 12, <https://doi.org/10.1186/s13046-018-1001-2>.
- [101] H. Roca, L.K. McCauley, Inflammation and skeletal metastasis, *Bonekey Rep.* 4 (2015) <https://doi.org/10.1038/bonekey.2015.75>.
- [102] K.L. Owen, B.S. Parker, Beyond the vicious cycle: The role of innate automimicry and tumor-inherent changes in dictating bone metastasis, *Mol. Immunol.* 110 (2019) 57-68, doi: 10.1016/j.molimm.2017.11.023. Epub 2017 Nov 27.
- [103] B. Ritter, F.R. Greten, Modulating inflammation for cancer therapy, *J. Exp. Med.* 216 (2019) 1234–1243, <https://doi.org/10.1084/jem.20181739>.

- [104] K. Maeda, Y. Kobayashi, M. Koide, S. Uehara, M. Okamoto, A. Ishihara, T. Kayama, M. Saito, K. Marumo, The Regulation of Bone Metabolism and Disorders by Wnt Signaling, *Int. J. Mol. Sci.* 20(22) (2019) 5525, doi:10.3390/ijms20225525.
- [105] L. Zhang, F. Zhou, P. ten Dijke, Signaling interplay between transforming growth factor- β receptor and PI3K/AKT pathways in cancer, *Trends Biochem Sci.* 38 (12) (2013) 612–620, doi:10.1016/j.tibs.2013.10.001.
- [106] J. Zhang, X.H. Yu, Y.G. Yan, C. Wang, W. J. Wang, PI3K/Akt signaling in osteosarcoma, *Clin. Chim. Acta.* 444 (2015)182-92, doi: 10.1016/j.cca.2014.12.041. Epub 2015 Feb 19.
- [107] A.Vallée, Y. Lecarpentier, J.-N. Vallée, Targeting the Canonical WNT/ β -Catenin Pathway in Cancer Treatment Using Non-Steroidal Anti-Inflammatory Drugs, *Cells.* 8(7) (2019) 726, doi:10.3390/cells8070726.
- [108] Y.P. Wu, W.S. Chen, S.J. Xu, N. Zhang, Osteoporosis as a potential contributor to the bone metastases, *Medical Hypotheses.* 75 (2010) 514–516, <https://doi.org/10.1016/j.mehy.2010.07.010>.
- [109] F. Salamanna, V. Borsari, S. Brogini, P. Torricelli, S. Cepollaro, M. Cadossi, M. Fini, A Human 3D In Vitro Model to Assess the Relationship Between Osteoporosis and Dissemination to Bone of Breast Cancer Tumor Cells, *J. Cell. Physiol.* 232 (2017) 1826–1834, <https://doi.org/10.1002/jcp.25708>.
- [110] D. Bellavia, F. Salamanna, L. Raimondi, A. De Luca, V. Carina, V. Costa, R. Alessandro, M. Fini, G. Giavaresi, Deregulated miRNAs in osteoporosis: effects in bone metastasis, *Cell. Mol. Life Sci.* 76 (2019) 3723–3744, <https://doi.org/10.1007/s00018-019-03162-w>.
- [111] L. Das Roy, J.M. Curry, M. Sahraei, D.M. Besmer, A. Kidiyoor, H.E. Gruber, P. Mukherjee, Arthritis augments breast cancer metastasis: role of mast cells and SCF/c-Kit signaling, *Breast Cancer Res.* 15 (2013) R32. <https://doi.org/10.1186/bcr3412>.
- [112] H.M. Chen, F.P. Chen, K.C. Yang, S.S. Yuan, Association of Bone Metastasis With Early-Stage Breast Cancer in Women With and Without Precancer Osteoporosis According to Osteoporosis Therapy Status, *JAMA Netw Open.* 2 (2019) e190429, <https://doi.org/10.1001/jamanetworkopen.2019.0429>.
- [113] A. Muhammad, S.B. Mada, I. Malami, G.E. Forcados, O.L. Erukainure, H. Sani, I.B. Abubakar, Postmenopausal osteoporosis and breast cancer: The biochemical links and beneficial effects of functional foods, *Biomed. Pharmacother.* 107 (2018) 571–582, <https://doi.org/10.1016/j.biopha.2018.08.018>.
- [114] O. Leal-Hernández, R. Roncero-Martín, I. Aliaga, M. Pedrera-Canal, J.M. Morán, S. Rico, L.M. Puerto Parejo and J.M. Lavado-García, P-53-Independent Apoptosis of Human Osteosarcoma Cells After Exposure to Oleuropein , *Lat. Am. J. Pharm.* 39 (2020) 668-672.

- [115] M.H. Elamin, M.H. Daghestani, S.A. Omer, M.A. Elobeid, P. Virk, E.M. Al-Olayan, Z.K. Hassan, O.B. Mohammed, A. Aboussekhra, Olive oil oleuropein has anti-breast cancer properties with higher efficiency on ER-negative cells, *Food Chem. Toxicol.* 53 (2013) 310–316, <https://doi.org/10.1016/j.fct.2012.12.009>.
- [116] A. Papachristodoulou, M. Tsoukala, D. Benaki, S. Kostidis, K. Gioti, N. Aligiannis, H. Pratsinis, D. Kletsas, A.-L. Skaltsounis, E. Mikros, R. Tenta, Oleuropein is a Powerful Sensitizer of Doxorubicin-mediated Killing of Prostate Cancer Cells and Exerts Its Action via Induction of Autophagy, *J. Cancer Res. Treat.* 4 (2018) 61–68, <https://doi.org/10.12691/jcrt-4-4-2>.
- [117] A.M. Nassir, I.A.A. Ibrahim, S. Md, M. Waris, T. MR Ain, I. Ahmad, N. Shahzad, Surface functionalized folate targeted oleuropein nano-liposomes for prostate tumor targeting: Invitro and invivo activity, *Life Sci. Mar* 1;220 (2019) 136-146, doi: 10.1016/j.lfs.2019.01.053.
- [118] W. Wang, J. Wu, Q. Zhang, X. Li, X. Zhu, Q. Wang, S. Cao, L. Du, Mitochondria-mediated apoptosis was induced by oleuropein in H1299 cells involving activation of p38 MAP kinase, *J. Cell Biochem.* 120 (2019) 5480–5494, <https://doi.org/10.1002/jcb.27827>.
- [119] C. Antognelli, R. Frosini, M.F. Santolla, M.J Peirce, V.N. Talesa, Oleuropein-Induced Apoptosis Is Mediated by Mitochondrial Glyoxalase 2 in NSCLC A549 Cells: A Mechanistic Inside and a Possible Novel Nonenzymatic Role for an Ancient Enzyme, *Oxidative Med. Cell. Longev.* 2019 (2019) 8576961, <https://doi.org/10.1155/2019/8576961>.
- [120] W. Mao, H. Shi, X. Chen, Y. Yin, T. Yang, M. Ge, M. Luo, D. Chen and X. QiAN, Anti-Proliferation and Migration Effects of Oleuropein on Human A549 Lung Carcinoma Cells, *Lat. Am. J. Pharm.* 31 (8) (2012) 1217-21.
- [121] Z. Bouallagui, J Han, Isoda, S. Sayadi, Hydroxytyrosol rich extract from olive leaves modulates cell cycle progression in MCF-7 human breast cancer cells, *Food Chem. Toxicol.* 49(2011)179–184, doi:10.1016/j.fct.2010.10.014.
- [122] J. Han, T.P.N. Talorete, P. Yamada, H. Isoda, Anti-proliferative and apoptotic effects of oleuropein and hydroxytyrosol on human breast cancer MCF-7 cells, *Cytotechnology.* 59 (2009) 45–53, doi: 10.1007/s10616-009-9191-2.
- [123] A. Chimento, I. Casaburi, C. Rosano, P. Avena, A. De Luca, C. Campana, E. Martire, M.F. Santolla, M. Maggiolini, V. Pezzi, Oleuropein and hydroxytyrosol activate GPER/GPR30-dependent pathways leading to apoptosis of ER-negative SKBR3 breast cancer cells, *Mol. Nutr. Food Res.* 58 (2014) 478–489, <https://doi.org/10.1002/mnfr.201300323>.

- [124] R. Sirianni, A. Chimento, A. De Luca, I. Casaburi, P. Rizza, A. Onofrio, D. Iacopetta, F. Puoci, S. Andò, M. Maggiolini and V. Pezzi, Oleuropein and hydroxytyrosol inhibit MCF-7 breast cancer cell proliferation interfering with ERK1/2 activation, *Mol. Nutr. Food Res.* 54 (2010) 833–840, DOI: 10.1002/mnfr.200900111.
- [125] S. Cao, X. Zhu, L. Du, P38 MAP kinase is involved in oleuropein-induced apoptosis in A549 cells by a mitochondrial apoptotic cascade, *Biomed. Pharmacother.* 95 (2017) 1425-1435, doi: 10.1016/j.biopha.2017.09.072.
- [126] Z.K. Hassan, M.H. Elamin, S.A. Omer, M.H. Daghestani, E.S. Al-Olayan, M.A. Elobeid, P. Virk, Oleuropein induces apoptosis via the p53 pathway in breast cancer cells. *Asian Pac. J. Cancer Prev.* 14(11) (2014) 6739-6742, doi: 10.7314/apjcp.2013.14.11.6739. PMID: 24377598.
- [127] S. Asgharzade, S. H. Sheikhshabani, E. Ghasempour, R. Heidari, S. Rahmati, M. Mohammadi, A. Jazaeri, Z. Amini-Farsani, The effect of oleuropein on apoptotic pathway regulators in breast cancer cells, *Eur J Pharmacol.* 2020;886:173509. doi:10.1016/j.ejphar.2020.173509.
- [128] B. Kaltschmidt, C. Banz-Jansen, T. Benhidjeb T, M. Beshay, C. Förster, J. Greiner, E. Hamelmann, N. Jorch, F. Mertzlufft, J. Pfitzenmaier, M. Simon, J. Schulte Am Esch, T. Vordemvenne, D. Wähnert, F. Weissinger, L. Wilkens, B. Kaltschmidt, A Role for NF- κ B in Organ Specific Cancer and Cancer Stem Cells. *Cancers (Basel)*. 11(5) (2019) 655, doi:10.3390/cancers11050655.
- [129] C.J. Li, P.Y. Chu, G.T. Yiang, M.Y. Wu, The Molecular Mechanism of Epithelial-Mesenchymal Transition for Breast Carcinogenesis, *Biomolecules*. 9(9) (2019) 476, doi:10.3390/biom9090476.
- [130] B. Demirkan, The Roles of Epithelial-to-Mesenchymal Transition (EMT) and Mesenchymal-to-Epithelial Transition (MET) in Breast Cancer Bone Metastasis: Potential Targets for Prevention and Treatment, *J. Clin. Med.* 2(4) (2013) 264-282, doi: 10.3390/jcm2040264.
- [131] A.H. Nwabo Kamdje, P. Takam Kamga, R. Tagne Simo, L. Vecchio, P.F. Seke Etet, J.M. Muller, G. Bassi, E. Lukong, R. Kumar Goel, J. Mbo Amvene, M. Krampera, Developmental pathways associated with cancer metastasis: Notch, Wnt, and Hedgehog, *Cancer Biol. Med.* 14 (2017) 109-120, doi: 10.20892/j.issn.2095-3941.2016.0032.
- [132] P. Juárez, T.A. Guise, TGF- β in cancer and bone: implications for treatment of bone metastases. *Bone*. 48(1) (2011) 23–29, doi:10.1016/j.bone.2010.08.004.
- [133] Y. Hao, D. Baker, P. Ten Dijke, TGF- β -Mediated Epithelial-Mesenchymal Transition and Cancer Metastasis, *Int. J. Mol. Sci.* 20(11)(2019) 2767, doi:10.3390/ijms20112767.

- [134] F. Verrecchia, F. R dini, Transforming Growth Factor- β Signaling Plays a Pivotal Role in the Interplay Between Osteosarcoma Cells and Their Microenvironment. *Front Oncol.* (8) (2018) 133, doi:10.3389/fonc.2018.0013.
- [135] H. Tsai and J. Yang, Epithelial-mesenchymal plasticity in carcinoma metastasis, *Genes Dev.* 27(20) (2013) 2192–2206, doi: 10.1101/gad.225334.113.
- [136] C. Chandhanayingyong, Y. Kim, J.R. Staples, C. Hahn, F.Y. Lee, MAPK/ERK Signaling in Osteosarcomas, Ewing Sarcomas and Chondrosarcomas: Therapeutic Implications and Future Directions. *Sarcoma.* 2012, 404810. doi:10.1155/2012/404810.
- [137] S.R. Lin, N. Mokgautsi, Y.N. Liu, Ras and Wnt Interaction Contribute in Prostate Cancer Bone Metastasis, *Molecules.* 25(10)(2020) 2380, doi: 10.3390/molecules25102380.
- [138] G. Danieau, S. Morice, F. R dini, F. Verrecchia, B.B. Royer, New Insights about the Wnt/ β -Catenin Signaling Pathway in Primary Bone Tumors and Their Microenvironment: A Promising Target to Develop Therapeutic Strategies? *Int. J. Mol. Sci.* 20(15) (2019)3751, doi:10.3390/ijms20153751.
- [139] B.K. Park, H. Zhang, Q. Zeng, J. Dai, E.T. Keller, T. Giordano, K. Gu, V. Shah, L. Pei, R.J. Zarbo RJ, L. McCauley, S. Shi, S. Chen, C.Y. Wang, NF-kappa B in breast cancer cells promotes osteolytic bone metastasis by inducing osteoclastogenesis via GM-CSF, *Nat. Med.* 13 (2007) 62–69, <https://doi.org/10.1038/nm1519>.
- [140] R. Jin, J.A. Sterling, J.R. Edwards, D.J. eGraff, C. Lee, S.I. Park, R.J. Matusik, Activation of NF-kappa B signaling promotes growth of prostate cancer cells in bone, *PLoS One.* 8(4) (2013) e60983, <https://doi.org/10.1371/journal.pone.0060983>.
- [141] R.W. Johnson, M.E. Sowder, A.J. Giaccia, Hypoxia and Bone Metastatic Disease, *Curr. Osteoporos. Rep.* 15(4) (2017) 231–238, doi:10.1007/s11914-017-0378-8.
- [142] F.N. Soki, S.I. Park, L.K. McCauley, The multifaceted actions of PTHrP in skeletal metastasis, *Future Oncol.* 8(7) (2012) 803–817, doi:10.2217/fon.12.76.
- [143] C. Nediani, J. Ruzzolini, A. Romani, L. Calorini, Oleuropein, a Bioactive Compound from *Olea europaea* L, as a Potential Preventive and Therapeutic Agent in Non-Communicable Diseases, *Antioxidants (Basel).* 8(12) (2019) 578, doi:10.3390/antiox8120578.
- [144] V. Goulas V. Exarchou, A.N. Troganis, E. Psomiadou, T. Fotsis, E. Briasoulis and I.P. Gerothanassis, Phytochemicals in olive-leaf extracts and their antiproliferative activity against cancer and endothelial cells, *Mol. Nutr. Food Res.* 53(5) (2009) 600-608, doi:10.1002/mnfr.200800204.

- [145] X. Lu, E. Mu, Y. Wei, S. Riethdorf, Q. Yang, M. Yuan, J. Yan, Y. Hua, B.J. Tiede, X. Lu, B.G. Haffty, K. Pantel, J. Massagué, Y. Kang, VCAM-1 promotes osteolytic expansion of indolent bone micrometastasis of breast cancer by engaging $\alpha 4\beta 1$ -positive osteoclast progenitors, *Cancer Cell*. 20(6) (2011) 701–714, doi:10.1016/j.ccr.2011.11.002.
- [146] M. Dell’Agli, R. Fagnani, N. Mitro, S. Scurati, M. Masciadri, L. Mussoni, G.V. Galli, E. Bosisio, M. Crestani, E. De Fabiani, E. Tremoli, D. Caruso, Minor Components of Olive Oil Modulate Proatherogenic Adhesion Molecules Involved in Endothelial Activation, *J. Agric. Food Chem.* 54 (2006) 3259–3264. doi: 10.1021/jf0529161.
- [147] H. Choi, A. Moon, Crosstalk between cancer cells and endothelial cells: implications for tumor progression and intervention, *Arch. Pharm. Res.* 41 (2018) 711–724, <https://doi.org/10.1007/s12272-018-1051-1>.
- [148] N. Yahfoufi, N. Alsadi, M. Jambi, C. Matar, The Immunomodulatory and Anti-Inflammatory Role of Polyphenols, *Nutrients*. 10 (11) (2018) 1618, <https://doi.org/10.3390/nu10111618>.
- [149] M. Paolillo, S. Schinelli, Extracellular Matrix Alterations in Metastatic Processes, *Int. J. Mol. Sci.* 20(19) (2019) 4947, doi:10.3390/ijms20194947.
- [150] H. Harjunpää, M. Lloret Asens, C. Guenther, S.C. Fagerholm, Cell Adhesion Molecules and Their Roles and Regulation in the Immune and Tumor Microenvironment, *Front. Immunol.* 10 (2019) 1078, doi:10.3389/fimmu.2019.01078.
- [151] N. Hashemi Goradel, M. Najafi, E. Salehi, B. Farhood, K. Mortezaee, Cyclooxygenase- 2 in cancer: A review, *J. Cell. Physiol.* 234 (2019) 5683– 5699, <https://doi.org/10.1002/jcp.27411>.
- [152] E. Hardy, C. Fernandez-Patron, Destroy to Rebuild: The Connection Between Bone Tissue Remodeling and Matrix Metalloproteinases, *Front. Physiol.* 2020;11:47, doi: 10.3389/fphys.2020.00047. eCollection 2020.
- [153] A. Dhawan, J. Friedrichs, M.V. Bonin, E.P. Bejestani, C. Werner, M. Wobus, T. Chavakis, M. Bornhäuser, Breast cancer cells compete with hematopoietic stem and progenitor cells for intercellular adhesion molecule 1-mediated binding to the bone marrow microenvironment, *Carcinogenesis*. 37(8) (2016) 759-767, doi: 10.1093/carcin/bgw057. Epub 2016 May 4.
- [154] M. Esposito, N. Mondal, T.M. Greco, Y. We, C. Spadazzi, S.C. Lin, H. Zheng, C. Cheung, J.L. Magnani, S.H. Lin, I.M. Cristea, R. Sackstein, Y. Kang, Bone vascular niche E-selectin induces mesenchymal-epithelial transition and Wnt activation in cancer cells to promote bone metastasis, *Nat. Cell Biol.* 21(5) (2019) 627–639, doi:10.1038/s41556-019-0309-2.

- [155] P. Przychodzen, A. Kuban-Jankowska, R. Wyszowska, G. Barone, G. Lo Bosco, F. Lo Celso, A. Kamm, A. Daca, T. Kostrzewa, M. Gorska-Ponikowska, PTP1B phosphatase as a novel target of oleuropein activity in MCF-7 breast cancer model, *Toxicol. In Vitro*. 61(2019) 104624, doi:10.1016/j.tiv.2019.104624.
- [156] B. Hilmarsson, E. Briem, S. Halldorsson, J. Krickler, S. Ingthorsson, S. Gustafsdottir, G.M. Mælandsmo, M.K. Magnusson, T. Gudjonsson, Inhibition of PTP1B disrupts cell–cell adhesion and induces anoikis in breast epithelial cells, *Cell Death Dis.* 8(5) (2017) e2769, <https://doi.org/10.1038/cddis.2017.17>.
- [157] G. Ren, M. Esposito, Y. Kang, Bone metastasis and the metastatic niche, *J. Mol. Med (Berl)*. 93(11) (2015) 1203–1212, doi:10.1007/s00109-015-1329-4.
- [158] Y. Liu, X. Cao, Characteristics and Significance of the Pre-metastatic Niche, *Cancer Cell*. 30(5) (2016) 668–681. doi: 10.1016/j.ccell.2016.09.011.
- [159] N.M. Byrne, M.A. Summers, M.M. McDonald, Tumor Cell Dormancy and Reactivation in Bone: Skeletal Biology and Therapeutic Opportunities, *JBMR Plus*. 3(3) (2019) e10125, doi:10.1002/jbm4.10125.
- [160] M.A. Rosillo, S. Montserrat-de-la-Paz, R. Abia, M. L. Castejon, M. C. Millan-Linares, C. Alarcon-de-la-Lastra, J.G. Fernandez-Bolaños, and F. Muriana, Oleuropein and its peracetylated derivative negatively regulate osteoclastogenesis by controlling the expression of genes involved in osteoclast differentiation and function, *Food Funct*. 11(5)(2020) 4038- 4048, doi:10.1039/d0fo00433b.
- [161] S. Kany, J.T. Vollrath, B. Relja, Cytokines in Inflammatory Disease, *Int. J. Mol. Sci.* 20(23) (2019) 6008, doi:10.3390/ijms20236008.
- [162] F. Salamanna, V. Borsari, D. Contartese, V. Costa, G. Giavaresi, M. Fini, What Is the Role of Interleukins in Breast Cancer Bone Metastases? A Systematic Review of Preclinical and Clinical Evidence. *Cancers(Basel)*. 11(12) (2018) 2019, doi:10.3390/cancers11122018.
- [163] A.C. Monteiro, A.C. Leal, T. Gonçalves-Silva, A.T.C. Mercadante, F. Kestelman, S.B. Chaves, R. J. P. Monteiro, A. Bonomo, T cells induce pre-metastatic osteolytic disease and help bone metastases establishment in a mouse model of metastatic breast cancer, *PLoS One*. 8(7) (2013) e68171, doi:10.1371/journal.pone.0068171.
- [164] H. Khan, H. Ullah, P.C.M.F. Castilho, A.S. Gomila, G. D’Onofrio, R. Filosa, F. Wang, S.M. Nabavi, M. Daglia, A.S. Silva, K.R.R. Rengasamy, J. Ou, X. Zou, J. Xiao, H. Cao, Targeting NF- κ B signaling pathway in cancer by dietary polyphenols, *Crit. Rev. Food Sci. Nutr.* (2019) 1–11. <https://doi.org/10.1080/10408398.2019.1661827>.
- [165] D. Verzella, M. Fischietti, D. Capece, D. Vecchiotti, F. Del Vecchio, G. Cicciarelli, V. Mastroiaco, A. Tessitore, E. Alesse, F. Zazzeroni, Targeting the NF- κ B pathway in prostate cancer: a promising therapeutic approach? *Curr. Drug Targets*. 17(3) (2016) 311- 320, doi:10.2174/1389450116666150907100715.

- [166] X. Wu, F. Li, L. Dang, C. Liang, A. Lu, G. Zhang, RANKL/RANK System-Based Mechanism for Breast Cancer Bone Metastasis and Related Therapeutic Strategies, *Front. Cell Dev. Biol.* 8 (2020) 76, doi: 10.3389/fcell.2020.00076. eCollection 2020.
- [167] A. Suvannasankha, J.M. Chirgwin, Role of bone-anabolic agents in the treatment of breast cancer bone metastases, *Breast Cancer Res* 16 (2014) 484, <https://doi.org/10.1186/s13058-014-0484-9>.
- [168] J. Adamik, D.L. Galson, G.D. Roodman, Osteoblast suppression in multiple myeloma bone disease, *J Bone Oncol.* 13 (2018) 62-70, doi:10.1016/j.jbo.2018.09.001.
- [169] Y. Xing, D. Cui, S. Wang, P. Wang, X. Xing, H. Li, Oleuropein represses the radiation resistance of ovarian cancer by inhibiting hypoxia and microRNA-299-targetted heparanase expression, *Food Funct.* 8(8) (2017) 2857–2864, doi: 10.1039/c7fo00552k.
- [170] J.A. Menendez, A. Vazquez-Martin, R. Colomer, J. Brunet, A. Carrasco-Pancorbo, R. Garcia-Villalba, A. Fernandez-Gutierrez and A. Segura-Carretero, Olive oil's bitter principle reverses acquired autoresistance to trastuzumab (Herceptin™) in HER2-overexpressing breast cancer cells, *BMC Cancer* 7 (2007) 80, <https://doi.org/10.1186/1471-2407-7-80>.
- [171] M. de Bock, E.B. Thorstensen, J.G. Derraik, H.V. Henderson, P.L. Hofman, W.S. Cutfield, Human absorption and metabolism of oleuropein and hydroxytyrosol ingested as olive (*Olea europaea* L.) leaf extract, *Mol. Nutr. Food Res.* 57(11) (2013) 2079- 2085, doi:10.1002/mnfr.201200795.
- [172] L. Rubi , A. Maci , M. J. Motilva, Impact of various factors on pharmacokinetics of bioactive polyphenols: an overview, *Curr Drug Metab.* 15 (2014) 62-76. doi:10.2174/1389200214666131210144115.
- [173] M.C.L. De las Hazas, C. Pi ol, A. Maci , M.-P. Romero, A. Pedret, R. Sol , L. Rubi , M.-J. Motilva, Differential absorption and metabolism of hydroxytyrosol and its precursors oleuropein and secoiridoids, *J. Funct. Food.* 22 (2016) 52–63, <https://doi.org/10.1016/j.jff.2016.01.030>.
- [174] F. Visioli, A. Davalos, M- C. L pez de las Hazas, M.C. Crespo, J. Tom - Carneiro, An overview of the pharmacology of olive oil and its active ingredients, *Br J. Pharmacol.* 177 (2020) 1316–1330. <https://doi.org/10.1111/bph.14782>.
- [175] D. Russo, D. Britti, S.M. Lepore, F. Trimboli, M. Celano, M. Oliverio, R.F. Rose, G. Loprete, N. Costa, C.D. Loreto, G. Damante, S. Bonacci, A. Procopio, Safety Evaluation of Subchronic Oral Administration of Oleuropein and its Semisynthetic Peracetylated Derivative, *J. Pharm. Pharmaceutics* 3(1) (2016) 40- 45, <https://doi.org/10.15436/2377-1313.16.014>.

- [176] A. Karković Marković, J. Torić, M. Barbarić, C. Jakobušić Brala, Hydroxytyrosol, Tyrosol and Derivatives and Their Potential Effects on Human Health, *Molecules*. 2019;24(10):2001. doi:10.3390/molecules24102001.
- [177] S. Bulotta, R. Corradino, M. Celano, J. Maiuolo, M. D'Agostino, M. Oliverio, A. Procopio, S. Filetti and D. Russo, Antioxidant and antigrowth action of peracetylated oleuropein in thyroid cancer cells, *J. Mol. Endocrinol.* 51(1) (2013)181-189. doi: 10.1530/JME-12-0241
- [178] G. Juli, M. Oliverio, D. Bellizzi, M.E. Gallo Cantafio, K. Grillone, G. Passarino, C. Colica, M. Nardi, M. Rossi, A. Procopio, P. Tagliaferri, P. Tassone, N. Amodio N, Anti-tumor Activity and Epigenetic Impact of the Polyphenol Oleacein in Multiple Myeloma, *Cancers (Basel)*. 11(7) (2019) 990, doi: 10.3390/cancers11070990.
- [179] G. Chindamo, S. Sapino, E. Peira, D. Chirio, M.C. Gonzalez, M.Gallarate, Bone Diseases: Current Approach and Future Perspectives in Drug Delivery Systems for Bone Targeted Therapeutics, *Nanomaterials (Basel)*. 10(5) (2020) 875, doi:10.3390/nano10050875.
- [180] F. Yang, Z. Zhao, B. Sun, Q. Chen, J. Sun, Z. He, C. Luo, Nanotherapeutics for Antimetastatic Treatment *Trends Cancer*. 6(8) (2020) 645-659, doi: <https://doi.org/10.1016/j.trecan.2020.05.001>.
- [181] I.M. Adjei, M.N. Temples, S.B. Brown, B. Sharma, Targeted Nanomedicine to Treat Bone Metastasis. *Pharmaceutics*. 10(4) (2018) 205, doi:10.3390/pharmaceutics10040205.
- [182] W. Gu, C. Wu, J. Chen, Y. Xiao, Nanotechnology in the targeted drug delivery for bone diseases and bone regeneration, *Int. J. Nanomedicine*. 8 (2013) 2305-2317, doi:10.2147/IJN.S44393.
- [183] S. Palagati, S. Sv and B.R. Kesavan, Application of computational tools for the designing of Oleuropein loaded nanostructured lipid carrier for brain targeting through nasal route, *Daru J. Pharm. Sci.* 27 (2019) 695 -708, doi:10.1007/s40199-019-00304-0.
- [184] C. Bonechi, A. Donati, G. Tamasi, A. Pardini, H. Rostom, G. Leone, S. Lamponi, M. Consumi, A. Magnani and C. Ross, Chemical characterization of liposomes containing nutraceutical compounds: Tyrosol, hydroxytyrosol and oleuropein, *Biophys. Chem.* 246 (2019) 25-34, <https://doi.org/10.1016/j.bpc.2019.01.002>.
- [185] S. Tabrez, N.R. Jabir, V.M. Adhami, M.I. Khan, M. Moulay, M.A. Kamal, H. Mukhtar, Nanoencapsulated dietary polyphenols for cancer prevention and treatment: successes and challenges, *Nanomedicine* 15, 1(2020)147–1162, doi: 10.2217/nmm-2019-0398.

Figure 1. Chemical structure of Oleuropein and its hydrolysis products

Figure 2. Pleiotropic effects of Oleuropein on cellular and molecular components involved in pre-metastatic niche formation.

OLe may inhibit (⚡) the expression and activity of growth factors, hormones, proinflammatory cytokines, adhesion molecules and enzymes, which regulate different cellular processes involved in cancer progression, including *i*) apoptosis, *ii*) cell cycle progression, *iii*) cell differentiation and *iv*) epithelial-to-mesenchymal transition (EMT). Most of these signalling molecules have been shown to be deregulated in several pathological conditions associated with bone loss and to contribute to the formation of the “*bone pre-metastatic niche*”. On the other hand, OLe has been shown to exert osteo-protective effects by fostering (⇒) the differentiation of human bone marrow mesenchymal stem cells into osteoblasts and by inhibiting (⚡) osteoclastogenesis.

Figure 3 Schematic representation of the effects of Oleuropein on molecular signalling pathways linking chronic inflammation and cancer progression

OLe may modulate the expression and activity of various signalling molecules, such as **1**) cytokines, chemokines [20,21,55,69,71,73,75], **2**) transcription factors [18,20,38,71,75-80], and **3**) enzymes [20,43,44,71,75,77] which, in physiological conditions, regulate several biological functions including the normal metabolic turnover of bone tissue while, their expression proves to be deregulated in various pathological conditions associated with altered bone remodelling such as chronic inflammatory processes and malignant bone disorders [7,20,47,68,71,75,83, 99-103,106].

Word count 10967 ; Figures 3; Tables 3.;

Effects of Oleuropein on tumor cell growth and bone remodeling: Potential clinical implications for the prevention and treatment of malignant bone diseases

Gaetano Leto^{a*}, Carla Flandina^a, Marilena Crescimanno^a, Marco Giammanco^b and Maria Vittoria Sepporta^c

^aLaboratory of Experimental Pharmacology, Department of Health Sciences, University of Palermo, 90127 Palermo, Italy.

^bDepartment of Surgical, Oncological and Oral Sciences University of Palermo, 90127 Palermo, Italy.

^cPediatric Unit, Department Women-Mother-Children, Pediatric Hematology-Oncology Research Laboratory, **Lausanne University Hospital**, Lausanne, Switzerland.

*Corresponding author: Gaetano Leto, Laboratory of Experimental Pharmacology, Department of Health Sciences, University of Palermo, 90127 Palermo, Italy. e-mail, gaetano.let@unipa.it; Orcid:0000-0002-0900-1259

Abstract

Oleuropein (Ole) is the main bioactive phenolic compound present in olive leaves, fruits and olive oil. This molecule has been shown to exert beneficial effects on several human pathological conditions. In particular, recent preclinical and observational studies have provided evidence that Ole exhibits chemo-preventive effects on different types of human tumors. Studies undertaken to elucidate the specific mechanisms underlying these effects have shown that this molecule may thwart several key steps of malignant progression, including tumor cell proliferation, survival, angiogenesis, invasion and metastasis, by modulating the expression and activity of several growth factors, cytokines, adhesion molecules and enzymes involved in these processes. Interestingly, experimental observations have highlighted the fact that most of these signalling molecules also appear to be actively involved in the homing and growth of disseminating cancer cells in bones and, ultimately, in the development of metastatic bone diseases. These findings, and the experimental and clinical data reporting the preventive activity of Ole on various pathological conditions associated with a bone loss, are indicative of a potential therapeutic role of this molecule in the prevention and treatment of cancer-related bone diseases. This paper provides a current overview regarding the molecular mechanisms and the experimental findings underpinning a possible clinical role of Ole in the prevention and development of cancer-related bone diseases.

Keywords: Bone; Cancer; Chemoprevention; Metastasis; Oleuropein; Polyphenols; Tumor progression

Chemical compounds: Elenolic Acid (PubChem CID: 169607); Hydroxytyrosol (PubChem CID: 82755); Oleuropein (PubChem CID: 5281544); Oleuropein aglycone (Pubchem CID: 56842347); Tyrosol (PubChem CID: 10393)

Abbreviations

BCa, Breast Cancer

BMSCs, Bone Marrow Mesenchymal Stem Cells

BMP, Bone Morphogenic Protein

BPH, Benign prostate hyperplasia epithelial cells

COX-2, Cyclooxygenase-2

CSC, Cancer Stem Cells

CXCRs, CXC Chemokine receptors

Elac, Elenolic Acid

ELAM-1 endothelial-leukocyte adhesion molecule 1

ECM, Extracellular matrix

EMT, Epithelial Mesenchymal Transition

EVOO, Extra Virgin Olive Oil

γ -GCS, γ -Glutamylcysteine Synthetase

Glo2, Mitochondrial Glyoxalase 2

HDAC2, Histone deacetylase 2

HDAC3, Histone deacetylase 3

HIF, Hypoxia Inducible Factor

HO-1, Heme Oxygenase-1

Hyt, Hydroxytyrosol

ICAM-1, Intercellular Adhesion Molecule-1

i-NOS, Inducible Form Of Nitric Oxide Synthase

IL-1 β , Interleukin-1 β

IL-6, Interleukin-6

IL-8, Interleukin-8

MAPK, Mitogen Activated Protein-Kinase

MCP-1, Monocyte Chemo-Attractant Protein-1

MMPS, Matrix Metalloproteinases

MMP-2, Matrix Metalloproteinases

MMP-9, Matrix Metalloproteinases

MM, Multiple Myeloma

mTOR, Mammalian Target Of Rapamycin

NF-kB, Nuclear Factor kB

NSLC, Non Small Cell Lung Cancer

OLE, Oleuropein.

OLE-A, Oleuropein Aglycone

OB, Osteoblast

OCL, Osteoclast

OPG/RANKL, Osteoprotegerin/Receptor Activator Of Nuclear Factor Kappa-B Ligand Expression Ratio

OS, Osteosarcoma

PARP, Poly (ADP-Ribose) Polymerase

PCa, Prostate Cancer
 PTPB1, Protein Tyrosine Phosphatase B1
 PTHrP, Parathyroid hormone-related protein
 PGs, Prostaglandins
 PI3K/AKT, Phosphatidylinositol-3 Kinase/ Protein Kinase B
 PPAR γ , Peroxisomal Proliferator–Activated Receptor
 RANK-L, Receptor Activator of NF- κ B Ligand
 RNI, Reactive Nitrogen Intermediates
 ROS, Reactive Oxygen Species
 RUNX2, Runt-Related Transcription Factor 2
 SOD2, Superoxide Dismutase 2
 STAT3, Signal Transducer and Activators of Transcription 3
 TIMP, Tissue Inhibitor of Metallproteinase
 TNF- α , Tumor Necrosis Factor- α
 TGF- β , Transforming Growth Factor β ,
 TRAP, Tartrate-Resistant Acid Phosphatase
 VEGF, Vascular Endothelial Growth Factor;
 VEGFR, VEGF Receptor
 VGSCs blocking voltage-gated sodium channels
 V-CAM-1, Vascular Cellular Adhesion Molecule-1
 Wnt/ β -Catenin, Wingless-Related Integration Site/ B-Catenin

1. Introduction

The skeleton is the third most common site of metastatic disease after lung and liver [1]. Many types solid cancers, in particular, breast, prostate, thyroid, lung, and kidney tumors preferentially metastasize to bone [1,2]. The incidence of bone metastasis by tumor type, reaches 65–90% in prostate cancer (PCa), about 65–75% in breast cancer (BCa), 60% in thyroid cancer, 30-40% in lung cancer, 40% in bladder, cancer, 20-25% in renal cell carcinoma. Skeletal involvement is less frequent in other malignancies such as melanoma (11-17%) and colorectal tumors (10%) [2,3]. The median-survival from diagnosis of bone metastasis from breast cancer, prostate cancer, thyroid cancer are 27 months, 25 months, and 23 months, respectively while, in patients with renal, bladder, lung, and melanoma median survival is lower (12 months, 8 months, 7-9 months and 6 months respectively) [2-4]. Furthermore, in some haematological malignancies such as in multiple myeloma (MM) the rate of incidence reaches 70–95% [4,5]. Interestingly, MM patients diagnosed after 2010 experienced improved clinical outcomes, the 2-year survival rate having increased from 69.9% in 2006 to 87.1% in 2012 [5,6]. On the other hand, primary malignant bone tumors are relatively rare, occurring at a rate of about one to 100,000 [4,5]. In adults, over 40% of primary bone cancers are chondrosarcomas. This is followed by osteosarcomas (28%), chordomas (10%), Ewing tumors (8%), and malignant fibrous histiocytoma and

fibrosarcomas (4%). The remainder of cases are several rare types of bone cancer [4,5]. In children and teenagers (those younger than 20 year), osteosarcoma (56%) and Ewing's sarcoma (34%) are much more common than chondrosarcoma (6%) [4,5]. The clinical treatment of primary malignant bone tumors or bone metastasis is currently based on therapeutic approaches encompassing systemic treatments, including chemotherapy, hormone therapy, immunotherapy, bone modifying agents, radiopharmaceuticals, and local treatments such as radiation therapy, radiofrequency ablation and surgery [7,8]. Unfortunately, to date, none of these therapeutic options have shown to exert a positive clinical impact on patients' survival [7,8]. Furthermore, the systemic administration of antitumor drugs and/or bone modifying agents and/or radiopharmaceuticals may negatively affect the normal metabolic turnover of bone tissue with detrimental consequences for cancer patients [9,10]. Hence, the need to identify new compounds that may be effective in preventing the growth and dissemination of malignant cells in the bone and, at the same time, are endowed with low toxicity and/or limited side effects [7,8,11]. In recent years an increasing number of preclinical studies have been directed towards the discovery and the development of new drugs based on natural products from plants environment [12-17]. The advantages of using natural products for therapeutic purposes in cancer treatment are manifold, as these compounds appear, in general, to be *i*) readily available, *ii*) non-toxic to normal human cells, *iii*) may act as multi-target agents since they may affect different signalling pathways that control cancer progression [12-18]. Among the natural compounds, phytochemicals such as polyphenols, terpenoids, alkaloids, phytosterols, and organosulfur compounds have been reported to produce beneficial effects on human health and, in particular, in pathological conditions such as inflammation and cancer [12-14]. In this setting, there is growing experimental evidence highlighting the fact that several extra virgin olive oil (EVOO)-derived phenolic compounds are endowed with anti-proliferative, anti-invasive and anti-metastatic properties [14,17-20]. Therefore, these molecules seem to be promising as potential chemo-preventive agents. In particular, oleuropein (OLe) (Fig.1), one of the main bioactive phenolic compounds present in olive leaves (*Olea europaea* L., Oleaceae), in unprocessed olive drupes and, in the aglycone form, in olive-oil exhibits a broad range of pharmacological properties [17,20-22], which may account for the therapeutic effects of this molecule observed in various human pathological conditions [14,18,20-22] including chronic inflammatory diseases associated with bone loss [20-22] and human cancers [17-19,23-25]. The finding that OLe may thwart the dissemination of cancer cells to distant organs [18,21-23] and exercises protective effects against the onset of various pathological conditions associated with bone loss [20-22] suggests the potential clinical usefulness of this molecule in the prevention and treatment of cancer-related bone diseases. In this review we discuss and critically appraise, in particular, the results from emergent data suggesting a potential role for OLe in the chemoprevention and clinical management of malignant bone tumors.

2. Oleuropein in human tumors

An increasing number of *vitro* studies have shown that **OLe** may inhibit the growth and survival of cancer cells from different human solid tumors including breast cancer [18,20,22-25] colorectal cancer [20,23-25], genito-urinary cancer [20,23-26], lung cancer [19,23-25], central nervous system tumors [19,20,23-25] melanoma [19,27,28], and haematological malignancies such as leukemias [19,20,29,30]. In line with these observations, *in vivo* studies have highlighted the fact that the administration of **OLe** to experimental animals may hinder the development of tumors such as skin cancer [31], soft tissue sarcoma [32], melanoma [27,28] or breast cancer [33-36] (Table 1) In particular, the administration of **OLe** to mice transplanted with B16F10 melanoma or MCF-7 human breast cancer cells, resulted in a significant inhibition of lymph-node or lung metastasis respectively [33-36] (Table 1). The numerous studies carried out in order to elucidate the specific mechanisms underlying the antitumor effects of **OLe** have highlighted the fact that this molecule, besides its inhibiting activity on tumor cell proliferation and survival [23-26], may also hinder other key steps of malignant progression such as cell migration [16,19,20,24,32,37-40], invasion[19,20,24,32,39,40,43], angiogenesis [20,24, 32,33,44,45] and dissemination of malignant cells to distant organs [15,20,24-26,28,32-36,46,47]. (Fig. 2) In this setting, molecular studies have revealed that **OLe** can modulate the expression and activity of several growth factors, hormones, cytokines, adhesion molecules and enzymes, which regulate different cellular processes involved in cancer progression including apoptosis [20,24,33,39,42,43,49,50], cell cycle progression [20,26,43,44,50], cell differentiation [20,24,30,51] and epithelial-to-mesenchymal transition (EMT) [20,37,52-56] (Fig. 2). These findings indicate the therapeutic potential of **OLe** as a chemo-preventive drug that might thwart early events in the metastatic process [24-28,35,36,44-48] (Fig. 2). However, at least *in vivo*, the probable contribution of a few active metabolites of **OLe**, endowed with anticancer activity such as hydroxytyrosol (Hyt), and **OLe**-aglycone (Ole A) (Fig. 1) may further account for the anticancer activity of **this molecule** [19,20,23-25,51,57]. In fact, experimental and clinical pharmacokinetic studies have shown that following its absorption in the intestinal tract, Ole undergoes phase I and phase II metabolic processes that generate several metabolites endowed with various pharmacological effects including **OLe**-A and Hyt [58-61]. In this setting, Mosele et al. [62] have provided evidence on the involvement of some lactic acid bacteria which can be found in human gastrointestinal tract (in particular *Lactobacillus plantarum*) in the metabolic conversion of **OLe** in **OLe**-A and then into Hyt.

3. Oleuropein and the bone microenvironment

Experimental and observational studies have provided evidence on the effectiveness of **OLe** in the prevention and treatment of various pathological conditions associated with bone loss [36,63-72]. These effects appear to be related to the modulating activity of this phenolic compound on multiple common signalling molecules underlying the

pathogenesis of malignant and non-malignant bone diseases [7,47,68,71]. On the basis of these observations it appears conceivable to speculate about a potential clinical role of **OLe** as a drug that might prevent the growth and dissemination of cancer cells in the bone [18,20,38,45-47,54,55,63] (Fig 2). In this context, compelling evidence shows that the protective effects exhibited by **OLe** on bone health appear to be due to the antioxidant [20,21,57-60] anti-inflammatory and immune-modulating properties of this molecule [20-22,67-71]. The antioxidant effects of **OLe** have been attributed to its ability to prevent the production of reactive oxygen species (ROS) and/or to act as radical chain breakers, and/or as a metal ion chelator [20,21,72]. Instead, the anti-inflammatory and immune-modulatory activities of **OLe** recognize multiple mechanisms and involve the capability of this compound to modulate the expression and activity of signalling molecules, such as growth factors [20,41,66,67,72-75], transcription factors [18,20,38,71,75-80], hormones [76,81,81], cytokines, chemokines [20,21,55,69,71,73,75] and enzymes [20,43,44,71,75,77], which, in physiological conditions, regulate the normal metabolic turnover of bone tissue while their expression proves to be deregulated in various pathological conditions associated with altered bone remodelling processes [20,44,45,71,82-84]. In particular, preclinical studies have highlighted the fact that the anti-inflammatory and immunomodulatory effects of **OLe** are the result of its inhibitory activity on nuclear factor- κ B (NF- κ B) and mitogen activated protein-kinase (MAPk) signalling pathways [20,66,71,79,84,85]. These phenomena eventuate in an attenuated production of various downstream molecular effectors modulating the immune-inflammatory response, such as tumor necrosis factor- α (TNF- α) [20,68,79,80,85,86], interleukin-1 β (IL-1 β) [20-22,71,85,86], interleukin-6 (IL-6) [20-22,71,86-89], interleukin-8 (IL-8) [8,20,21,47,68,72,88], monocyte chemo-attractant protein-1 (MCP-1) [89], prostaglandins (PGs), in particular prostaglandin E₂ (PGE₂), and enzymes such as cyclooxygenase-2 (COX-2) [20,24,44,70,71,85,86,90], matrix metalloproteinases (MMPs) [20,21,44,45,51,71] and the inducible form of nitric oxide synthase (iNOS) [20,65,71,80,85]. On the other hand, these findings are in line with the results from experimental studies indicating that the bone-protective effects of **OLe** appear to be related mainly to its modulating activity on inflammatory signalling pathways rather than directly on bone metabolism [63-68]. Nevertheless, recent findings have shown that **OLe** may facilitate osteoblast (OB) proliferation and differentiation [66,82,91-93], whilst it suppresses osteoclastogenesis [64,65,82,89-91]. These phenomena were associated with an increase in the expression levels of osteogenic transcription factors such as runt-related transcription factor 2 (Runx2) and osterix, with an increased osteoprotegerin/receptor activator of nuclear factor kappa-B ligand (OPG/RANKL) expression ratio [91,94] and the up-regulation of other osteoblastic markers such as type I collagen, osteocalcin or alkaline phosphatase [67,77,94,95]. In particular, **OLe** elicits osteoprotective effects by fostering the differentiation of human bone marrow mesenchymal stem cells into osteoblasts [91,96] and by reducing tartrate-resistant acid phosphatase (TRAP) activity in osteoclasts formed from mouse spleen cells [65]. Furthermore, recent *in vitro* studies have shown that **OLe** modulates

osteoblastogenesis by suppressing the expression and activity of perioxosome proliferator-activated receptor γ (PPAR γ), which functions as a transcriptional regulator of adipocyte differentiation and inhibitor of OB [91,96-98].

4. Oleuropein and malignant bone disease progression

Most of the molecular pathways involved in pathological conditions associated with disorders of bone remodeling and which may be targeted by Ole, have also been shown to contribute to the homing and growth of cancer cells in the bone [20,75,99-103] (Fig 3). The findings that various chronic inflammatory bone diseases and bone-related cancers share common molecular mechanisms underlying their pathogenesis [20,75,83,99-103] may, in part, explain the reason why chronic inflammation is actually considered as one of the main pathological processes that may foster the homing and growth of tumor cell in the bone tissue [20,103,104] (Fig. 3). On the other hand, these observations are consistent with the results from several experimental studies showing that chronic inflammation may function as a promoting factor in the early stages of a malignant progression [101, 103-107] (Fig. 3). These data may also, partially, explain the findings that disturbances of the bone microenvironment, induced by osteoporosis and/or other immunoinflammatory bone diseases, appear to facilitate the growth and dissemination of some types of cancer cells in the bone tissue [106-113]. Ultimately, these observations indicate that Ole might well exert its potential chemopreventive and therapeutic effects on cancer-related bone disease through multiple mechanisms involved in early events of cancer progression and in the regulation of bone remodelling processes (Fig. 2, Fig. 3).

4.1. Oleuropein and tumor cell proliferation and survival

A consistent body of *in vitro* studies have demonstrated that Ole may hinder the proliferation, survival and migration of malignant cells from human primary bone tumor, such as osteosarcoma [22,38,63,114], and from human tumors which preferentially metastasize to the bone, such as breast cancer [20,23-25,38-40,45,49,115], prostate cancer [20,23-25,116,117] and lung cancer [23-25,117-120]. The mechanisms underlying the anti-proliferative effects of Ole on osteosarcoma cells still need to be fully understood [37,63,114,116]. Conversely, accumulating evidence shows that Ole can inhibit the growth and survival of human breast, prostate or lung cancer cell lines by *i*) delaying or halting different phases of cell cycle progression [20,26,119-123], *ii*) by activating different pro-apoptotic molecular signalling pathways [20,26,40,45,76,113,116-118,123-127] and/or *iii*) by inhibiting the expression of the transcription factor NF- κ B which regulates many genes driving cancer growth and progression (Table 2).

4.2. Oleuropein and Epithelial to Mesenchymal transition (EMT).

OLE has been demonstrated to modulate *in vitro* multiple signalling pathways associated with the different types of EMT, including type 3 EMT [36,46,51-54,119], which has been recognized as an initial, critical event for metastasis formation by individually-invading carcinoma cells [119,122,124,128-131]. Almost all the key regulators of EMT are expressed in the bone microenvironment where their cross-talk contributes to favouring the homing and growth of cancer cells [37,47,51-54,129-131] (Fig. 2, Table 3). In particular, these studies have highlighted the promoting role of signalling pathways such as TGF- β /BMP[52,74,132-134], MAP/ERK[52,120,125,134-136], PI3K/AKT [52,105,137], Wnt/ β -catenin [54-56,74,125-131], NF- κ B [128,133,137,140,141], HIF [36,142], and PTHrP [142,143] on the growth and dissemination of cancer cells in the bone (Table 3). **OLE** has been reported to preserve bone health by interfering with various cellular processes triggered by these molecular pathways [20,66,68,71,85,87,92-94,135]. On the basis of these findings, numerous preclinical investigations have been undertaken to assess the therapeutic effectiveness of this molecule, alone or in association with antitumor drugs in the prevention and treatment of several human solid tumors [13,15,43,45,50-53,71,111-113,134].

4.3. Oleuropein and tumor angiogenesis.

Tumor angiogenesis is a crucial event for the growth and dissemination of circulating tumor cells to distant organs including the bones [128-130]. Several experimental observations highlight the fact that **OLE** appears to thwart tumor neovascularization by decreasing the expression levels of various signaling molecules involved in this process [32,33,36,44, 89,118,123-129,144] (Fig. 2, Table 3). In particular, these investigations have shown that the treatment of endothelial cells [75,76,89,90,143,144] and/or human cancer cells from different tumors [34,35,76,124,125,143,145], with **OLE** results in a decrease in the expression level of specific growth factors and receptors such as VEGF and VEGFR2 [31,33,36,75,90,118,129], pro-inflammatory cytokines such as IL-6, IL-8, IL-1 β TNF- α [19,20,22,48,80,90,143-146], adhesion molecules such as vascular cellular adhesion molecule-1 (VCAM-1) intercellular adhesion molecule-1 (ICAM-1) and E-selectin [20,22,39,52,71,75,145,146], inflammatory and proteolytic enzymes such as COX-2 [31,39,44,71,75,98] and MMPs [31,37,44,45,52,71,75,118,130] (Table 3). Most of these effects can also be attributed to the direct up-stream inhibiting activity of **OLE** on NF- κ B, a transcription factor that modulates the expression of various genes and related signalling pathways involved in almost all the hallmarks of malignant progression [20,39,79,131,132, 143,147].

4.4. Oleuropein and tumor cell migration, adhesion and invasion.

Besides tumor neo-vascularization, some of the molecules mentioned above, such as pro-inflammatory cytokines, adhesion molecules or COX-2 and MMPs, also modulate the adhesion, migration and invasion of tumor cells [44,147-153] (Table 3). For instance, *in vitro* studies have demonstrated that adhesion molecules such as VCAM-1, ICAM-1 and E-selectin contribute to the escaping of tumor cells from dormancy and bone metastasis formation [146,150,152-154]. Moreover, emerging evidence has shown that **OLe** may indirectly affect the adhesive properties of tumor cells by inhibiting protein tyrosine phosphatase B1 (PTPB1), a regulator of cell adhesion and migration in normal and cancer cells [155,156]. Furthermore, MMP-2 and MMP-9 have been reported to actively contribute to the migratory activity of tumor cells, to the degradation of the extracellular matrix (ECM) and to regulating the expression of cytoskeletal proteins, growth factors, cytokines and chemokines, which favour the preferential adhesion of disseminating cancer cells to the ECM underlying epithelial cells and vascular endothelial cells [44,134,135,150,152]. Consistent with these observations, recent *in vitro* studies demonstrated that the anti-invasive activity induced by **OLe** on tumor cells paralleled the down-regulation of MMP-9 and MMP-2 expression and the up-regulation of their intracellular inhibitors TIMP-1,-3,-4 [23,31,44,45,55,71,132,143,152] (Table 3)

4.5. **Oleuropein** and tumor-bone niche

Most of the signalling molecules mentioned above have also been shown to contribute to the formation of a permissive microenvironment, the so-called “(pre)metastatic niche”, which supports the homing, colonization and growth of cancer cells in the bone and, their subsequent survival, dormancy and resistance to clinical treatments [157-159] (Fig. 2, Fig.3, Table 3). Therefore, the targeting of these signalling molecules may be regarded as a novel potential therapeutic tool in the prevention and, eventually, eradication of the establishment of malignant cells in the bone. In this regard, preclinical evidence has suggested the possibility that **OLe** might thwart the formation of the (pre)metastatic bone niche (Fig. 2). For instance, pro-inflammatory interleukins (ILs) such as IL-6, IL-8, IL-1 β , TNF- α have been shown actively to contribute to the homing of malignant cells in the bone [19,20,90,143,147,160-163] (Fig. 3) **OLe** can down-regulate the expression of these cytokines in several pathological conditions associated with an altered bone resorption and in early events of the metastatic process, namely EMT transition [37,52-55,130,147] and angiogenesis [20,36,75,143,148] (Fig. 3, Table 3). Moreover, proteolytic enzymes such as MMP-2 and MMP-9 have also been shown to contribute to the formation of the “(pre)metastatic niche” by modifying the ECM structure and by promoting the release growth factors and chemokines which fuel the “vicious cycle” of bone metastasis [44,90,130-133,147,152] (Table 3). As described above, **OLe** has been proven to inhibit the expression levels of MMP-2 and MMP-9 in human metastatic tumor cells while it may increase the expression of intracellular inhibitors of these enzymes, namely TIMP-1, TIM,-3 and TIMP-4 [45,51,54,75,132,152]. Moreover, **OLe** has been reported to indirectly

affect the expression and activity of these proteolytic enzymes through the up-stream inhibition NF- κ B [39,51,54,71,148,149,165] (Fig. 3). Furthermore, experimental evidence has shown that, the osteo-protective effects of **OLE** also appear to be due to its inhibiting activity on RANKL-induced NF- κ B activation, a phenomenon which facilitates osteoclast differentiation and bone remodeling [68,70,93,94,161,166]. On the other hand, *in vitro* studies have highlighted the fact that the RANKL/RANK/OPG system appears to exert an active role in the formation of the pre-metastatic niche and in fostering the metastatic spread to bone [7,100,149,153,157,165,166]. On the basis of these observations it is conceivable to hypothesize a potential clinical role of **OLE** as a novel drug that may thwart the formation of a permissive microenvironment enabling tumor cells to grow and disseminate in the bone [160,165].

5. Discussion

Despite the therapeutic advances registered in recent years, the current treatments for bone-related cancers based on the systemic administration of anti-resorptive drugs and/or anti-tumour agents and/or radiopharmaceutical, still experience several drawbacks, including the negative effects elicited by these agents on the normal bone metabolism. These effects may result in detrimental consequences for cancer patients, such as increased risk of osteoporosis and fractures [7,8,9]. Furthermore, none of the currently available clinical treatments have been shown to exert a positive impact on patients' survival [7,8,9]. Hence, the need to develop new molecules endowed with both antitumor activity and low toxicity and/or limited side effects [12-17,24]. In this setting, an increasing number of preclinical studies are currently aimed at identifying and/or developing new molecules based on natural products from plants [12-17]. In particular, a notable number of preclinical investigations have provided convincing evidence on the chemo-preventive effects of several extra virgin olive oil (EVOO)-derived phenolic compounds, including oleuropein, in combatting the growth of several human solid tumors [17-20,22,23,35,36]. Furthermore, experimental and observational studies have shown that **OLE** exhibits protective effects on bone health due to its property of modulating various molecular signalling pathways involved in the regulation of bone resorption in normal and pathological conditions, including bone-related tumors [7,16,65-68,75,107-109,143]. Therefore, the results emerging from these studies make this **molecule** an attractive candidate as a novel potential agent in the prevention and treatment of bone-related cancers [17-20,54,45,53,65,66,143]. Interestingly, as **OLE** appears to be endowed with both anti-tumor [19,20,23,24,25,35,53,142] and bone-anabolic effects [20,65,66,81,82,91-94,98,143], this molecule might be regarded as a potential, additional therapeutic option, in the treatment of specific tumors such as multiple myeloma (MM), which is known to cause severe osteolytic bone loss due to hyper-activation of osteoclasts and suppression of the differentiation of bone marrow mesenchymal stem cells (BMSCs) into functional osteoblasts [91,96,97,167,168] (Table 3). Furthermore these findings also indicate the administration of **OLE** may be effective in the prevention of bone loss induced by the systemic

administration of antitumor drugs and/or bone modifying agents and/or radiopharmaceuticals [8,9,10,19]. Although, the data from *in vitro* studies are promising in this regard, unfortunately, there is still a lack of *in vivo* studies with the specific aim to evaluate the therapeutic effectiveness of this compound on primary bone tumors or metastatic bone diseases by using suitable experimental animal tumor models. Nevertheless, the accumulating *in vitro* evidence on the capability of OLe to target different molecular pathways involved in the pathogenesis of cancer-related bone diseases, prompts the need for extensive future experimental and clinical investigations.

6. Conclusions

Oleuropein has been reported to target several signaling pathways involved in the modulation of bone remodelling processes in normal and pathological conditions including malignant bone diseases [45,52,53,68,144,161]. These observations suggest the potential usefulness of OLe, administered alone and/or in association with cytotoxic drugs and/or inhibitors of bone metabolism and/or radiopharmaceuticals to prevent the growth and spreading of cancer cells in the bone [19,20,29,37,38,168]. These drug associations might also result in improved therapeutic activity, in a reduction of side effects and in a decreasing probability of developing drug resistance [19,20,24,28,37,76,169,170]. However, there are several drawbacks in transferring these experimental in clinic data. One of the major drawbacks regards the bioavailability and, more in general, the pharmacokinetic properties of OLe [171-174]. The rate of absorption and bioavailability of this molecule appears to be influenced by several factors, such as the route and form of administration, interaction with food, age, sex, different extraction and analytical methods used [171-174]. Experimental and clinical studies have shown that the oral administration of OLe results in rapid absorption, metabolism and renal clearance [117,171-174]. Furthermore, following absorption this compound undergoes extensive phase I and phase II metabolic processes [56,169,173,174]. The potential therapeutic effectiveness of OLe is closely related to the possibility that this molecule may reach in adequate concentrations its specific molecular targets in human tissues [20,143]. Therefore, these phenomena may affect the bioavailability of OLe and its systemic transfer at adequate concentrations to the target tissues, ultimately blunting the therapeutic effectiveness of this compound. However, in this context, data on the effective organ distribution of OLe in human tissues including the bone are currently lacking. Nevertheless, a promising strategy to overcome these hurdles may rely on the development of new semi-synthetic OLe derivatives endowed with better bioavailability and possibly, improved biological activity and spectrum of activity [19,20,160,175-178] and by the use of novel drug delivery systems based on nanotechnology [179-184]. In this latter case, preclinical *in vivo* studies have reported that the nano-encapsulation of OLe could prolong circulation, improve localization, enhance efficacy and reduce the chances of multidrug resistance [117,183-185]. In conclusion, the increasing number of preclinical *in vitro* studies suggesting a potential chemo-preventive role

of **OLE** in cancer-related bone disease, warrant future studies to assess the impact of this phenolic compound on the clinical management and outcome of patients with primary tumors that reside in, or form metastasis in the bone [13,15,36,43,54,51,67,134,151,156].

Acknowledgements

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflict of Interest statement

The authors declare that there are no conflicts of interest.

References

- [1] R.E. Coleman, Clinical features of metastatic bone disease and risk of skeletal morbidity, *Clin. Cancer Res.* 12(2006) 6243s–6249s, doi: 10.1158/1078-0432.CCR-06-0931.
- [2] J.F. Huang, J. Shen, X. Li, R. Rengan, N. Silvestris, M. Wang, L. Derosa, X. Zheng, A. Belli, X. L. Zhang, Y.M. Li, A. Wu, Incidence of patients with bone metastases at diagnosis of solid tumors in adults: a large population-based study, *Ann. Transl. Med.* 8(7) (2020): 482. doi:10.21037/atm.2020.03.55.
- [3] R.K. Hernandez, S.W. Wade, A. Reich, M. Pirolli, A. Liede, G.H. Lyman, Incidence of bone metastases in patients with solid tumors: analysis of oncology electronic medical records in the United States, *BMC Cancer.* 18(1)(2018) : 44. doi: [10.1186/s12885-017-3922-0](https://doi.org/10.1186/s12885-017-3922-0).
- [4] R.L. Siegel, K.D. Miller, A. Jemal, Cancer statistics, 2019, *CA Cancer J. Clin.* 69(1)(2019):7-34. doi:10.3322/caac.21551.
- [5] SEER Cancer Stat Facts: Bone and Joint Cancer. National Cancer Institute. Bethesda, MD, [HYPERLINK "https://seer.cancer.gov/statfacts/html/bones.html"](https://seer.cancer.gov/statfacts/html/bones.html) . <https://seer.cancer.gov/statfacts/html/bones.html>.
- [6] R. Fonseca, S. Abouzaid, M. Bonafede, Q. Cai, K. Parikh, L. Cosler, P. Richardson, Trends in overall survival and costs of multiple myeloma, 2000-2014, *Leukemia.* 31(9)(2017)1915-1921. doi:10.1038/leu.2016.380.
- [7] S. D'Oronzo, R. Coleman, J. Brown, F. Silvestris, Metastatic bone disease: Pathogenesis and therapeutic options. Up-date on bone metastasis management, *J. Bone Oncol.* 15(2019)4, <https://doi.org/10.1016/j.jbo.2018.10.004>. eCollection 2019 Apr.
- [8] I. Makhoul, C.O. Montgomery, D. Gaddy, L. J. Suva, The best of both worlds - managing the cancer, saving the bone, *Nat. Rev. Endocrinol.* 12 (2016) 29–42, <https://doi.org/10.1038/nrendo.2015.185>.

- [9] M.K. Skjødt, M. Frost, B. Abrahamsen, Side effects of drugs for osteoporosis and metastatic bone disease, *Br. J. Clin. Pharmacol.* 85 (2019) 1063–1071, <https://doi.org/10.1111/bcp.13759>.
- [10] P.G. Turner, J.M. O'Sullivan, ^{223}Ra and other bone-targeting radiopharmaceuticals—the translation of radiation biology into clinical practice, *Br. J. Radiol.* 88 (2015) 20140752, <https://doi.org/10.1259/bjr.20140752>.
- [11] G. Leto, Current status and future directions in the treatment of bone metastases from breast cancer, *Clin. Exp. Pharmacol. Physiol.* 46 (2019) 968–971, <https://doi.org/10.1111/1440-1681.13139>.
- [12] L. Sun, W. Zhou, H. Zhang, Q. Guo, W. Yang, B. Li, Z. Sun, S. Gao, R. Cui, Modulation of Multiple Signaling Pathways of the Plant-Derived Natural Products in Cancer. *Front Oncol* 9(2019). 1153. <https://doi.org/10.3389/fonc.2019.01153>.
- [13] S. Sur, R.B. Ray, Bitter Melon (*Momordica Charantia*), a Nutraceutical Approach for Cancer Prevention and Therapy. *Cancers (Basel)*. 2020;12(8):E2064. Published 2020 Jul 27. doi:10.3390/cancers12082064
- [14] B. Salehi, P. Zucca, M. Sharifi-Rad, R. Pezzani, S. Rajabi, W.N. Setzer, E.M. Varoni, M. Iriti, F. Kobarfard, J. Sharifi-Rad, Phytotherapeutics in cancer invasion and metastasis, *Phytother. Res.* 32 (2018) 1425–1449, <https://doi.org/10.1002/ptr.6087>.
- [15] P. Juárez, Plant-derived anticancer agents: a promising treatment for bone metastasis, *Bonekey Rep.* 3 (2014) 599, <https://doi.org/10.1038/bonekey.2014.94>.
- [16] A. Kapinova, P. Kubatka, A. Liskova, D. Baranenko, P. Kruzliak, M. Matta, D. Busselberg, B. Malicherova, A. Zulli, T.K. Kwon, E. Jezkova, D. Blahutova, P. Zubor, J. Danko, Controlling metastatic cancer: the role of phytochemicals in cell signaling, *J. Cancer Res. Clin. Oncol.* 145 (2019) 1087–1109, <https://doi.org/10.1007/s00432-019-02892-5.h>.
- [17] A. Maruca, R. Catalano, D. Bagetta, F. Mesiti, F.A. Ambrosio, I. Romeo, F. Moraca, R. Rocca, F. Ortuso, A. Artese, G. Costa, S. Alcaro, A. Lupia, The Mediterranean Diet as source of bioactive compounds with multi-targeting anti-cancer profile., *Eur. J. Med. Chem.* 181 (2019) 111579, doi: 10.1016/j.ejmech.2019.111579. Epub 2019 Jul 3.
- [18] I. Casaburi, F. Puoci, A. Chimento, R. Sirianni, C. Ruggiero, P. Avena, V. Pezzi, Potential of olive oil phenols as chemopreventive and therapeutic agents against cancer: a review of in vitro studies, *Mol. Nutr. Food Res.* 57 (2013) 71–83, <https://doi.org/10.1002/mnfr.201200503>.
- [19] J. Torić, A.K. Marković, C.J. Brala, M. Barbarić, Anticancer effects of olive oil polyphenols and their combinations with anticancer drugs, *Acta Pharm.* 69 (2019) 461–482, <https://doi.org/10.2478/acph-2019-0052>.
- [20] M.L. Castejón, T. Montoya, C. Alarcón-de-la-Lastra, M. Sánchez-Hidalgo, Potential Protective Role Exerted by Secoiridoids from *Olea europaea* L. in Cancer, Cardiovascular, Neurodegenerative, Aging-Related, and Immunoinflammatory Disease, *Antioxidants (Basel)*. 9 (2) (2020) 149, <https://doi.org/10.3390/antiox9020149>.
- [21] I. Hassen, H. Casabianca, K. Hosni, Biological activities of the natural antioxidant oleuropein: Exceeding the expectation-A mini-review, *J. Funct. Foods.* 18 (2015) 926–940, <https://doi.org/10.1016/j.jff.2014.09.001>.

- [22] S. Rigacci, M. Stefani, Nutraceutical Properties of Olive Oil Polyphenols. An Itinerary from Cultured Cells through Animal Models to Humans, *Int. J. Mol. Sci.* 17 (2016), <https://doi.org/10.3390/ijms17060843>.
- [23] H. Shamshoum, F. Vlaveciski, E. Tsiani, Anticancer effects of oleuropein, *Biofactors*. 43 (2017) 517–528, <https://doi.org/10.1002/biof.1366>.
- [24] M. Imran, M. Nadeem, S.A. Gilani, S. Khan, M.W. Sajid, R.M. Amir, Antitumor Perspectives of Oleuropein and Its Metabolite Hydroxytyrosol: Recent Updates, *J. Food Sci.* 83(2018) 1781–1791, <https://doi.org/10.1111/1750-3841.14198>.
- [25] A. Ahmad Farooqi, S. Fayyaz, A.S. Silva, A. Sureda, S.F. Nabavi, A. Mocan, S.M. Nabavi, A. Bishayee, Oleuropein and Cancer Chemoprevention: The Link is Hot, *Molecules*. 22(2017), <https://doi.org/10.3390/molecules22050705>.
- [26] R. Acquaviva, C. Di Giacomo, V. Sorrenti, F. Galvano, R. Santangelo, V. Cardile, S. Gangia, N. D’Orazio, N.G. Abraham, L. Vanella, Antiproliferative effect of oleuropein in prostate cell lines, *Int. J. Oncol.* 41 (2012) 31–38, <https://doi.org/10.3892/ijo.2012.1428>.
- [27] S.A. Mijatovic, G.S. Timotijevic, D.M. Miljkovic, J.M. Radovic, D.D. Maksimovic-Ivanic, D.P. Dekanski, S.D. Stosic-Grujicic, Multiple antimelanoma potential of dry olive leaf extract, *Int. J. Cancer*. 128 (2011) 1955–1965, <https://doi.org/10.1002/ijc.25526>.
- [28] J. Ruzzolini, S. Peppicelli, E. Andreucci, F. Bianchini, A. Scardigli, A. Romani, G. la Marca, C. Nediani, L. Calorini, Oleuropein, the Main Polyphenol of *Olea europaea* Leaf Extract, Has an Anti-Cancer Effect on Human BRAF Melanoma Cells and Potentiates the Cytotoxicity of Current Chemotherapies. *Nutrients*. 10 (2018), <https://doi.org/10.3390/nu10121950>.
- [29] M. Crescimanno, M.V. Sepporta, E. Tripoli, C. Flandina, M. Giammanco, F.M. Tumminello, D. Di Majo, M. Tolomeo, M. La Guardia, G. Leto, Effects of extra virgin olive oil phenols on HL60 cell lines sensitive and resistant to anthracyclines, *J. Biol. Res. - Bollettino della Società Italiana di Biologia Sperimentale*. 82, 1 (Jan. 2009), <https://doi.org/10.4081/jbr.2009.4729>.
- [30] S. Fagan, N. Fulco, J. Gilbertson, A.R. Van Dyke, S.A. Phelan, Anti-cancer effects of oleuropein olive leaf extract in K562 leukemia cells, *Cancer Res.* 78 (13 Suppl.) (2018) 4388, <https://doi.org/10.1158/1538-7445.am2018-4388>.
- [31] Y. Kimura, M. Sumiyoshi, Olive Leaf Extract and Its Main Component Oleuropein Prevent Chronic Ultraviolet B Radiation-Induced Skin Damage and Carcinogenesis in Hairless Mice, *J. Nutr.* 139 (2009) 2079–2086, <https://doi.org/10.3945/jn.109.104992>.
- [32] H.K. Hamdi, R. Castellon, Oleuropein, a non-toxic olive iridoid, is an anti-tumor agent and cytoskeleton disruptor, *Biochem. Biophys. Res. Commun.* 334 (2005) 769–778, <https://doi.org/10.1016/j.bbrc.2005.06.161>.

- [33] R. Liman, F.K. Coban, I.H. Cigerci, I. Bulduk, S. Bozkurt, Antiangiogenic and Apoptotic Effects of Oleuropein on Breast Cancer Cells, *Br. J. Pharm. Res.* 16 (2017) 1-10, <https://doi.org/10.9734/bjpr/2017/33403>.
- [34] M.H. Elamin, A.B. Elmahi, M.H. Daghestani, E.M. Al-Olayan, R.A Al-Ajmi, A.F. Alkhuriji, S. S.Hamed, M. .F Elkhadragy, Synergistic Anti-Breast-Cancer Effects of Combined Treatment With Oleuropein and Doxorubicin In Vivo, *Altern. Ther. Health Med.* 25(3) (2019) 17-24.
- [35] M.V. Sepporta, R. Fuccelli, P. Rosignoli, G. Ricci, M. Servili, G. Morozzi, R. Fabiani, Oleuropein inhibits tumour growth and metastases dissemination in ovariectomised nude mice with MCF-7 human breast tumour xenografts, *J. Funct. Foods.* 8 (2014) 269–273, <https://doi.org/10.1016/j.jff.2014.03.027>.
- [36] H. Song, D.Y. Lim, J.I. Jung, H.J. Cho, S.Y. Park, G.T. Kwon, Y.H. Kang, K.W. Lee, M.S. Choi, and J. Park, Dietary oleuropein inhibits tumor angiogenesis and lymphangiogenesis in the B16F10 melanoma allograft model: a mechanism for the suppression of high-fat diet-induced solid tumor growth and lymph node metastasis, *Oncotarget.* 8(19) (2017) 32027- 32042, doi:10.18632/oncotarget.16757.
- [37] J. Choupani, M.R. Alivand, S. M Derakhshan, M. Zaeifzadeh, M.S. Khaniani, Oleuropein inhibits migration ability through suppression of epithelial-mesenchymal transition and synergistically enhances doxorubicin-mediated apoptosis in MCF-7 cells, *J. Cell Physiol.* 234 (2019) 9093–9104, <https://doi.org/10.1002/jcp.27586>.
- [38] P. Przychodzen, R. Wyszowska, M. Gorzynik-Debicka, T. Kostrzewa, A. Kuban-Jankowska, M. Gorska-Ponikowska, Anticancer Potential of Oleuropein, the Polyphenol of Olive Oil, With 2-Methoxyestradiol, Separately or in Combination, in Human Osteosarcoma Cells, *Anticancer Res.* 39 (2019) 1243–1251, <https://doi.org/10.21873/anticancer.13234>.
- [39] L. Liu, K.S. Ahn, M.K. Shanmugam, H. Wang, H. Shen, F. Arfuso, A. Chinnathambi, S.A. Alharbi, Y. Chang, G. Sethi, F.R. Tang, Oleuropein induces apoptosis via abrogating NF- κ B activation cascade in estrogen receptor-negative breast cancer cells, *J. Cell Biochem.* 120 (2019) 4504–4513, <https://doi.org/10.1002/jcb.27738>.
- [40] H.Y. Lu, J.S. Zhu, Z. Zhang, W.J. Shen, S. Jiang, Y.F. Long, B. Wu, T. Ding, F. Huan, S.L. Wang, Hydroxytyrosol and Oleuropein Inhibit Migration and Invasion of MDA-MB-231 Triple-Negative Breast Cancer Cell via Induction of Autophagy, *Anticancer Agents Med. Chem.* 19 (2019) 1983–1990, <https://doi.org/10.2174/1871520619666190722101207>.
- [41] G. Aktas and H. Ayan, Oleuropein: A Potential Inhibitor for Prostate Cancer Cell Motility by Blocking Voltage-Gated Sodium Channels, *Nutr. Cancer* (2020), doi: 10.1080/01635581.2020.1807575.
- [42] M. Abtin, M.R. Alivand, M.S. Khaniani, M. Bastami, M. Zaeifzadeh, S.M. Derakhshan, Simultaneous downregulation of miR-21 and miR-155 through oleuropein for breast cancer prevention and therapy, *J. Cell Biochem.* 119 (2018) 7151–7165, <https://doi.org/10.1002/jcb.26754>.

- [43] M. Liu, J. Wang, B. Huang, A. Chen, X. Li, Oleuropein inhibits the proliferation and invasion of glioma cells via suppression of the AKT signaling pathway, *Oncol. Rep.* 36 (2016) 2009–2016, <https://doi.org/10.3892/or.2016.4978>.
- [44] M. Seçme, C. Eroğlu, Y. Dodurga, G. Bağcı, Investigation of anticancer mechanism of oleuropein via cell cycle and apoptotic pathways in SH-SY5Y neuroblastoma cells, *Gene.* 585 (2016) 93–99, <https://doi.org/10.1016/j.gene.2016.03.038>.
- [45] E. Scoditti, N. Calabriso, M. Massaro, M. Pellegrino, C. Storelli, G. Martines, R. De Caterina, M.A. Carluccio, Mediterranean diet polyphenols reduce inflammatory angiogenesis through MMP-9 and COX-2 inhibition in human vascular endothelial cells: a potentially protective mechanism in atherosclerotic vascular disease and cancer, *Arch. Biochem. Biophys.* 527 (2012) 81–89, <https://doi.org/10.1016/j.abb.2012.05.003>.
- [46] Z.K. Hassan, M.H. Elamin, M.H. Daghestani, S.A. Omer, E.M. Al-Olayan, M.A. Elobeid, P. Virk, O.B. Mohammed, Oleuropein induces anti-metastatic effects in breast cancer, *Asian Pac. J. Cancer Prev.* 13 (2012) 4555–4559, <https://doi.org/10.7314/apjcp.2012.13.9.4555>.
- [47] Q. Zhou, L.L. Bennett, S. Zhou, Multifaceted ability of naturally occurring polyphenols against metastatic cancer, *Clin. Exp. Pharmacol. Physiol.* 43 (2016) 394–409, <https://doi.org/10.1111/1440-1681.12546>.
- [48] S. Fayyaz, T. Aydin, A. Cakir, M.L. Gasparri, P.B. Panici, A.A. Farooqi, Oleuropein Mediated Targeting of Signaling Network in Cancer, *Curr. Topics Med. Chem.* 16 (2016) 2477–248, <https://doi.org/10.2174/1568026616666160212123706>.
- [49] C.D. Goldsmith, D.R. Bond, H.Jankowski, J. Weidenhofer, C.E. Stathopoulos, P.D. Roach, C.J. Scarlett CJ, The Olive Biophenols Oleuropein and Hydroxytyrosol Selectively Reduce Proliferation, Influence the Cell Cycle, and Induce Apoptosis in Pancreatic Cancer Cells. *Int. J. Mol. Sci.* 19(7) (2018) 1937, doi:10.3390/ijms19071937.
- [50] S. Bayat, S.M. Derakhshan, N.M. Derakhshan, M.S. Khaniani, M.R. Alivand, Downregulation of HDAC2 and HDAC3 via oleuropein as a potent prevention and therapeutic agent in MCF-7 breast cancer cells, *J. Cell. Biochem.* 120 (2019) 9172–9180, <https://doi.org/10.1002/jcb.28193>.
- [51] I. Samet, J. Han, L. Jlaiel, S. Sayadi, H. Isoda, Olive (*Olea europaea*) Leaf Extract Induces Apoptosis and Monocyte/Macrophage Differentiation in Human Chronic Myelogenous Leukemia K562 Cells: Insight into the Underlying Mechanism, *Oxid. Med. Cell. Longev.* (2014) 927619, doi: 10.1155/2014/927619.
- [52] M. Dell'Agli, R. Fagnani, G. V. Galli, O. Maschi, F. Gilardi, S. Bellosta, M. Crestani, E. Bosisio, E. De Fabiani, D. Caruso, Olive Oil Phenols Modulate the Expression of Metalloproteinase 9 in THP-1 Cells by Acting on Nuclear Factor-kappa B Signaling, *J. Agric. Food Chem.* 58 (2010) 2246–2252, <https://doi.org/10.1021/jf9042503>.

- [53] A. Vazquez-Martin, S. Fernandez-Arroyo, S. Cufi, C. Oliveras-Ferraros, J. Lozano-Sanchez, L. Vellon, V. Micol, J. Joven, A. Segura-Carretero, J.A. Menendez, Phenolic Secoiridoids in Extra Virgin Olive Oil Impede Fibrogenic and Oncogenic Epithelial-to-Mesenchymal Transition: Extra Virgin Olive Oil As a Source of Novel Antiaging Phytochemicals, *Rejuvenation Res.* 15 (2012) 3–21, <https://doi.org/10.1089/rej.2011.1203>.
- [54] R.A. Razali, Y. Lokanathan, M.D. Yazid, A.S. Ansari, A. Bin Saim, R.B.H. Idrus, Modulation of Epithelial to Mesenchymal Transition Signaling Pathways by *Olea Europaea* and Its Active Compounds, *Int. J. Mol. Sci.* 20(14) (2019) 3492, <https://doi.org/10.3390/ijms20143492>.
- [55] Y.Q. Ci, J.P. Qiao, M. Han, Molecular Mechanisms and Metabolomics of Natural Polyphenols Interfering with Breast Cancer Metastasis, *Molecules.* 21 (2016) 1634, <https://doi.org/10.3390/molecules21121634>.
- [56] H. Amawi, C.R. Ashby, T. Samuel, R. Peraman, A.K. Tiwari, Polyphenolic Nutrients in Cancer Chemoprevention and Metastasis: Role of the Epithelial-to-Mesenchymal (EMT) Pathway, *Nutrients.* 9 (2017), <https://doi.org/10.3390/nu9080911>.
- [57] F. Xu, Y. Li, M. Zheng, X. Xi, X. Zhang, C. Han, Structure Properties, Acquisition Protocols, and Biological Activities of Oleuropein Aglycone, *Front. Chem.* 6 (2018) 239. doi: 10.3389/fchem.2018.00239. eCollection 2018.
- [58] M.P. Carrera-González, M.J. Ramírez-Expósito, M.D. Mayas, J. M. Martínez-Martos, Protective role of oleuropein and its metabolite hydroxytyrosol on cancer, *Trends Food Sci. Technol.* 31 (2013) 92-99. doi: 10.1016/j.tifs.2013.03.003.
- [59] H.K. Obied, P.D. Prenzler, S.H. Omar, R. Ismael, M. Servili, S. Esposito, A. Taticchi, R. Selvaggini, S. Urbani, Pharmacology of Olive Biophenols, in: J.C. Fishbein, J.M. Heilman (Eds), *Adv. in Mol. Toxicol.* Elsevier: Amsterdam, The Netherlands, 2012; Vol. 6, pp. 195–223.
- [60] M.N. Vissers, P.L. Zock, A.J. Roodenburg, R. Leenen, M.B. Katan, Olive oil phenols are absorbed in humans, *J. Nutr.* 132(3) (2002) 409-417, doi:10.1093/jn/132.3.409.
- [61] R. García-Villalba, M. Larrosa, S. Possemiers, F. A. Tomás-Barberán, J. C. Espín, Bioavailability of phenolics from an oleuropein-rich olive (*Olea europaea*) leaf extract and its acute effect on plasma antioxidant status: comparison between pre- and postmenopausal women, *Eur. J. Nutr.* 53 (2014) 1015–1027, <https://doi.org/10.1007/s00394-013-0604-9>.
- [62] J.I. Mosele, S. Martín-Peláez, A. Macià, M. Farràs, R.-M. Valls, U. Catalán, M.-J. Motilva, Faecal microbial metabolism of olive oil phenolic compounds: In vitro and in vivo approaches, *Mol. Nutr. Food Res.* 58 (2014) 1809–1819. doi: 10.1002/mnfr.20140012.

- [63] J.M. Morana, O. Leal-Hernande, M.L. Canal-Macías, R. Roncero-Martin, R. Guerrero-Bonmatty, I. Aliaga, J.D. Zamorano, Antiproliferative Properties of Oleuropein in Human Osteosarcoma Cells, *Nat. Prod. Commun.* 11 (2016) 491–492, <https://www.ncbi.nlm.nih.gov/pubmed/27396201>.
- [64] Puel, J. Mathey, A. Agalias, S. Kati-Coulibaly, J. Mardon, C. Obled, M.J. Davicco, P. Lebecque, M.N. Horcajada, A.L. Skaltsounis, V. Coxam, Dose-response study of effect of oleuropein, an olive oil polyphenol, in an ovariectomy/inflammation experimental model of bone loss in the rat, *Clin. Nutr.* 25 (2006) 859–868, <https://doi.org/10.1016/j.clnu.2006.03.009>.
- [65] K. Hagiwara, T. Goto, M. Araki, H. Miyazaki, H. Hagiwara, Olive polyphenol hydroxytyrosol prevents bone loss, *Eur. J. Pharmacol.* 662 (2011) 78–84, <https://doi.org/10.1016/j.ejphar.2011.04.023>.
- [66] K.Y. Chin, S. Ima-Nirwana, Olives and Bone: A Green Osteoporosis Prevention Option, *Int. J. Environ. Res. Public Health.* 13 (2016). <https://doi.org/10.3390/ijerph13080755>.
- [67] K.Y. Chin, K.L. Pang, Therapeutic Effects of Olive and Its Derivatives on Osteoarthritis: From Bench to Bedside, *Nutrients.* 9 (2017) <https://doi.org/10.3390/nu9101060>.
- [68] V. Nicolin, N. De Tommasi, S.L. Nori, F. Costantinides, F. Berton, R. Di Lenarda, Modulatory Effects of Plant Polyphenols on Bone Remodeling: A Prospective View From the Bench to Bedside, *Front. Endocrinol. (Lausanne).* 10 (2019) 494, <https://doi.org/10.3389/fendo.2019.00494>.
- [69] E. Impellizzeri, E. Esposito, E. Mazzon, I. Paterniti, R. Di Paola, V.M. Morittu, A. Procopio, D. Britti, S. Cuzzocrea, Oleuropein aglycone, an olive oil compound, ameliorates development of arthritis caused by injection of collagen type II in mice, *J. Pharmacol. Exp. Ther.* 339 (2011) 859–869, <https://doi.org/10.1124/jpet.111.182808>.
- [70] M.N. Horcajada, C. Sanchez, F. Membrez Scalfò, P. Drion, F. Comblain, S. Taralla, A.F. Donneau, E.A. Offord, Y. Henrotin, Oleuropein or rutin consumption decreases the spontaneous development of osteoarthritis in the Hartley guinea pig, *Osteoarthritis Cartilage.* 23 (2015) 94–102, <https://doi.org/10.1016/j.joca.2014.08.016>.
- [71] Z. Feng, X. Li, J. Lin, W. Zheng, Z. Hu, J. Xuan, W. Ni, X. Pan, Oleuropein inhibits the IL-1 β -induced expression of inflammatory mediators by suppressing the activation of NF- κ B and MAPKs in human osteoarthritis chondrocytes, *Food Funct.* 8 (2017) 3737–3744, <https://doi.org/10.1039/c7fo00823f>.
- [72] S. Bulotta, M. Oliverio, D. Russo, A. Procopio, Biological Activity of Oleuropein and its Derivatives. In: *Natural Products*, Ramawat K., Mérillon JM. (eds), Chapter: 156, Publisher: Springer-Verlag Berlin Heidelberg, (2013) 3605-3638, doi 10.1007/978-3-642-22144-6_156.

- [73] A Cabarkapa, L. Zivkovic, S. Borozan, M. Zlatkovic-Svenda, D. Dekanski, I. Jancic, M. Radak-Perovic, V. Bajic, B. Spremo-Potparevic, Dry Olive Leaf Extract in Combination with Methotrexate Reduces Cell Damage in Early Rheumatoid Arthritis Patients. A Pilot Study, *Phytother. Res.* 30 (2016) 1615–1623, <https://doi.org/10.1002/ptr.5662>.
- [74] M. Cruz-Lozano, A. González-González, J.A. Marchal, E. Muñoz-Muela, M.P. Molina, F.E. Cara, A.M. Brown, G. García-Rivas, C. Hernández-Brenes, J.A. Lorente, P. Sanchez-Rovira, J.C. Chang, S. Granados-Principal, Hydroxytyrosol inhibits cancer stem cells and the metastatic capacity of triple-negative breast cancer cell lines by the simultaneous targeting of epithelial-to-mesenchymal transition, Wnt/ β -catenin and TGF β signaling pathways, *Eur. J. Nutr.* 58(8) (2019) 3207-3219, <https://doi.org/10.1007/s00394-018-1864-1>.
- [75] F. Margheri, B. Menicacci, A. Laurenzana, M. Del Rosso, G. Fibbi, M.G. Cipolleschi, J. Ruzzolini, C. Nediani, A. Mocali, L. Giovannelli, Oleuropein aglycone attenuates the pro-angiogenic phenotype of senescent fibroblasts: A functional study in endothelial cells, *J. Funct. Foods.* 53 (2019) 219–226, <https://doi.org/10.1016/j.jff.2018.12.026>.
- [76] A. Boss, K.S. Bishop, G. Marlow, M.P.G. Barnett, L.R. Ferguson, Evidence to Support the Anti-Cancer Effect of Olive Leaf Extract and Future Directions, *Nutrients.* 8(8) (2016) 513, <https://doi.org/10.3390/nu8080513>.
- [77] C. Santangelo, R. Vari, B. Scazzocchio, P. De Sanctis, C. Giovannini, M. D'Archivio, R. Masella, Anti-inflammatory Activity of Extra Virgin Olive Oil Polyphenols: Which Role in the Prevention and Treatment of Immune-Mediated Inflammatory Diseases?, *Endocr. Metab. Immune Disord. Drug Targets.* 18 (2018) 36–50, <https://doi.org/10.2174/1871530317666171114114321>.
- [78] M. Rosillo, C. Alarcón-de-la-Lastra, M.L. Castejón, T. Montoya, M. Cejudo-Guillén, M. Sánchez-Hidalgo, Polyphenolic extract from extra virgin olive oil inhibits the inflammatory response in IL-1 β -activated synovial fibroblasts, *Br. J. Nutr.* 121 (2019) 55–62, <https://doi.org/10.1017/S0007114518002829>.
- [79] I. Peluso, T. Magrone, D. Villaño Valencia, C.O. Chen, M. Palmery, Antioxidant, Anti-Inflammatory, and Microbial-Modulating Activities of Nutraceuticals and Functional Foods, *Oxid. Med. Cell Longev.* (2017) 7658617, <https://doi.org/10.1155/2017/7658617>.
- [80] K. Qabaha, F. Al-Rimawi, A. Qasem, S.A. Naser, Oleuropein Is Responsible for the Major Anti-Inflammatory Effects of Olive Leaf Extract, *J. Med. Food.* 21 (2018) 302–305, <https://doi.org/10.1089/jmf.2017.0070>.
- [81] J.M. Fernández-Real, M. Bulló, J.M. Moreno-Navarrete, W. Ricart, E. Ros, R. Estruch, J. Salas-Salvadó, A Mediterranean diet enriched with olive oil is associated with higher serum total osteocalcin levels in elderly men at high cardiovascular risk, *J. Clin. Endocrinol. Metab.* 97 (2012) 3792–3798, <https://doi.org/10.1210/jc.2012-2221>.

- [82] E. Torre, Molecular signaling mechanisms behind polyphenol-induced bone anabolism, *Phytochem. Rev.* 16 (2017), 1183–1226, <https://doi.org/10.1007/s11101-017-9529-x>.
- [83] X. Feng, J.M. McDonald, Disorders of Bone Remodeling, *Ann. Rev. Pathol.* 6 (2011) 121–145, <https://doi.org/10.1146/annurev-pathol-011110-130203>.
- [84] K.B.S. Paiva, J.M. Granjeiro, Matrix Metalloproteinases in Bone Resorption, Remodeling, and Repair, in: *Matrix Metalloproteinases and Tissue Remodeling in Health and Disease*, R.A. Khalil (Ed.), Target Tissues and Therapy, (2017) 203–303, <https://doi.org/10.1016/bs.pmbts.2017.05.001>.
- [85] M.L. Castejón, M. Rosillo, T. Montoya, A. González-Benjumea, J.G. Fernández-Bolaños, C. Alarcón-de-la-Lastra, Oleuropein down-regulated IL-1 β -induced inflammation and oxidative stress in human synovial fibroblast cell line SW982, *Food Funct.* 8 (2017), 1890–1898. <https://doi.org/10.1039/c7fo00210f>.
- [86] X. Mao, B. Xia, M. Zheng, Z. Zou, Assessment of the anti-inflammatory, analgesic and sedative effects of oleuropein from *Olea europaea* L, *Cell. Mol. Biol. (Noisy-le-grand)* 65(1) (2019)52-55.
- [87] M.M. Taskan, H. Balci Yuce, O. Karatas, F. Gevrek, H. Toker, Evaluation of the effect of oleuropein on alveolar bone loss, inflammation, and apoptosis in experimental periodontitis, *J. Periodontal Res.* 54(6) (2019) 624-632, <https://doi.org/10.1111/jre.12662>.
- [88] B. Menicacci, C. Cipriani, F. Margheri, A. Mocali, L. Giovannelli, Modulation of the Senescence-Associated Inflammatory Phenotype in Human Fibroblasts by Olive Phenols, *Int. J. Mol. Sci.* 18 (2017) <https://doi.org/10.3390/ijms18112275>.
- [89] G. Sindona, A. Caruso, A. Cozza, S. Fiorentini, B. Lorusso, E. Marini, M. Nardi, A. Procopio, S. Zicari, Anti-inflammatory effect of 3,4-DHPEA-EDA [2-(3,4 -hydroxyphenyl) ethyl (3S, 4E)-4-formyl-3-(2-oxoethyl)hex-4-enoate] on primary human vascular endothelial cells, *Curr. Med. Chem.* 19 (2012) 4006–4013, <https://doi.org/10.2174/092986712802002536>.
- [90] S. Lamy, A. Ben Saad, A. Zgheib, B. Annabi, Olive oil compounds inhibit the paracrine regulation of TNF- α -induced endothelial cell migration through reduced glioblastoma cell cyclooxygenase-2 expression, *J. Nutr. Biochem.* 27 (2016) 136–145, <https://doi.org/10.1016/j.jnutbio.2015.08.026>.
- [91] O. García-Martínez, E. De Luna-Bertos, J. Ramos-Torrecillas, C. Ruiz, E. Milia, M.L. Lorenzo, B. Jimenez, A. Sánchez-Ortiz, A. Rivas, Phenolic Compounds in Extra Virgin Olive Oil Stimulate Human Osteoblastic Cell Proliferation, *PLoS One.* 11 (2016) e0150045, <https://doi.org/10.1371/journal.pone.0150045>.
- [92] R. Santiago-Mora, A. Casado-Díaz, M.D. De Castro, J.M. Quesada-Gómez, Oleuropein enhances osteoblastogenesis and inhibits adipogenesis: the effect on differentiation in stem cells derived from bone marrow, *Osteoporosis Int.* 22 (2011) 675–684, <https://doi.org/10.1007/s00198-010-1270-x>.

- [93] R.B.H Idrus, A.B. Saim, *Olea europaea* and Its Constituents Promote Bone Health by Enhancing Osteoblast Differentiation and Proliferation: A Review, *Pharmacognosy J.* 11(1) (2019) 1–7, doi: 10.5530/pj.2019.1.1.
- [94] L. Dudaric, A. Fuzinac-Smojver, D. Muhvic, J. Giacometti, The role of polyphenols on bone metabolism in osteoporosis, *Food Res. Int.* 77 (2015) 290–298, <https://doi.org/10.1016/j.foodres.2015.10.017>.
- [95] R. Filip, S. Possemiers, A. Heyerick, I. Pinheiro, G. Raszewski, M.J. Davicco, V. Coxam, Twelve-month consumption of a polyphenol extract from olive (*Olea europaea*) in a double blind, randomized trial increases serum total osteocalcin levels and improves serum lipid profiles in postmenopausal women with osteopenia, *J. Nutr. Health Aging.* 19 (2015) 77–86, <https://doi.org/10.1007/s12603-014-0480-x>.
- [96] M. Svobodova, I. Andreadou, A.L. Skaltsounis, J. Kopecky, P. Flachs, Oleuropein as an inhibitor of peroxisome proliferator-activated receptor gamma, *Genes Nutr.* 9 (2014) 376, <https://doi.org/10.1007/s12263-013-0376-0>.
- [97] Casado-Díaz, J. Anter, S. Müller, P. Winter, J.M. Quesada-Gómez, G. Dorado, Transcriptomic analyses of the anti-adipogenic effects of oleuropein in human mesenchymal stem cells, *Food Funct.* 8 (2017) 1254–1270, <https://doi.org/10.1039/c7fo00045f>.
- [98] Z.C. Xiong, P. Luo, J. Zhou, M.S. Tan, 15-Deoxy-Delta(12,14)-prostaglandin J(2) as a potential regulator of bone metabolism via PPAR gamma-dependent and independent pathways: a review, *Drug Des. Develop. and Ther.* 13 (2019) 1879–1888, <https://doi.org/10.2147/dddt.s206695>.
- [99] J. Fornetti, A.L. Welm, S.A. Stewart, Understanding the Bone in Cancer Metastasis, *J. Bone Miner. Res.* 33 (2018) 2099–2113, <https://doi.org/10.1002/jbmr.3618>.
- [100] M. Infante, A. Fabi, F. Cognetti, S. Gorini, M. Caprio, A. Fabbri, RANKL/RANK/OPG system beyond bone remodeling: involvement in breast cancer and clinical perspectives, *J. Exp. Clin. Cancer Res.* 38 (2019) 12, <https://doi.org/10.1186/s13046-018-1001-2>.
- [101] H. Roca, L.K. McCauley, Inflammation and skeletal metastasis, *Bonekey Rep.* 4 (2015) <https://doi.org/10.1038/bonekey.2015.75>.
- [102] K.L. Owen, B.S. Parker, Beyond the vicious cycle: The role of innate automimicry and tumor-inherent changes in dictating bone metastasis, *Mol. Immunol.* 110 (2019) 57-68, doi: 10.1016/j.molimm.2017.11.023. Epub 2017 Nov 27.
- [103] B. Ritter, F.R. Greten, Modulating inflammation for cancer therapy, *J. Exp. Med.* 216 (2019) 1234–1243, <https://doi.org/10.1084/jem.20181739>.

- [104] K. Maeda, Y. Kobayashi, M. Koide, S. Uehara, M. Okamoto, A. Ishihara, T. Kayama, M. Saito, K. Marumo, The Regulation of Bone Metabolism and Disorders by Wnt Signaling, *Int. J. Mol. Sci.* 20(22) (2019) 5525, doi:10.3390/ijms20225525.
- [105] L. Zhang, F. Zhou, P. ten Dijke, Signaling interplay between transforming growth factor- β receptor and PI3K/AKT pathways in cancer, *Trends Biochem Sci.* 38 (12) (2013) 612–620, doi:10.1016/j.tibs.2013.10.001.
- [106] J. Zhang, X.H. Yu, Y.G. Yan, C. Wang, W. J. Wang, PI3K/Akt signaling in osteosarcoma, *Clin. Chim. Acta.* 444 (2015)182-92, doi: 10.1016/j.cca.2014.12.041. Epub 2015 Feb 19.
- [107] A.Vallée, Y. Lecarpentier, J.-N. Vallée, Targeting the Canonical WNT/ β -Catenin Pathway in Cancer Treatment Using Non-Steroidal Anti-Inflammatory Drugs, *Cells.* 8(7) (2019) 726, doi:10.3390/cells8070726.
- [108] Y.P. Wu, W.S. Chen, S.J. Xu, N. Zhang, Osteoporosis as a potential contributor to the bone metastases, *Medical Hypotheses.* 75 (2010) 514–516, <https://doi.org/10.1016/j.mehy.2010.07.010>.
- [109] F. Salamanna, V. Borsari, S. Brogini, P. Torricelli, S. Cepollaro, M. Cadossi, M. Fini, A Human 3D In Vitro Model to Assess the Relationship Between Osteoporosis and Dissemination to Bone of Breast Cancer Tumor Cells, *J. Cell. Physiol.* 232 (2017) 1826–1834, <https://doi.org/10.1002/jcp.25708>.
- [110] D. Bellavia, F. Salamanna, L. Raimondi, A. De Luca, V. Carina, V. Costa, R. Alessandro, M. Fini, G. Giavaresi, Deregulated miRNAs in osteoporosis: effects in bone metastasis, *Cell. Mol. Life Sci.* 76 (2019) 3723–3744, <https://doi.org/10.1007/s00018-019-03162-w>.
- [111] L. Das Roy, J.M. Curry, M. Sahraei, D.M. Besmer, A. Kidiyoor, H.E. Gruber, P. Mukherjee, Arthritis augments breast cancer metastasis: role of mast cells and SCF/c-Kit signaling, *Breast Cancer Res.* 15 (2013) R32. <https://doi.org/10.1186/bcr3412>.
- [112] H.M. Chen, F.P. Chen, K.C. Yang, S.S. Yuan, Association of Bone Metastasis With Early-Stage Breast Cancer in Women With and Without Precancer Osteoporosis According to Osteoporosis Therapy Status, *JAMA Netw Open.* 2 (2019) e190429, <https://doi.org/10.1001/jamanetworkopen.2019.0429>.
- [113] A. Muhammad, S.B. Mada, I. Malami, G.E. Forcados, O.L. Erukainure, H. Sani, I.B. Abubakar, Postmenopausal osteoporosis and breast cancer: The biochemical links and beneficial effects of functional foods, *Biomed. Pharmacother.* 107 (2018) 571–582, <https://doi.org/10.1016/j.biopha.2018.08.018>.
- [114] O. Leal-Hernández, R. Roncero-Martín, I. Aliaga, M. Pedrera-Canal, J.M. Morán, S. Rico, L.M. Puerto Parejo and J.M. Lavado-García, P-53-Independent Apoptosis of Human Osteosarcoma Cells After Exposure to Oleuropein, *Lat. Am. J. Pharm.* 39 (2020) 668-672.

- [115] M.H. Elamin, M.H. Daghestani, S.A. Omer, M.A. Elobeid, P. Virk, E.M. Al-Olayan, Z.K. Hassan, O.B. Mohammed, A. Aboussekhra, Olive oil oleuropein has anti-breast cancer properties with higher efficiency on ER-negative cells, *Food Chem. Toxicol.* 53 (2013) 310–316, <https://doi.org/10.1016/j.fct.2012.12.009>.
- [116] A. Papachristodoulou, M. Tsoukala, D. Benaki, S. Kostidis, K. Gioti, N. Aligiannis, H. Pratsinis, D. Kletsas, A.-L. Skaltsounis, E. Mikros, R. Tenta, Oleuropein is a Powerful Sensitizer of Doxorubicin-mediated Killing of Prostate Cancer Cells and Exerts Its Action via Induction of Autophagy, *J. Cancer Res. Treat.* 4 (2018) 61–68, <https://doi.org/10.12691/jcrt-4-4-2>.
- [117] A.M. Nassir, I.A.A. Ibrahim, S. Md, M. Waris, T. MR Ain, I. Ahmad, N. Shahzad, Surface functionalized folate targeted oleuropein nano-liposomes for prostate tumor targeting: Invitro and invivo activity, *Life Sci. Mar* 1;220 (2019) 136-146, doi: 10.1016/j.lfs.2019.01.053.
- [118] W. Wang, J. Wu, Q. Zhang, X. Li, X. Zhu, Q. Wang, S. Cao, L. Du, Mitochondria-mediated apoptosis was induced by oleuropein in H1299 cells involving activation of p38 MAP kinase, *J. Cell Biochem.* 120 (2019) 5480–5494, <https://doi.org/10.1002/jcb.27827>.
- [119] C. Antognelli, R. Frosini, M.F. Santolla, M.J Peirce, V.N. Talesa, Oleuropein-Induced Apoptosis Is Mediated by Mitochondrial Glyoxalase 2 in NSCLC A549 Cells: A Mechanistic Inside and a Possible Novel Nonenzymatic Role for an Ancient Enzyme, *Oxidative Med. Cell. Longev.* 2019 (2019) 8576961, <https://doi.org/10.1155/2019/8576961>.
- [120] W. Mao, H. Shi, X. Chen, Y. Yin, T. Yang, M. Ge, M. Luo, D. Chen and X. QiAN, Anti-Proliferation and Migration Effects of Oleuropein on Human A549 Lung Carcinoma Cells, *Lat. Am. J. Pharm.* 31 (8) (2012) 1217-21.
- [121] Z. Bouallagui, J Han, Isoda, S. Sayadi, Hydroxytyrosol rich extract from olive leaves modulates cell cycle progression in MCF-7 human breast cancer cells, *Food Chem. Toxicol.* 49(2011)179–184, doi:10.1016/j.fct.2010.10.014.
- [122] J. Han, T.P.N. Talorete, P. Yamada, H. Isoda, Anti-proliferative and apoptotic effects of oleuropein and hydroxytyrosol on human breast cancer MCF-7 cells, *Cytotechnology.* 59 (2009) 45–53, doi: 10.1007/s10616-009-9191-2.
- [123] A. Chimento, I. Casaburi, C. Rosano, P. Avena, A. De Luca, C. Campana, E. Martire, M.F. Santolla, M. Maggiolini, V. Pezzi, Oleuropein and hydroxytyrosol activate GPER/GPR30-dependent pathways leading to apoptosis of ER-negative SKBR3 breast cancer cells, *Mol. Nutr. Food Res.* 58 (2014) 478–489, <https://doi.org/10.1002/mnfr.201300323>.

- [124] R. Sirianni, A. Chimento, A. De Luca, I. Casaburi, P. Rizza, A. Onofrio, D. Iacopetta, F. Puoci, S. Andò, M. Maggiolini and V. Pezzi, Oleuropein and hydroxytyrosol inhibit MCF-7 breast cancer cell proliferation interfering with ERK1/2 activation, *Mol. Nutr. Food Res.* 54 (2010) 833–840, DOI: 10.1002/mnfr.200900111.
- [125] S. Cao, X. Zhu, L. Du, P38 MAP kinase is involved in oleuropein-induced apoptosis in A549 cells by a mitochondrial apoptotic cascade, *Biomed. Pharmacother.* 95 (2017) 1425-1435, doi: 10.1016/j.biopha.2017.09.072.
- [126] Z.K. Hassan, M.H. Elamin, S.A. Omer, M.H. Daghestani, E.S. Al-Olayan, M.A. Elobeid, P. Virk, Oleuropein induces apoptosis via the p53 pathway in breast cancer cells. *Asian Pac. J. Cancer Prev.* 14(11) (2014) 6739-6742, doi: 10.7314/apjcp.2013.14.11.6739. PMID: 24377598.
- [127] S. Asgharzade, S. H. Sheikhshabani, E. Ghasempour, R. Heidari, S. Rahmati, M. Mohammadi, A. Jazaeri, Z. Amini-Farsani, The effect of oleuropein on apoptotic pathway regulators in breast cancer cells, *Eur J Pharmacol.* 2020;886:173509. doi:10.1016/j.ejphar.2020.173509.
- [128] B. Kaltschmidt, C. Banz-Jansen, T. Benhidjeb T, M. Beshay, C. Förster, J. Greiner, E. Hamelmann, N. Jorch, F. Mertzlufft, J. Pfitzenmaier, M. Simon, J. Schulte Am Esch, T. Vordemvenne, D. Wähnert, F. Weissinger, L. Wilkens, B. Kaltschmidt, A Role for NF- κ B in Organ Specific Cancer and Cancer Stem Cells. *Cancers (Basel)*. 11(5) (2019) 655, doi:10.3390/cancers11050655.
- [129] C.J. Li, P.Y. Chu, G.T. Yiang, M.Y. Wu, The Molecular Mechanism of Epithelial-Mesenchymal Transition for Breast Carcinogenesis, *Biomolecules*. 9(9) (2019) 476, doi:10.3390/biom9090476.
- [130] B. Demirkan, The Roles of Epithelial-to-Mesenchymal Transition (EMT) and Mesenchymal-to-Epithelial Transition (MET) in Breast Cancer Bone Metastasis: Potential Targets for Prevention and Treatment, *J. Clin. Med.* 2(4) (2013) 264-282, doi: 10.3390/jcm2040264.
- [131] A.H. Nwabo Kamdje, P. Takam Kamga, R. Tagne Simo, L. Vecchio, P.F. Seke Etet, J.M. Muller, G. Bassi, E. Lukong, R. Kumar Goel, J. Mbo Amvene, M. Krampera, Developmental pathways associated with cancer metastasis: Notch, Wnt, and Hedgehog, *Cancer Biol. Med.* 14 (2017) 109-120, doi: 10.20892/j.issn.2095-3941.2016.0032.
- [132] P. Juárez, T.A. Guise, TGF- β in cancer and bone: implications for treatment of bone metastases. *Bone*. 48(1) (2011) 23–29, doi:10.1016/j.bone.2010.08.004.
- [133] Y. Hao, D. Baker, P. Ten Dijke, TGF- β -Mediated Epithelial-Mesenchymal Transition and Cancer Metastasis, *Int. J. Mol. Sci.* 20(11)(2019) 2767, doi:10.3390/ijms20112767.

- [134] F. Verrecchia, F. R dini, Transforming Growth Factor- β Signaling Plays a Pivotal Role in the Interplay Between Osteosarcoma Cells and Their Microenvironment. *Front Oncol.* (8) (2018) 133, doi:10.3389/fonc.2018.0013.
- [135] H. Tsai and J. Yang, Epithelial-mesenchymal plasticity in carcinoma metastasis, *Genes Dev.* 27(20) (2013) 2192–2206, doi: 10.1101/gad.225334.113.
- [136] C. Chandhanayingyong, Y. Kim, J.R. Staples, C. Hahn, F.Y. Lee, MAPK/ERK Signaling in Osteosarcomas, Ewing Sarcomas and Chondrosarcomas: Therapeutic Implications and Future Directions. *Sarcoma.* 2012, 404810. doi:10.1155/2012/404810.
- [137] S.R. Lin, N. Mokgautsi, Y.N. Liu, Ras and Wnt Interaction Contribute in Prostate Cancer Bone Metastasis, *Molecules.* 25(10)(2020) 2380, doi: 10.3390/molecules25102380.
- [138] G. Danieau, S. Morice, F. R dini, F. Verrecchia, B.B. Royer, New Insights about the Wnt/ β -Catenin Signaling Pathway in Primary Bone Tumors and Their Microenvironment: A Promising Target to Develop Therapeutic Strategies? *Int. J. Mol. Sci.* 20(15) (2019)3751, doi:10.3390/ijms20153751.
- [139] B.K. Park, H. Zhang, Q. Zeng, J. Dai, E.T. Keller, T. Giordano, K. Gu, V. Shah, L. Pei, R.J. Zarbo RJ, L. McCauley, S. Shi, S. Chen, C.Y. Wang, NF-kappa B in breast cancer cells promotes osteolytic bone metastasis by inducing osteoclastogenesis via GM-CSF, *Nat. Med.* 13 (2007) 62–69, <https://doi.org/10.1038/nm1519>.
- [140] R. Jin, J.A. Sterling, J.R. Edwards, D.J. eGraff, C. Lee, S.I. Park, R.J. Matusik, Activation of NF-kappa B signaling promotes growth of prostate cancer cells in bone, *PLoS One.* 8(4) (2013) e60983, <https://doi.org/10.1371/journal.pone.0060983>.
- [141] R.W. Johnson, M.E. Sowder, A.J. Giaccia, Hypoxia and Bone Metastatic Disease, *Curr. Osteoporos. Rep.* 15(4) (2017) 231–238, doi:10.1007/s11914-017-0378-8.
- [142] F.N. Soki, S.I. Park, L.K. McCauley, The multifaceted actions of PTHrP in skeletal metastasis, *Future Oncol.* 8(7) (2012) 803–817, doi:10.2217/fon.12.76.
- [143] C. Nediani, J. Ruzzolini, A. Romani, L. Calorini, Oleuropein, a Bioactive Compound from *Olea europaea* L, as a Potential Preventive and Therapeutic Agent in Non-Communicable Diseases, *Antioxidants (Basel).* 8(12) (2019) 578, doi:10.3390/antiox8120578.
- [144] V. Goulas V. Exarchou, A.N. Troganis, E. Psomiadou, T. Fotsis, E. Briasoulis and I.P. Gerothanassis, Phytochemicals in olive-leaf extracts and their antiproliferative activity against cancer and endothelial cells, *Mol. Nutr. Food Res.* 53(5) (2009) 600-608, doi:10.1002/mnfr.200800204.

- [145] X. Lu, E. Mu, Y. Wei, S. Riethdorf, Q. Yang, M. Yuan, J. Yan, Y. Hua, B.J. Tiede, X. Lu, B.G. Haffty, K. Pantel, J. Massagué, Y. Kang, VCAM-1 promotes osteolytic expansion of indolent bone micrometastasis of breast cancer by engaging $\alpha 4\beta 1$ -positive osteoclast progenitors, *Cancer Cell*. 20(6) (2011) 701–714, doi:10.1016/j.ccr.2011.11.002.
- [146] M. Dell’Agli, R. Fagnani, N. Mitro, S. Scurati, M. Masciadri, L. Mussoni, G.V. Galli, E. Bosisio, M. Crestani, E. De Fabiani, E. Tremoli, D. Caruso, Minor Components of Olive Oil Modulate Proatherogenic Adhesion Molecules Involved in Endothelial Activation, *J. Agric. Food Chem.* 54 (2006) 3259–3264. doi: 10.1021/jf0529161.
- [147] H. Choi, A. Moon, Crosstalk between cancer cells and endothelial cells: implications for tumor progression and intervention, *Arch. Pharm. Res.* 41 (2018) 711–724, <https://doi.org/10.1007/s12272-018-1051-1>.
- [148] N. Yahfoufi, N. Alsadi, M. Jambi, C. Matar, The Immunomodulatory and Anti-Inflammatory Role of Polyphenols, *Nutrients*. 10 (11) (2018) 1618, <https://doi.org/10.3390/nu10111618>.
- [149] M. Paolillo, S. Schinelli, Extracellular Matrix Alterations in Metastatic Processes, *Int. J. Mol. Sci.* 20(19) (2019) 4947, doi:10.3390/ijms20194947.
- [150] H. Harjunpää, M. Lloret Asens, C. Guenther, S.C. Fagerholm, Cell Adhesion Molecules and Their Roles and Regulation in the Immune and Tumor Microenvironment, *Front. Immunol.* 10 (2019) 1078, doi:10.3389/fimmu.2019.01078.
- [151] N. Hashemi Goradel, M. Najafi, E. Salehi, B. Farhood, K. Mortezaee, Cyclooxygenase- 2 in cancer: A review, *J. Cell. Physiol.* 234 (2019) 5683– 5699, <https://doi.org/10.1002/jcp.27411>.
- [152] E. Hardy, C. Fernandez-Patron, Destroy to Rebuild: The Connection Between Bone Tissue Remodeling and Matrix Metalloproteinases, *Front. Physiol.* 2020;11:47, doi: 10.3389/fphys.2020.00047. eCollection 2020.
- [153] A. Dhawan, J. Friedrichs, M.V. Bonin, E.P. Bejestani, C. Werner, M. Wobus, T. Chavakis, M. Bornhäuser, Breast cancer cells compete with hematopoietic stem and progenitor cells for intercellular adhesion molecule 1-mediated binding to the bone marrow microenvironment, *Carcinogenesis*. 37(8) (2016) 759-767, doi: 10.1093/carcin/bgw057. Epub 2016 May 4.
- [154] M. Esposito, N. Mondal, T.M. Greco, Y. We, C. Spadazzi, S.C. Lin, H. Zheng, C. Cheung, J.L. Magnani, S.H. Lin, I.M. Cristea, R. Sackstein, Y. Kang, Bone vascular niche E-selectin induces mesenchymal-epithelial transition and Wnt activation in cancer cells to promote bone metastasis, *Nat. Cell Biol.* 21(5) (2019) 627–639, doi:10.1038/s41556-019-0309-2.

- [155] P. Przychodzen, A. Kuban-Jankowska, R. Wyszowska, G. Barone, G. Lo Bosco, F. Lo Celso, A. Kamm, A. Daca, T. Kostrzewa, M. Gorska-Ponikowska, PTP1B phosphatase as a novel target of oleuropein activity in MCF-7 breast cancer model, *Toxicol. In Vitro*. 61(2019) 104624, doi:10.1016/j.tiv.2019.104624.
- [156] B. Hilmarsson, E. Briem, S. Halldorsson, J. Krickler, S. Ingthorsson, S. Gustafsdottir, G.M. Mælandsmo, M.K. Magnusson, T. Gudjonsson, Inhibition of PTP1B disrupts cell–cell adhesion and induces anoikis in breast epithelial cells, *Cell Death Dis.* 8(5) (2017) e2769, <https://doi.org/10.1038/cddis.2017.17>.
- [157] G. Ren, M. Esposito, Y. Kang, Bone metastasis and the metastatic niche, *J. Mol. Med (Berl)*. 93(11) (2015) 1203–1212, doi:10.1007/s00109-015-1329-4.
- [158] Y. Liu, X. Cao, Characteristics and Significance of the Pre-metastatic Niche, *Cancer Cell*. 30(5) (2016) 668-681. doi: 10.1016/j.ccell.2016.09.011.
- [159] N.M. Byrne, M.A. Summers, M.M. McDonald, Tumor Cell Dormancy and Reactivation in Bone: Skeletal Biology and Therapeutic Opportunities, *JBMR Plus*. 3(3) (2019) e10125, doi:10.1002/jbm4.10125.
- [160] M.A. Rosillo, S. Montserrat-de-la-Paz, R. Abia, M. L. Castejon, M. C. Millan-Linares, C. Alarcon-de-la-Lastra, J.G. Fernandez-Bolaños, and F. Muriana, Oleuropein and its peracetylated derivative negatively regulate osteoclastogenesis by controlling the expression of genes involved in osteoclast differentiation and function, *Food Funct*. 11(5)(2020) 4038- 4048, doi:10.1039/d0fo00433b.
- [161] S. Kany, J.T. Vollrath, B. Relja, Cytokines in Inflammatory Disease, *Int. J. Mol. Sci.* 20(23) (2019) 6008, doi:10.3390/ijms20236008.
- [162] F. Salamanna, V. Borsari, D. Contartese, V. Costa, G. Giavaresi, M. Fini, What Is the Role of Interleukins in Breast Cancer Bone Metastases? A Systematic Review of Preclinical and Clinical Evidence. *Cancers(Basel)*. 11(12) (2018) 2019, doi:10.3390/cancers11122018.
- [163] A.C. Monteiro, A.C. Leal, T. Gonçalves-Silva, A.T.C. Mercadante, F. Kestelman, S.B. Chaves, R. J. P. Monteiro, A. Bonomo, T cells induce pre-metastatic osteolytic disease and help bone metastases establishment in a mouse model of metastatic breast cancer, *PLoS One*. 8(7) (2013) e68171, doi:10.1371/journal.pone.0068171.
- [164] H. Khan, H. Ullah, P.C.M.F. Castilho, A.S. Gomila, G. D’Onofrio, R. Filosa, F. Wang, S.M. Nabavi, M. Daglia, A.S. Silva, K.R.R. Rengasamy, J. Ou, X. Zou, J. Xiao, H. Cao, Targeting NF- κ B signaling pathway in cancer by dietary polyphenols, *Crit. Rev. Food Sci. Nutr.* (2019) 1–11. <https://doi.org/10.1080/10408398.2019.1661827>.
- [165] D. Verzella, M. Fischietti, D. Capece, D. Vecchiotti, F. Del Vecchio, G. Cicciarelli, V. Mastroiaco, A. Tessitore, E. Alesse, F. Zazzeroni, Targeting the NF- κ B pathway in prostate cancer: a promising therapeutic approach? *Curr. Drug Targets*. 17(3) (2016) 311- 320, doi:10.2174/1389450116666150907100715.

- [166] X. Wu, F. Li, L. Dang, C. Liang, A. Lu, G. Zhang, RANKL/RANK System-Based Mechanism for Breast Cancer Bone Metastasis and Related Therapeutic Strategies, *Front. Cell Dev. Biol.* 8 (2020) 76, doi: 10.3389/fcell.2020.00076. eCollection 2020.
- [167] A. Suvannasankha, J.M. Chirgwin, Role of bone-anabolic agents in the treatment of breast cancer bone metastases, *Breast Cancer Res* 16 (2014) 484, <https://doi.org/10.1186/s13058-014-0484-9>.
- [168] J. Adamik, D.L. Galson, G.D. Roodman, Osteoblast suppression in multiple myeloma bone disease, *J Bone Oncol.* 13 (2018) 62-70, doi:10.1016/j.jbo.2018.09.001.
- [169] Y. Xing, D. Cui, S. Wang, P. Wang, X. Xing, H. Li, Oleuropein represses the radiation resistance of ovarian cancer by inhibiting hypoxia and microRNA-299-targetted heparanase expression, *Food Funct.* 8(8) (2017) 2857–2864, doi: 10.1039/c7fo00552k.
- [170] J.A. Menendez, A. Vazquez-Martin, R. Colomer, J. Brunet, A. Carrasco-Pancorbo, R. Garcia-Villalba, A. Fernandez-Gutierrez and A. Segura-Carretero, Olive oil's bitter principle reverses acquired autoresistance to trastuzumab (Herceptin™) in HER2-overexpressing breast cancer cells, *BMC Cancer* 7 (2007) 80, <https://doi.org/10.1186/1471-2407-7-80>.
- [171] M. de Bock, E.B. Thorstensen, J.G. Derraik, H.V. Henderson, P.L. Hofman, W.S. Cutfield, Human absorption and metabolism of oleuropein and hydroxytyrosol ingested as olive (*Olea europaea* L.) leaf extract, *Mol. Nutr. Food Res.* 57(11) (2013) 2079- 2085, doi:10.1002/mnfr.201200795.
- [172] L. Rubi , A. Maci , M. J. Motilva, Impact of various factors on pharmacokinetics of bioactive polyphenols: an overview, *Curr Drug Metab.* 15 (2014) 62-76. doi:10.2174/1389200214666131210144115.
- [173] M.C.L. De las Hazas, C. Pi ol, A. Maci , M.-P. Romero, A. Pedret, R. Sol , L. Rubi , M.-J. Motilva, Differential absorption and metabolism of hydroxytyrosol and its precursors oleuropein and secoiridoids, *J. Funct. Food.* 22 (2016) 52–63, <https://doi.org/10.1016/j.jff.2016.01.030>.
- [174] F. Visioli, A. Davalos, M- C. L pez de las Hazas, M.C. Crespo, J. Tom - Carneiro, An overview of the pharmacology of olive oil and its active ingredients, *Br J. Pharmacol.* 177 (2020) 1316–1330. <https://doi.org/10.1111/bph.14782>.
- [175] D. Russo, D. Britti, S.M. Lepore, F. Trimboli, M. Celano, M. Oliverio, R.F. Rose, G. Loprete, N. Costa, C.D. Loreto, G. Damante, S. Bonacci, A. Procopio, Safety Evaluation of Subchronic Oral Administration of Oleuropein and its Semisynthetic Peracetylated Derivative, *J. Pharm. Pharmaceutics* 3(1) (2016) 40- 45, <https://doi.org/10.15436/2377-1313.16.014>.

- [176] A. Karković Marković, J. Torić, M. Barbarić, C. Jakobušić Brala, Hydroxytyrosol, Tyrosol and Derivatives and Their Potential Effects on Human Health, *Molecules*. 2019;24(10):2001. doi:10.3390/molecules24102001.
- [177] S. Bulotta, R. Corradino, M. Celano, J. Maiuolo, M. D'Agostino, M. Oliverio, A. Procopio, S. Filetti and D. Russo, Antioxidant and antigrowth action of peracetylated oleuropein in thyroid cancer cells, *J. Mol. Endocrinol.* 51(1) (2013)181-189. doi: 10.1530/JME-12-0241
- [178] G. Juli, M. Oliverio, D. Bellizzi, M.E. Gallo Cantafio, K. Grillone, G. Passarino, C. Colica, M. Nardi, M. Rossi, A. Procopio, P. Tagliaferri, P. Tassone, N. Amodio N, Anti-tumor Activity and Epigenetic Impact of the Polyphenol Oleacein in Multiple Myeloma, *Cancers (Basel)*. 11(7) (2019) 990, doi: 10.3390/cancers11070990.
- [179] G. Chindamo, S. Sapino, E. Peira, D. Chirio, M.C. Gonzalez, M.Gallarate, Bone Diseases: Current Approach and Future Perspectives in Drug Delivery Systems for Bone Targeted Therapeutics, *Nanomaterials (Basel)*. 10(5) (2020) 875, doi:10.3390/nano10050875.
- [180] F. Yang, Z. Zhao, B. Sun, Q. Chen, J. Sun, Z. He, C. Luo, Nanotherapeutics for Antimetastatic Treatment *Trends Cancer*. 6(8) (2020) 645-659, doi: <https://doi.org/10.1016/j.trecan.2020.05.001>.
- [181] I.M. Adjei, M.N. Temples, S.B. Brown, B. Sharma, Targeted Nanomedicine to Treat Bone Metastasis. *Pharmaceutics*. 10(4) (2018) 205, doi:10.3390/pharmaceutics10040205.
- [182] W. Gu, C. Wu, J. Chen, Y. Xiao, Nanotechnology in the targeted drug delivery for bone diseases and bone regeneration, *Int. J. Nanomedicine*. 8 (2013) 2305-2317, doi:10.2147/IJN.S44393.
- [183] S. Palagati, S. Sv and B.R. Kesavan, Application of computational tools for the designing of Oleuropein loaded nanostructured lipid carrier for brain targeting through nasal route, *Daru J. Pharm. Sci.* 27 (2019) 695 -708, doi:10.1007/s40199-019-00304-0.
- [184] C. Bonechi, A. Donati, G. Tamasi, A. Pardini, H. Rostom, G. Leone, S. Lamponi, M. Consumi, A. Magnani and C. Ross, Chemical characterization of liposomes containing nutraceutical compounds: Tyrosol, hydroxytyrosol and oleuropein, *Biophys. Chem.* 246 (2019) 25-34, <https://doi.org/10.1016/j.bpc.2019.01.002>.
- [185] S. Tabrez, N.R. Jabir, V.M. Adhami, M.I. Khan, M. Moulay, M.A. Kamal, H. Mukhtar, Nanoencapsulated dietary polyphenols for cancer prevention and treatment: successes and challenges, *Nanomedicine* 15, 1(2020)147–1162, doi: 10.2217/nmm-2019-0398.

Figure 1. Chemical structure of Oleuropein and its hydrolysis products

Figure 2. Pleiotropic effects of Oleuropein on cellular and molecular components involved in pre-metastatic niche formation.

OLE may inhibit (⚡) the expression and activity of growth factors, hormones, proinflammatory cytokines, adhesion molecules and enzymes, which regulate different cellular processes involved in cancer progression, including *i*) apoptosis, *ii*) cell cycle progression, *iii*) cell differentiation and *iv*) epithelial-to-mesenchymal transition (EMT). Most of these signalling molecules have been shown to be deregulated in several pathological conditions associated with bone loss and to contribute to the formation of the “*bone pre-metastatic niche*”. On the other hand, OLE has been shown to exert osteo-protective effects by fostering (⇒) the differentiation of human bone marrow mesenchymal stem cells into osteoblasts and by inhibiting (⚡) osteoclastogenesis.

Figure 3 Schematic representation of the effects of Oleuropein on molecular signalling pathways linking chronic inflammation and cancer progression

OLE may modulate the expression and activity of various signalling molecules, such as *1*) cytokines, chemokines [20,21,55,69,71,73,75], *2*) transcription factors [18,20,38,71,75-80], and *3*) enzymes [20,43,44,71,75,77] which, in physiological conditions, regulate several biological functions including the normal metabolic turnover of bone tissue while, their expression proves to be deregulated in various pathological conditions associated with altered bone remodelling such as chronic inflammatory processes and malignant bone disorders [7,20,47,68,71,75,83, 99-103,106].

Table 1In vivo effects of **OLE** on some human tumors which metastasize to the bone

Tumor model	Treatment	Effects	Mechanisms	References
Nu/nu athymic ovariectomised mice MCF7 human breast cancer	OLE enriched diet (125mg OLE /kg of diet up to 35 days)	tumor growth lung metastasis	▼ cell proliferation ▼ cell migration cell invasion	[35]
BALB/c OlaHsd-foxn1 nude mice MDA-MB-231 human breast cancer	OLE 50 mg/kg i.p. once a week for 4 weeks	tumor growth	▼ NF-kB Bcl-2 Cyclin D1	[34]
<u>BALB/c nude mice</u> 22Rv1 Human <u>prostate cancer</u>	OLE , 25 mg/kg/ every24h/30 days	tumor growth apoptosis	▼ cell proliferation	[117]
Swiss albino mice Soft tissue sarcoma	1% OLE in drinking water, consumed ad libitum for 12 days	tumor growth	▼ cell proliferation cell migration cell invasion	[32]
C57BL/6N mice BF16F10 melanoma	High Fat Diet + OLE 0.02% or 0.04%	tumor growth lymph-node metastasis	▼ Angiogenesis ▼ Lymphangiogenesis	[36]

▼, **inhibition**

Table 2.

In vitro effects of **OLe** on the growth and survival of tumor cells from human neoplasms which preferentially metastasize to the bone

Tumor	Cell line	Effects		Mechanisms		References
Breast Cancer	MCF-7	cell proliferation apoptosis	▼ ▲	DNA damage PARP levels	▲	[33]
	MCF-7	cell proliferation apoptosis	▼ ▲	block of G ₁ to S phase transition		[121,122]
	MCF-7 MDA-MB-231	cell proliferation apoptosis	▼ ▲	cell cycle arrest in the G2M cell cycle delaying the at S-phase cyclin D1 Erk1/2 Bax Bcl-2	▼ ▼ ▲ ▲	[34,115,124]
	MCF-7 MDA-MB-231	apoptosis	▲	NF-kB	▼	[34,37,115]
	MCF-7	apoptosis	▲	P53, Bax Bcl-2	▲ ▼	[126]
	SKBR3 (ER-)	apoptosis	▲	Activation GPER/GPR30 dependent pathway	▲	[123]
	MCF7	apoptosis	▲	HDAC2 , HDAC3	▼	[50]
	MCF7	cell proliferation	▼	PTP1B	▼	[155]
	MCF-7 MDA-MD-231	cell cycle apoptosis	▼ ▲	miR-125b, miR-16, miR-34a, p53, p21, and TNFRS10B	▲	[42]
				bcl-2, mcl1, miR- 21 and miR- 155 miR-221, miR-29a and miR-21	▼	[127]
Prostate cancer	LNCaP, DU145 and BPH-1	apoptosis autophagy	▲ ▲	pAkt, γ-GCS, HO-1 pro-oxidant activity in cancer cells but not in normal cells	▼ ▼ ▼ ▲	[26]

	22Rv1	apoptosis	▲	mitochondrial membrane potential caspase-3	▼ ▲	[117]
	Highly metastatic Dunning rat prostate cancer	cell motility	▼	blocking voltage-gated sodium channels (VGSCs).	▲	[41]
Thyroid cancer	TPC-1 and BCPAP	cell proliferation	▼	block in S phase AKT and ERK phosphorylation H ₂ O ₂ -induced ROS levels ROS levels	▼ ▼ ▼	[176]
Lung Cancer	A549 NSLC	apoptosis	▲	<u>phosphorylation</u> of p38MAPK protein mitochondrial membrane potential mitochondrial cytochrome C release Bax/Bcl2 ratio caspase 9 and caspase 3 SOD2 upregulation Akt signaling pathway mitochondrial Glo2 expression	▲ ▼ ▲ ▲ ▲ ▲ ▼ ▼	[110,111,125]
	H1299 lung cancer	apoptosis	▲	Activation of p38MAPK		[109]
Osteosarcoma	143B OS, Saos2, p53-null MG-63	cell proliferation apoptosis autophagy	▼ ▲ ▲	Bax/Bcl-2, ratio Caspase -3	▲ ▲	[38,63, 114]

▼ inhibition; ▲ promotion

Table 3

Effects of **OLe** on the key events and signalling pathways underlying the dissemination of cancer cells in the bone microenvironment

Event	Mechanisms	Effects	References	
EMT	TGF- β / MAP/ERK	▼	[54,74,132-134], [55,120,125,134-136]	
	PI3K/AKT	▼	[54,105,137]	
	Wnt/ β -catenin, NF- κ B	▼	[54-56,74,131,138-140] [127,133,138,140,141]	
	HIF and PTHrP	▼	[36,142] [143]	
	Migration /Adhesion	EMT	▼	[37]
		PTPB1	▼	[155,156]
Invasion	MMPs	▼	[23,31,45,46,55,71,132,143,152]	
	AKT	▼		
Angiogenesis	VEGF, VEGFR-2	▼	[33, 90]	
	HIF-1 α , VEGF-A VEGF-D	▼	[36]	
	NF- κ B, IL-6, IL-8, IL-1 β TNF- α	▼	[20,39,79,131,132, 143,147] [19,20,22,48,80,90,143-146]	
	VCAM-1, ICAM-1, E-selectin	▼	[20,22,39,52,71,75,145,146]	
	COX-2	▼	[31,39,45,71,75,98]	
	MMPs	▼	[31,37,45,46,51,71,75,118,130]	
	Bone micro-environment (pre-metastatic niche)	IL-6, IL-8, IL-1 β , TNF- α	▼	[19,20,90,143,147,160-164].
EMT transition		▼	[37,53-56,130-147]	
Angiogenesis		▼	[20, 36, 75,143,148]	
MMP2, MMP9		▼	[45,90,130-134, 147,152]	
TIMP-1, TIM,-3 and TIMP-4		▲	[46,51,71,75,131,152]	
NF- κ B		▼	[20,39,51,55,71,148,149,165]	
Osteoclast		▼	[20, 64-68, 89,91,143,160]	
Osteoblast		▲	[20, 64-68, 91-93,143]	
HSC		▼	[20,92 97,143]	
MSC		▲	[20, 92,96,143]	
RANKL-induced NF- κ B activation		▼	[68,70,93,94,161,166].	
RANKL/RANK/OPG system		▼/▲	[100,149,153,157,165,166].	
PPAR γ		▼	[91,96-98]	

▼ inhibition ; ▲ promotion

Fig.1

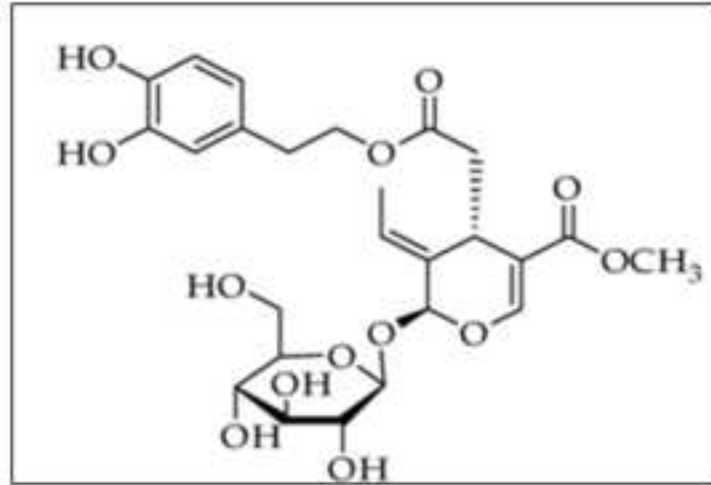
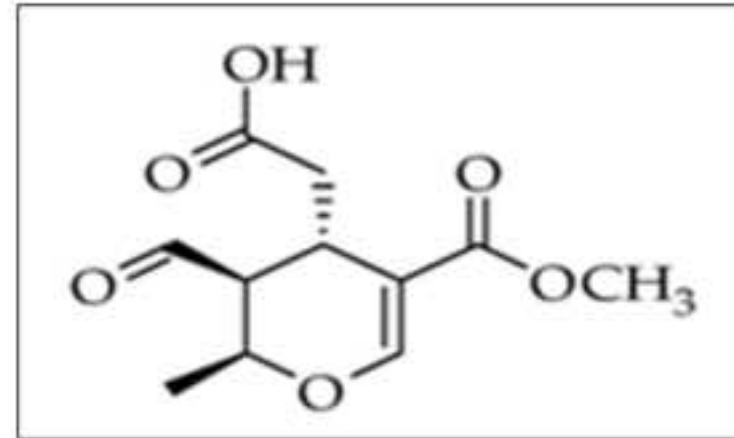
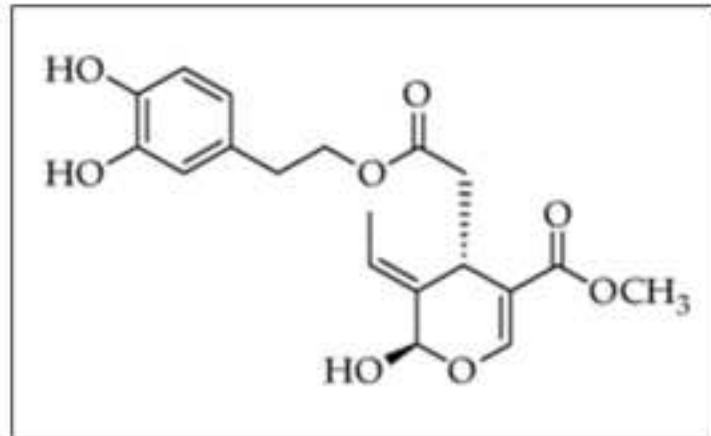
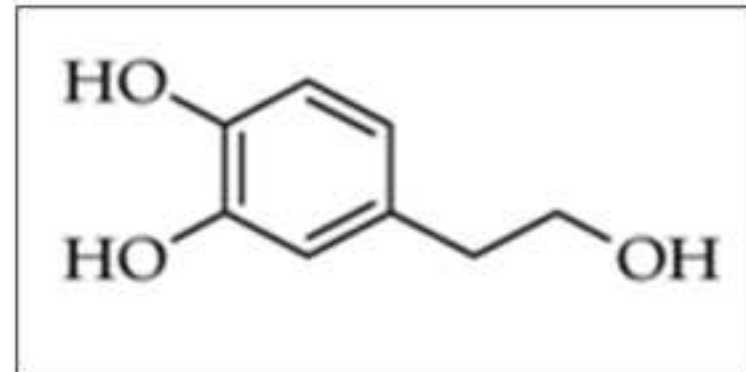
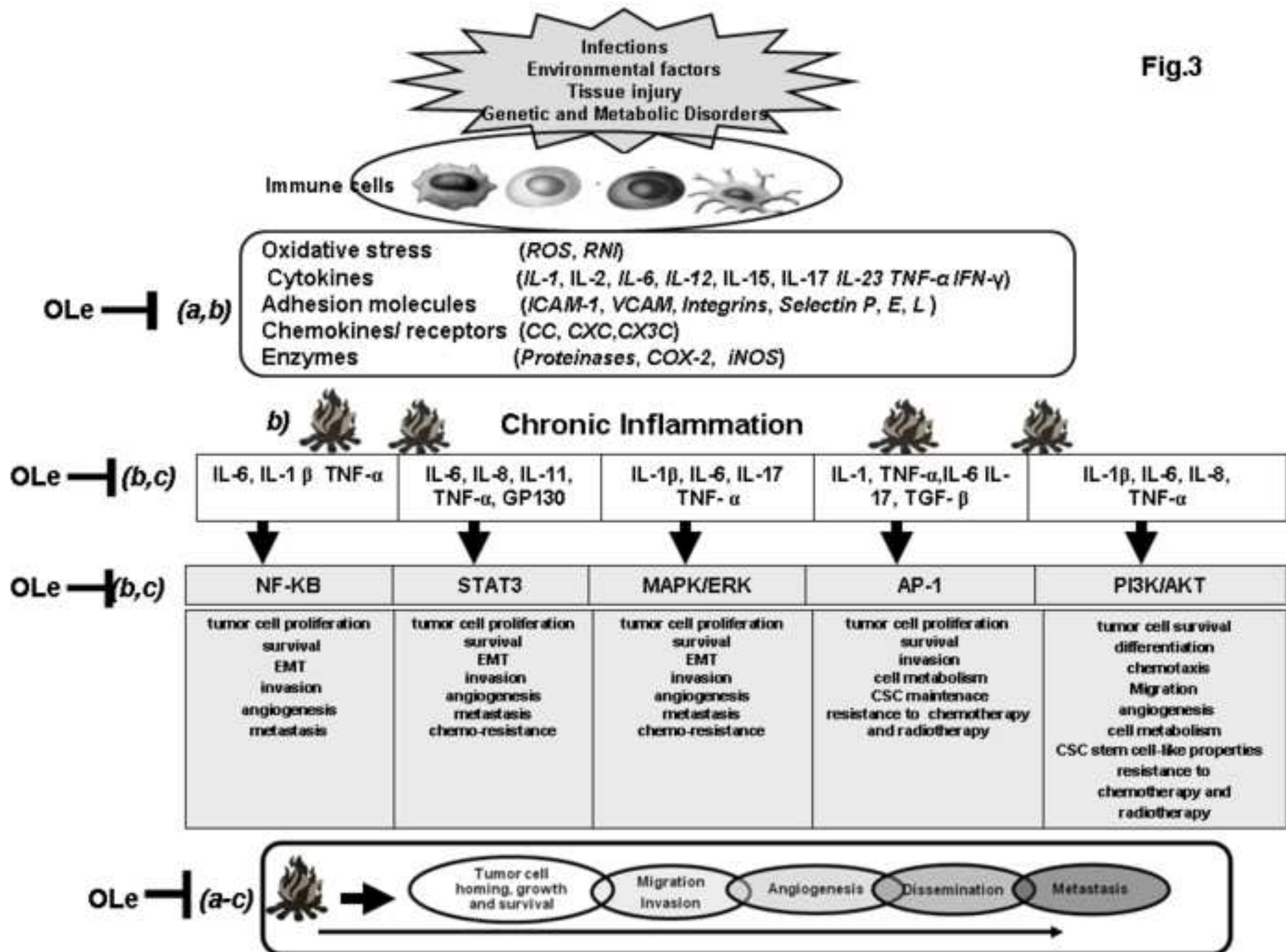
**Oleuropein (Ole)****Hydroxytyrosol (Hytr)****Oleuropein Aglycone (Ole-A)****Elenolic Acid (EIAC)**

Fig.3



Highlights

- Oleuropein, a phenolic compound present in olive leaves and fruits, exerts beneficial effects against various human diseases including cancer.
- This compounds may thwart cancer progression by modulating the expression and activity of several molecular pathways fostering this process.
- Most of these pathways also appear to favour the homing and growth of disseminating cancer cells in bones.
- These findings suggest a potential therapeutic role of this molecule in the prevention and treatment of malignant bone diseases.
- This paper provides a current overview regarding the results of recent experimental studies underpinning a possible clinical role of Ole in the prevention and development of cancer-related bone diseases.