EMULSIONS BASED ON FATTY ACID FROM VEGETABLE OILS FOR COSMETICS

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Abstract

- 12 Vegetable oils are sources of saturated, monounsaturated and polyunsaturated fatty acids. In this
- work, we studied the suitability of a mixture of vegetable oils and some fatty acids (similar to the those
- of the human skin) as a cosmetic emulsion. Hypericum (Hypericum perforatum L.) 1%, thistle
- 15 (Silybum marianum (L.) Gaertn.) 42%, linen (Línum usitatíssimum L.) 1.1%, wheat germ (Triticum
- 16 aestivum L.) 1%, sesame (Sésamum índicum L.) 1%, mustard (Sinápis álba L.) 2%, pumpkin
- 17 (Cucúrbita pépo L.) 5%, were selected as vegetable oils. The emulsion based on the proposed
- mixture of vegetable oils has a higher antioxidant activity (1.623 OD) compared with that based on
- mineral oil (0.427 OD). Finally, the emulsion provides a good balance of moisture and fat on the skin
- for one hour, in contrast to the short-term effect of the emulsion based on mineral oil. The obtained
- 21 data are promising for the formulation of emulsion with specific fatty acids composition.

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23 **Keywords:** vegetable oils, fatty acids, emulsion, antioxidant activity, moisture, skin.

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1. Introduction

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On the territory of modern Ukraine the steppe is 240 thousand km², or 40% of the entire territory of the country, and is the largest zonal natural complex (Pirko, 2004). The steppe is an area where in dry conditions develops the largest variety of herbaceous plants on the planet, which begin to bloom immediately after the snow melts, from the beginning of February. If in the forest there are from 2 to 5 types of plants per 1 m2, then in the steppe – more than 100 (Vasylyuk, 2021). Crop production in the steppe zone specializes in the production of winter wheat, barley, corn, soybeans, sunflower seeds, rapeseed, and the cultivation of vegetable crops (Bazalii et al., 2015). Vegetable raw materials have been recently used for numerous applications, including paper fabrication (Modica et al., 2020; Sottile et al., 2021), synthesis of anti-inflammatory products (Maione et al., 2016), protection of artworks (D'Agostino et al., 2021) and cosmetic emulsions (Chu and Nyam, 2020; Saribekova et al., 2021; Khafid et al., 2022). Among the natural materials, fatty vegetable oils (triglyceride) are valuable components of medicinal and decorative cosmetics. As a part of cosmetic compositions vegetable oils carry out, as a rule, some functions. They play the role of emollients, serve as a base in which other structural and functional components of the composition are located, perform a transport function of delivery of biologically active substances through the lipid barrier of the skin. The triglycerides of fatty acids, which form the basis of vegetable oils, are actively involved in the formation of structure, function and repair of cell membranes of the lipid barrier. Triglycerides of polyunsaturated fatty acids play an important role in the skin's immune system, participating in the synthesis of prostaglandins (Shepel, 2010; Hernandez and Margolina, 2017). In determining the composition of fatty acids (triglycerides), which would provide the most positive effect of a particular cosmetic product, it is advisable to use triglycerides that are part of the skin's lipid barrier in a normal healthy skin considering also their natural proportions (Shepel, 2010). Fatty acids, which are naturally present in the human epidermis, are mainly a mixture of linoleic C18:2 (21.5%), oleic C18:1 (15.1%), palmitic C16:0 (14.0%), stearic C18:0 (11.1%) and lignoceric C24:0 (10.0%)

- acids, Fig. 1 (Moore et al., 2020; Knox and O'Boyle, 2021). Acids such as arachidonic C20:4 (6.2%),
- 52 behenic C22:0 (2.7%), palmitinoleic C16:1 (2.3%), arachinoic C20:0 (1.6%), myristic C14:0 (1.1%),
- linolenic C18:3 (1.0%) are present in much smaller amounts but still play a key role in a healthy skin.

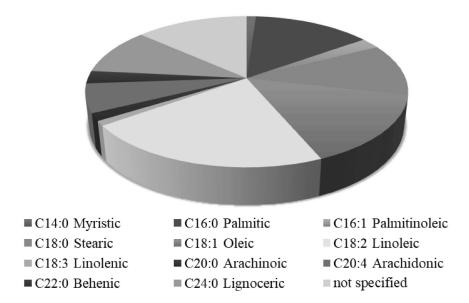


Figure 1. The average percentage of fatty acids in the epidermis of normal human skin.

It is reported (Shepel, 2010) that the mixture of oils, close in fatty acid composition of normal skin, was palmitic (2-3%), palmitoleic (13-17%), stearic (1-2%), oleic (30-36%), linoleic (20-26%) and linolenic (2-3%) for a cosmetic formulation. In addition, it is emphasizes the importance of maintaining the ratio of linoleic/oleic 1:1.8 and linoleic/linolenic acids 10:1. This quantitative ratio of linoleic/oleic and linoleic/linolenic acids is also reported by other authors (Nosenko et al., 2015, Mank and Polonska, 2016). Finally, the optimal ratio between saturated, mono- and polyunsaturated fatty acids is 1:1:1 when designing compositions of natural oils, which corresponds to that of lipid layers of the skin barrier layer (Mank and Polonska, 2016).

Recent literature reports slightly different values from the fatty acid composition of epidermis in normal skin (Moore et al., 2020; Knox and O'Boyle, 2021).

The more recent tendency is to select the composition of natural vegetable oils according to the fatty acid composition of the sebum of normal skin. Such a calculated mixture of oils should ensure the balance of moisture and fat on the skin treatment, as well as the antioxidant properties due to the presence of biologically active compounds in vegetable oils. As follows, the aim of the work is to develop a mixture of vegetable oils from the steppe zone of southern Ukraine with a fatty acid composition as close as possible to the fatty acid composition of the epidermis of normal skin for use as a fat phase in cosmetic emulsions.

2. Materials and Methods

2.1. Materials

79 2.1.1. Unrefined vegetable oil

In the work, we investigated samples of the unrefined vegetable oils typical of the steppe zone of southern Ukraine obtained by the press method: samples of oil of brier (*Rosa cinnamomea sensu L.*), sunflower (*Helianthus annuus L.*), wheat germ (*Triticum aestivum L.*), thistle (*Silybum marianum (L.*) *Gaertn.*), grape seeds (*Vitis vinifera L.*), sesame (*Sésamum índicum L.*), sea-buckthorn (*Hippóphaë rhamnóides L.*), camelina (*Camelina sativa (L.) Crantz*), linen (*Línum usitatíssimum L.*), pumpkin (*Cucúrbita pépo L.*), mustard (*Sinápis álba L.*), hemp (*Cánnabis sativa L.*); and method of extraction by oil corn (*Zéa máys L.*) refined deodorized: samples of oil of hypericum (*Hypericum perforatum L.*), chamomile (*Matricaria chamomilla L.*) and bidens (*Bídens tripartíta L.*), supplier company Leko Style (Ukraine).

2.1.2. Components of the cosmetic emulsion

Emulsifier – non-ionic emulsion system Emulgade SE-PF (INCI: Glyceryl Stearate (and) Ceteareth-20 (and) Ceteareth-12 (and) Cetearyl Alcohol (and) Cetyl Palmitate) – BASF SE, Germany; structuring

- 92 cetearyl alcohol (INCI: Cetearyl Alcohol) BASF SE, Germany; conservative Cosgard (INCI:
- 93 Benzyl alcohol, Dehydroacetic acid, Aqua) Lonza, Switzerland; pH regulator citric acid (INCI:
- 94 Citric acid) Nanjing Jiayi Sunway Chemical Co., Ltd, China; silicone dimethicone (INCI:
- 95 Dimethicone) BRB International BV, Netherlands.

97 2.2. Method of research of unrefined vegetable oils

- 99 2.2.1. Fatty acid composition of oils
- 100 The study was performed on a gas chromatograph Crystal-2000M, Agilent DB-FFAP column 50 m ×
- $0.320 \text{ mm} \times 0.50 \text{ }\mu\text{m}$ (Kuryata and Polyvanyi, 2018).
- 102 2.2.2. Refractive index
- The determination of the refractive index of unrefined vegetable oils was performed using the
- refractometric method (Bhagyaraj et al., 2020). To determine the refractive index of unrefined
- vegetable oils, a refractometer type IRF-22 was used.
- 106 *2.2.3. Color number on the iodine scale*
- The color of the test oil was evaluated by comparison with the color of standard iodine solutions in
- the range from 1 to 100 mg of iodine (Shimamoto et al., 2016).
- 109 2.2.4. Determination of acid number by titrometric method
- A portion of the test oil (emulsion) was added to a 250 mL conical flask. Add 50 mL of neutralized
- solvent mixture (diethyl ether and ethyl alcohol) and mix. The resulting solution was rapidly titrated
- with potassium or sodium hydroxide solution until a pale pink color persisted for 15 s. The acid number
- 113 (mg KOH/g) was calculated by the formula:

$$AN = \frac{V \cdot c \cdot 56.1}{m},\tag{1}$$

where c is the exact value of the molar concentration of the alkaline titrant, mol/1000 mL; V is the volume of alkaline titrant determined during indicator titration or calculated for the equivalent point during potentiometric titration, mL; 56.1 is the molar mass of potassium hydroxide, g/mol; m is the amount of oil (emulsion), g (Aricetti and Tubino, 2012).

2.3. Emulsion

2.3.1. Preparation of the emulsion

- To prepare the emulsion, oil, cetearyl alcohol and Emulgade SE-PF were heated in a glass in a water bath at 60°C. In parallel, the calculated amount of water and citric acid was heated in the second glass in a water bath to the same temperature (60°C). Both phases were mixed at a speed of 200 min⁻¹ for 5 min. The temperature of the mixture was periodically measured during the emulsification process.

 After cooling the emulsion to 30°C was introduced conservative Cosgard, stirring was continued for 5 min.
- *2.3.2. Emulsion colloidal stability*
 - The glass tubes were filled to 2/3 of the volume with the test emulsion, placed in a thermostat and kept at 42°C for 20 min. The height of the test tubes is 120 mm, the diameter is 14 mm. After that, the tubes were centrifuged at a speed of 100 s⁻¹ for 5 min. The emulsion was considered stable if no more than a drop of the aqueous phase or a layer of the oil phase of not more than 0.5 cm was isolated in the test tubes after centrifugation (Estanqueiro et al., 2014).
- *2.3.3. Determination of emulsion thermal stability*
 - Tubes with a capacity of 25 mL were filled to 2/3 of the volume with the tested emulsion, making sure that no air bubbles remained in them, closed with stoppers and placed in a thermostat with a temperature of 40 42°C. The contents of the tubes after 1 hour were gently stirred with a glass rod to remove air. The emulsions were kept in a thermostat for 24 hours. The emulsion was considered stable

if no aqueous phase evolution was observed in the tubes after thermostating, and an oil phase layer of no more than 0.5 cm is allowed to be released (Estanqueiro et al., 2014).

2.3.4. Determination of hydrogen index

The determination was performed in an aqueous extract using a pH meter ADWA AD1200. To prepare an aqueous extract of the emulsion, 10.00 g of the sample was placed in a beaker, 90 mL of distilled water was added, heated with stirring to 70°C until the emulsion was completely destroyed (separation of the oil layer), cooled to 25°C and the aqueous layer separated.

2.3.5. Determining the amount of moisture and fat on the skin

The amount of moisture and fat was determined using a tester with a digital high-sensitivity sensor Skin Detector SG-7D. The study involved 60 youth volunteers (22 – 35 years old), including 30 males and 30 females during autumn.

2.3.6. Antioxidant activity

The antioxidant activity of the cosmetic emulsion was determined using the Oyaizu method (FRAP method) [Faria et al., 2005; Harhaun et al., 2020]. A total of 0.2 mL of the sample was mixed with phosphate buffer (0.5 mL; 0.2 M; and pH 6.6) and potassium ferricyanide [K₃Fe(CN)₆] (0.5 mL; 1% solution). The resulting mixture was kept at 50°C for 20 min, then 0.5 mL of a 30% solution of trichloroacetic acid was added to the mixture and filtered. To 0.5 mL of the obtained filtrate was added ferric (III) chloride FeCl₃ (0.1 mL, 0.1% solution). The optical density was determined using a ULAB 102 spectrophotometer (Shanghai Metash Instruments Co., Ltd., Shanghai, China) at a wavelength of 700 nm. The increase in the absorption of the reaction mixture indicates an increase in the ability of the samples to reduce ferric iron (Fe³⁺ \rightarrow Fe²⁺). A 20% solution of ascorbic acid was used as a reference solution.

2.4. Statistical processing of the obtained results

The software Origin 6.1 (OriginLab Corporation, UK) was used for statistical processing of the obtained results. The level of statistical significance is p<0.05.

3. Results and discussion

At the first stage of the study, the characteristics of the studied samples of vegetable oils were determined: refractive index, acid number and color number on the iodine scale (Table 1).

Table 1. Characteristics of the studied samples of vegetable oils

0.1	Refractive index	Acid number,	Color number on	
Oil	Refractive index		the iodine scale	
Brier	1.475	0.8	70	
Sunflower	1.473	0.3	15	
Wheat germ	1.471	0.3	25	
Thistle	1.471	0.7	10	
Grape seeds	1.474	0.7	20	
Sesame	1.472	0.4	5	
Bidens	1.474	0.4	10	
Sea buckthorn	1.441	1.6	Over 100	
Chamomile	1.474	0.4	5	
Hypericum	1.473	0.4	25	
Camelina	1.476	0.9	40	
Linen	1.481	2.2	40	
Pumpkin	1.473	0.5	50	
Mustard	1.474	0.8	25	
Hemp	1.476	1.4	20	

According to Table 1, the studied samples of vegetable oils are identical in terms of refraction (Gunstone, 2004; Mukhametov et al., 2022). The acid number corresponds to the indicators specified in the quality certificates of vegetable oil. Analysis of the color number shows that the most colored are samples of sea buckthorn oil – more than 100 units and brier oil – 70 units of iodine scale, whilst the least colored were samples of sesame oil and chamomile – 5 units of iodine scale. The data obtained should be taken into account when creating a composition of vegetable oils due to possible undesirable color (Sharma et al., 2022).

The fatty acid composition of the studied samples of vegetable oil is provided in Table 2.

Table 2. Fatty acid mass percentage composition of vegetable oils

kin mustard hemp	0.074 0.043		0.023	5.468	5.468	5.468	5.468	5.468 0.172	5.468 0.172 2.727 5 32.035	5.468 0.172 2.727 2.727 5 32.035	5.468 0.172 2.727 32.035 47.046 5.041	5.468 0.172 2.727 5 32.035 2 47.046 5.041 0.604	5.468 0.172 2.727 2.727 47.046 5.041 0.604 5.291	5.468 0.172 2.727 2.727 3.2.035 4.7.046 5.041 0.604 5.291	5.468 0.172 2.727 32.035 47.046 5.041 0.604 5.291 0.417	5.468 0.172 2.727 5 32.035 2 47.046 5.041 0.604 5.291 0.417	5.468 0.172 2.727 2.727 2.47.046 5.041 0.604 5.291 0.417 0.664	5.468 0.172 2.727 3.2.035 47.046 5.041 0.604 5.291 0.417 0.158
linen pumpkin		0.049 0.10/									m /0 +	m 10 H	60 VO #	m vo +		m /n +		co 10 H
hypericum camelina	0.070 0.070		0.025	0.025						m 81								
chamomile	0.072 0.00			6.569 6.4														
bidens sea buckthorn	74 0.078		0.015						10									
sesame	0.021 0.074			9.448 6.563					10									
thistle grape seeds	0.085 0.070		0.036	0.036 7.088 6.694														
wheat germ	0.075			6.630														
Oil brier sunflower	0.070 0.064			6.413 6.340														
Fatty acid b	$C14:0^{a}$ 0		C15:0															

^a The blue color indicates the fatty acids found in normal skin.

In the analysis of the fatty acid composition of the studied samples of vegetable oil, the main attention was paid to the fatty acids of normal skin (Fig. 1): C14: 0, C16: 0, C16: 1, C18: 0, C18: 1, C18: 2, C18: 3, C20: 0, C22: 0, C24: 0. Based on the data in Table 2, samples of oils with the maximum content of fatty acids detected in normal skin were identified (Table 3).

Table 3. Samples of oils with the maximum content of certain fatty acids

Fatty acid	Oil	Value, %
Myristic C14:0	Pumpkin	0.107
Palmitic C16:0	Pumpkin	9.455
Palmitinoleic C16:1	Mustard	0.172
Stearic C18:0	Thistle	5.786
Oleic C18:1	Sesame	40.536
Linoleic C18:2	Hypericum	61.992
Linolenic C18:3	Linen	51.094
Arachinoic C20:0	Thistle	3.724
Arachidonic C20:4	_	_
Behenic C22:0	Thistle	2.480
Lignoceric C24:0	Wheat germ	0.036

Therefore one can conclude that among the studied samples of vegetable oil, the following should be selected for the proper skin treatment formulation: pumpkin, mustard, thistle, sesame, hypericum, linen and wheat germ. It should be noted that arachidonic acid C20:4 was not detected in any of the studied samples of vegetable oil (Vicentini-Polette et al., 2021). The percentage of selected vegetable oils was calculated, which in the ratio of essential fatty acids is as close as possible to the ratio of fatty acids in normal healthy skin (Table 4).

Table 4. Estimated composition of basic fatty acids of the calculated mixture

Oil	Concentration, %
Hypericum	1.0
Thistle	42.0
Linen	1.1
Wheat germ	1.0
Sesame	1.0
Mustard	2.0
Pumpkin	5.0

The fatty acid composition of normal skin (a) and the calculated mixture of oils (b) are compared in Fig. 2.

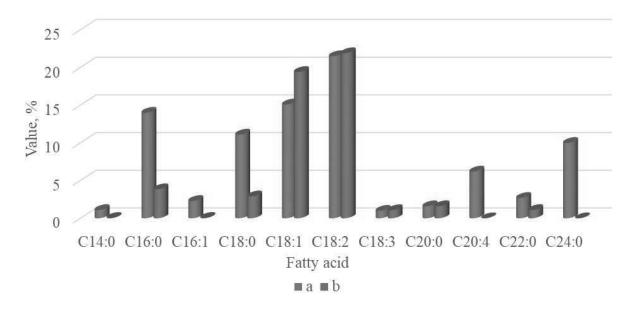


Figure 2. Fatty acid composition: a – normal skin, b – calculated mixture of vegetable oils.

The calculated mixture of vegetable oils does not contain arachidonic acid C20:4 (Figure 2). Also the content of fatty acids such as myristic C14:0, palmitic C16:0, palmitinoleic C16:1, stearic C18:0, behenic C22:0 and lignoceric C24:0 is relatively small (Figure 2). Optimal values have been achieved for acids such as oleic C18:1, linoleic C18:2, linolenic C18:3 and arachinoic C20:0 (Figure 2).

For the sake of comparison, we developed mixture of vegetable oils, – Emulsion I and a similar composition with the replacement of the mixture of oils with mineral oil – Emulsion II (Table 5). The Emulsion II was proposed by us in a recent paper (Kunik et al., 2019).

Table 5. Prescription composition of emulsion cosmetics

C	Concentration, %							
Component	Emulsion I	Emulsion II						
Mixture of vegetable	25.0							
oils	25.0	_						
Mineral oil	_	25.0						
Emulgade SE-PF	3.0							
Cetearyl alcohol	3.0							
Dimethicone	3.0							
Cosgard	0.5							
Citric acid	To pH 5.5							
Distilled water	To 100							

The obtained samples of the emulsion were investigated to determine the organoleptic, physical and chemical characteristics (Table 6).

Table 6. Organoleptic, physical and chemical characteristics of the obtained samples of

emulsions

Indicator	Emulsion I	Emulsion II				
Appearance	Homogeneous emulsion, without impurities					
Color	Yellow	White				
Scent	Characteristic scent of vegetable	Neutral				
	oils					
Colloidal stability	Stable					
Thermal stability	Stable					
Hydrogen index	5.5					
(pH)	3.3					
Antioxidant						
activity, optical	1.623	0.427				
density D						

The obtained data show that the obtained emulsions are stable colloidal system. The antioxidant activity of the emulsion I significantly exceeds the antioxidant activity of the Emulsion II based on mineral oil. The obtained data are consistent with (Saha and Ghosh, 2011). Next, we determined the effect of the developed composition of the cosmetic emulsion on the

amount of moisture and fat on the skin of the forearm (Table 7).

Table 7. Determining the effect of moisture and fat on the skin

Time	Indicators of without cosm emulsion		Cosmetic emu		Cosmetic emulsion based on unrefined vegetable oils		
	moisture, %	fat, %	moisture, %	fat, %	moisture, %	fat, %	
Before	21.0/20.4 ^a	31.3/33.5°	21.0/20.4	31.1/33.1	21.1/20.4	31.0/32.9	
application Immediately							
after	20.9/20.0 ^b	30.5/32.7	48.4/45.9	32.5/35.3	52.9/50.3	38.5/40.3	
application							
After 1 hour	21.3/19.8	31.9/33.4	25.2/21.7	33.6/36.1	35.6/33.4	44.4/45.2	
After 2 hours	20.8/20.1	31.2/33.2	21.5/20.8	32.0/33.2	29.8/26.5	37.8/39.6	
After 3 hours	21.0/20.0	31.2/33.5	21.2/20.5	31.8/33.9	25.2/23.0	38.7/39.2	
After 4 hours	21.2/20.1	31.1/33.0	21.0/20.3	31.6/33.2	20.7/19.5	31.0/33.8	

²²⁶ The average amount of moisture and fat on the skin of female and male volunteers.

According to Table 7, immediately after applying both emulsions on the skin of volunteers, a balance of moisture and fat was observed. An hour later, the balance of moisture and fat was observed only after applying a sample of emulsion I. After two hours, the balance of moisture and fat was lost, although the moisture and fat content of the skin after applying the emulsion I was higher than after applying the emulsion II with mineral oil. Three hours after the application

^b The red color indicates an imbalance of moisture and fat on the skin, green – the balance.

^c The level of statistical significance is p<0.05.

of emulsion I, the moisture and fat content of the skin was still 20 - 24% higher than the moisture and fat content of the skin without applying the emulsion. The indicators of moisture and fat content on the skin did not differ from the indicators of skin without emulsion application after 4 hours from application. It should be noted that with the systematic application of the developed composition of the emulsion with a mixture of oils, it is possible to expect a longer effect of the balance of moisture and fat on the skin.

The results also show that women's skin, unlike men's, is more moisturized and less oily. The obtained data are fully consistent with literature reports (Patel et al., 2017; Rahrovan et al., 2018). This work proves the perspective of taking into account the composition of fatty acids in the formulation of emulsions for cosmetic purposes. Further research will be aimed at prolonging the

moisturizing effect by including in the mixture of vegetable oils rich in myristic C14:0, palmitic

C16:0, palmitoleic C16:1, stearic C18:0, arachidonic C20:4 and lignoceric C24:0 fatty acids.

4. Conclusions

Selected samples of vegetable oil from steppe zone of southern Ukraine were characterized from the physico-chemical point of view (refractive index, color and acid number). And also by determining the fatty acid composition. A formulation for cosmetic application was prepared by considering the ratio of fatty acids as close as possible to that in healthy skin. An additional emulsion formulation containing mineral oil was considered and the performances were compared to the vegetable oil based one. It was demonstrated that the vegetable oil based emulsion has higher antioxidant activity and it provides a balance of moisture and fat on the skin for longer time compared to the emulsion based on mineral oil.

Declaration of Interest statement

No potential conflict of interest was reported by the authors.

260 **REFERENCES**

- Aricetti, J., Tubino, M., 2012. A Visual titration method for the determination of the acid
- number of oils and fats: a green alternative. J. Am. Oil Chem.' Soc. 89(11), 2113–2115.
- 263 <u>https://doi:10.1007/s11746-012-2111-1</u>.
- Bazalii, V., Zinchenko, O., Lavrynenko, Yu., Salatenko, V., Kokovikhin, S., Domaratskyi E.,
- 265 2015. Horticulture. Grin D.S., Kherson.
- Bhagyaraj, S.M., Oluwafemi, O.S., Krupa, I., 2020. Chapter 13 Polymers in optics. Polymer
- 267 Science and Innovative Applications. 423–455. https://doi.org/10.1016/B978-0-12-816808-
- 268 0.00013-5.
- 269 Chu, C., Nyam, K., 2020. Kenaf (Hibiscus cannabinus L.) seed oil: Application as cosmetic
- product ingredients. Ind. Crops Prod. 156, 112871.
- 271 https://doi.org/10.1016/j.indcrop.2020.112871.
- D'Agostino, G., Badalamenti, N., Franco, P., Bruno, M., Gallo, G., 2021. The chemical
- composition of the flowers essential oil of *Inula crithmoides* (Asteraceae) growing in aeolian
- islands, Sicily (Italy) and its biocide properties on microorganisms affecting historical art crafts.
- 275 Nat. Prod. Res. 36(12), 2993–3001. https://doi:10.1080/14786419.2021.1938040.
- Estanqueiro, M., Conceição, J., Amaral, M.H., Santos, D., Silva, J.B., Lobo, J.M., 2014.
- 277 Characterization and stability studies of emulsion systems containing pumice. Braz. J. Pharm.
- 278 Sci. 50(2), 361–369. https://doi:10.1590/S1984-82502014000200016.
- Faria, A., Oliveira, J., Neves, P., Gameiro, P., Santos-Buelga, C., Freitas, V., Mateus, N., 2005.
- Antioxidant properties of prepared blueberry (Vaccinium Myrtillus) extracts. J. Agric. Food
- 281 Chem. 53(17), 6896–902. https://doi: 10.1021/jf0511300.
- Gunstone, F.D., 2004. The chemistry of oils and fats. Sources, composition, properties and

- uses. Oxford: Blackwell Publishing Ltd.
- Harhaun, R., Kunik, O., Saribekova, D., Lazzara, G., 2020. Biologically active properties of
- plant extracts in cosmetic emulsions. Microchem. J. 104543.
- 286 https://doi:10.1016/j.microc.2019.104543.
- Hernandez, E., Margolina, A., 2017. New cosmetology. Fundamentals of modern cosmetology.
- 288 Moscow: Cosmetics and Medicine.
- Khafid, M., Islamiati, D., Ika, Yu., Sisunandar, S., Binar, A., 2022. Oil from kopyor coconut
- 290 (Cocos nucifera var. Kopyor) for cosmetic application. Ind. Crops Prod. 115221.
- 291 https://doi.org/10.1016/j.indcrop.2022.115221.
- Knox, S., O'Boyle, N.M., 2021. Skin lipids in health and disease: A review. Chem. Phys.
- 293 Lipids. 236, 1–14. https://doi:10.1016/j.chemphyslip.2021.105055.
- Kunik, O., Saribyekova, D., Saleba, L., Ivakhnenko, H., Panchenko, Y., 2019. Research of
- 295 physical and chemical properties of cosmetic emulsions of oil in water type based on
- polyorganosyloxanes and their alternative substitute. Chem. Chem. Technol. 13(4), 526–534.
- 297 https://doi:10.23939/chcht13.04.
- Kuryata, V., Polyvanyi, S., 2018. Formation and functioning of source-sink relation system of
- oil poppy plants under treptolem treatment in connection with productivity of crop. Ukr. J. Ecol.,
- 300 8(1), 11–20. https://doi:10.15421/2018 182.
- Maione, F., Russo, R., Khan, H., Mascolo, N., 2016. Medicinal plants with anti-inflammatory
- activities. Nat. Prod. Res. 30(12), 1343–1352. https://doi:10.1080/14786419.2015.1062761.
- Mank, W., Polonska, T., 2016. Vegetable oils compositions for cosmetic products. Scientific
- Works of National University of Food Technologies. 22(3), 217–223.
- 305 http://sw.nuft.edu.ua/Archiv/2016/swnuft 22 3.pdf (accessed 19 May 2022).
- Modica, A., Rosselli, S., Catinella, G., Sottile, F., Catania, C.A., Cavallaro, G., Lazzara, G.,

- 307 Botta, L., Spinella, A., Bruno M., 2020. Solid state 13C-NMR methodology for the cellulose
- 308 composition studies of the shells of *Prunus dulcis* and their derived cellulosic materials.
- 309 Carbohydr. Polym. 116290. https://doi:10.1016/j.carbpol.2020.116290.
- Moore, E.M., Wagner, C., Komarnytsky, S., 2020. The enigma of bioactivity and toxicity of
- botanical oils for skin care. Front. pharmacol. 11(785), 1–17.
- 312 https://doi:10.3389/fphar.2020.00785.
- Mukhametov, A., Mamayeva, L., Kazhymurat, A., Akhlan, T., Yerbulekova, M., 2022. Study of
- vegetable oils and their blends using infrared reflectance spectroscopy and refractometry. Food
- 315 Chemistry. X (2022). https://doi.org/10.1016/j.fochx.2022.100386.
- Nosenko, T., Voloshchenko, T., Sidorenko, T., 2015. Cosmetic oil for skin cleansing. Bulletin
- of the National Technical University «KhPI». 30(1139), 72–78.
- 318 <u>http://nbuv.gov.ua/UJRN/vcpixx 2015 30 13</u> (accessed 19 May 2022).
- Patel, K., Gajjar, P.C., Mehta, H.H., Mehta, H., Solanki, J., 2017. Study of moisture content on
- various skin sites in different seasons in Indian population. Int. J. Clin. Exp. 4, 190–194.
- 321 <u>https://doi:10.4103/ijcep.ijcep_3_18</u>.
- Pirko, V., 2004. Settlement and economic development of Steppe Ukraine in the XVI-XVIII
- 323 centuries. Eastern Publishing House, Donetsk.
- Rahrovan, S., Fanian, F., Mehryan, P., Humbert, P., Firooz, A., 2018. Male versus female skin:
- What dermatologists and cosmeticians should know. Int. J. Womens Dermatol. 4(3), 122–130.
- 326 <u>https://doi:10.1016/j.ijwd.2018.03.002</u>.
- Saha, S., Ghosh, M. 2011. Antioxidant effect of vegetable oils containing conjugated linolenic
- acid isomers against induced tissue lipid peroxidation and inflammation in rat model. Chem.-
- 329 Biol. Interact. 190 (2–3), 109-120. https://doi.org/10.1016/j.cbi.2011.02.030.
- Saribekova, D., Kunik, O., Harhaun, R., Saleba, L., Cavallaro, G., 2021. The use of silicones as

- extractants of biologically active substances from vegetable raw materials. Appl. Sci. 11(22),
- 332 10625. https://doi:10.3390/app112210625.
- 333 Sharma, M., Hussain, Sh., Shalima, T., Aav, R., Bhat, R., 2022. Valorization of seabuckthorn
- pomace to obtain bioactive carotenoids: An innovative approach of using green extraction
- techniques (ultrasonic and microwave-assisted extractions) synergized with green solvents
- 336 (edible oils). Ind. Crops Prod. 175. 114257. https://doi.org/10.1016/j.indcrop.2021.114257.
- Shepel, V., 2010. On the composition of mixtures of vegetable oils for cosmetic compositions.
- Publications of the Touche Flora company.
- 339 https://tusheflora.ru/publication/articles/o sostavlenii smesej rastitelnih masel dlya kosmetich
- 340 <u>eskih_kompozitcij/</u> (accessed 19 May 2022).
- 341 Shimamoto, G.G., Aricetti, J.A., Tubino, M., 2016. A simple, fast, and green titrimetric method
- for the determination of the iodine value of vegetable oils without Wijs solution (ICl). Food Anal.
- 343 Methods. 9(24), 79–83. https://doi:/10.1007/s12161-016-0401-1.
- Sottile, F., Modica, A., Rosselli, S., Catania, C.A., Cavallaro, G., Lazzara, G., Bruno, M., 2021.
- Hand-made paper obtained by green procedure of cladode waste of *Opuntia ficus indica* (L.)
- 346 Mill. from Sicily. Nat. Prod. Res. 35(3), 359–368. https://doi:10.1080/14786419.2019.1631820.
- Vasylyuk, O. The steppe is the most efficient storage of atmospheric carbon, 2021. Institute of
- Zoology named after I.I. Schmalhausen of the National Academy of Sciences of Ukraine.
- 349 https://www.nas.gov.ua/UA/Messages/Pages/View.aspx?MessageID=8147 (accessed 22 August
- 350 2022).
- Vicentini-Polette, C., Ramos Paulo, R., Gonçalves, C., De Oliveira, A. 2021. Determination of
- free fatty acids in crude vegetable oil samples obtained by high-pressure processes. Food Chem.:
- 353 X. 12. 100166. https://doi.org/10.1016/j.fochx.2021.100166.