

Photostability assessment of natural pyrethrins using halloysite nanotube carrier system

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1 Abstract

2 Natural pyrethrins are one of the most used pesticides and insect repellents employed for domestic
3 or agronomic use. However, they are highly hydrophobic and suffer from insolubility in water
4 media and ready decomposition by photochemical, and oxidative actions. These aspects hamper
5 their long-term storage and applicability. Herein, we report the synthesis and characterization of a
6 nanomaterial based on the loading of pyrethrum extract (PE) into natural, low-cost, and eco-
7 compatible halloysite nanotubes (Hal). The Hal/PE nanomaterial was thoroughly characterized, and
8 the morphology was imaged by TEM and SEM investigations.

9 Release experiments showed a slow release of pyrethrins from the carrier, and photodegradation
10 studies, both in solution and in a solid state, highlighted that the pyrethrum extract in the Hal/PE
11 nanomaterial showed high stability to UV irradiation, as proved by UPLC-ESI-MS, UV-*vis*, and
12 circular dichroism measurements.

13 Furthermore, to check if the obtained nanomaterial possesses the insecticidal activity of the
14 pyrethrum, *in vivo* tests on two model insects, *Galleria mellonella* and *Tenebrio molitor*, were
15 performed. The pyrethrins loaded onto Hal showed the same activity as standard pyrethrins, at half
16 dose compared to them.

17 Keywords: pyrethrins, halloysite, photoprotection, nanopesticides.

18

1. Introduction

Agriculture is the major source of food in the world. Nowadays, environmental pollution, climate change, and agricultural crop pests jeopardize plant cultivation hampering overall food production (Gomez-Zavaglia et al., 2020; Praveen and Sharma, 2019). In this context, commonly used pesticides are more and more inefficient due to the occurrence of resistance phenomena (Bombo et al., 2019; Martin, 2004).

Plant extracts or their bioactive compounds are increasingly used as alternatives to synthetic pesticides for sustainable pest control, with less impact on the environment and human health.

Pyrethrum, a natural extract of *Tanacetum cinerariifolium* (Trevir.) Sch.Bip., represents one of the most commonly used pesticides and insect repellents, both for agricultural and domestic purposes (Kalaitzaki et al., 2015; Markham et al., 2021; Papanikolaou et al., 2018). The extract constituents, responsible of its insecticidal and repellent activity are six esters (pyrethrin I, pyrethrin II, cinerin I, cinerin II, jasmolin I and jasmolin II), commonly known as pyrethrins. The biological activities of the pyrethrum constituents depend on the structures and stereochemical characteristics of both the acid and alcohol components. Among the six components, pyrethrin I and II are the most dominant and active (Jeran et al., 2021).

Unfortunately, although the pyrethrum extract possesses a high insecticidal action and low residual power, it suffers from ready decomposition by thermal, photochemical, and oxidative actions which hamper its long-term storage and applicability (Zhang et al., 2019). In addition, due to its hydrophobic chemical composition, organic solvents are necessary in its formulations which contribute to environmental pollution.

In recent years, the use of nanomaterials as a carrier for pesticides (Kah et al., 2018; Zhong et al., 2017), that can take over organic solvents in their formulations, has offered some advantages in terms of enhanced stability, toward sustainability and environmental considerations compared to the traditional approaches for extensive potential future use of natural insecticides on large scale (Li et al., 2022; Zhai et al., 2020; Zhong et al., 2017). For example, hexenoic acid, an insect pheromone,

1 was successfully loaded onto zinc layered hydroxide to develop a new generation of
2 environmentally safe pesticide nanomaterial for crop protection (Ahmad et al., 2015). Similarly,
3 some essential oils were loaded onto different clays obtaining an aroma-controlled release systems
4 for pest control (de Oliveira et al., 2022; Saucedo-Zuñiga et al., 2021).

5 Halloysite is a natural clay mineral (JOUSSEIN et al., 2005) derived from weathering rocks present
6 in tropical and subtropical areas since the origin of life. It is an aluminosilicate clay with a
7 predominantly hollow tubular structure with dimensions in the nanometric range and usually, it is
8 referred to as halloysite nanotubes (Hal) (Kogure, 2016; R. Price, 2001). Chemically Hal is similar
9 to the platy kaolinite ($\text{Al}_2\text{Si}_2\text{O}_5(\text{OH})_4 \cdot n\text{H}_2\text{O}$) with an inner surface mainly composed of aluminol
10 groups and an external one constituted by siloxane groups with the presence of few silanols groups
11 as structural defects that are fundamental for chemical modifications of the external surface (Long
12 et al., 2022; Massaro et al., 2018; Massaro et al., 2022a). Generally, the length of the tubes is in the
13 range of 0.2–1.5 μm , while their inner and outer diameters are in the ranges of 10–30 nm and 40–70
14 nm, respectively. The different chemical composition makes the tubes undergo ionization in
15 aqueous media, in a wide range of pH (Bretti et al., 2016). In particular, Hal possesses positive
16 charges in the inner lumen and negative ones at the external surface. The Hal adsorption capacity
17 towards several organic molecules is well documented and the resultant nanomaterials have found
18 applications in several fields (Abdullayev and Lvov, 2011; Massaro et al., 2020; R. Price, 2001).
19 The Hal ability of the controlled release of organic molecules from the lumen is probably the most
20 attractive aspect of this nanomaterial (Ghezzi et al., 2018; Glotov et al., 2021; Hasani et al., 2021;
21 Massaro et al., 2022b; Riela et al., 2021; Santos et al., 2018).

22 Hal is considered a safe material, able to penetrate the cell membranes (Massaro et al., 2022b; Prinz
23 Setter and Segal, 2020), that exerts relevant toxicity in *vitro* at the concentration of 1000 $\mu\text{g}/\text{mL}$
24 (Ahmed et al., 2015) whereas in *vivo* the oral administration limit is at ca. 20 mg/kg BW (Long et
25 al., 2018; Wang et al., 2018). Furthermore, it was found that by feeding nematodes with them, no
26 toxic effects were induced (Fakhrullina et al., 2015). The understanding of the phytotoxicity of Hal

1 is helpful for their applications in the environmental protection and agricultural fields (Huang et al.,
2 2021). In this context, some studies were reported about the environmental impact of Hal (Chen et
3 al., 2021a). Recent studies indeed showed that Hal possesses no phytotoxic effects on *Raphanus*
4 *sativus* L (Bellani et al., 2016) and on wheat, while having even positive effect on the growth of
5 tobacco cells and secondary metabolite (Chen et al., 2021b). Furthermore, few studies report the use
6 of Hal as a nanocontainer for pesticides which benefit from sustained and controlled release. Ge et
7 al. for example reported the encapsulation of myrcene into Hal for further environmental
8 applications (Chen et al., 2022; Li et al., 2022). Up to now, for the best of our knowledge, no
9 studies are reported about the delivery of natural pyrethrins by Hal and the subsequent *in vivo*
10 studies on insects as potential pesticides.

11 This study is focused on the advantages that pyrethrins should have if carried by Hal in terms of
12 prevention of photodegradation (da Rocha et al., 2022) (both in solution and in solid state),
13 controlled release (Shao et al., 2022) and toxic effect on model insects.

14 To realize these goals, herein we report the preparation of Hal loaded with pyrethrins from natural
15 pyrethrum extract PESTANALTM (Hal/PE) for domestic or agronomic use. The successful loading
16 of the extract into the Hal lumen was confirmed by thermogravimetric analysis (TGA) and Fourier
17 transform infrared (FT-IR) spectroscopy and the morphology of the nanomaterial was imaged by
18 transmission and scanning electron microscopy measurements (TEM and SEM, respectively). The
19 photostability of the nanomaterial under UV irradiation both in solution and in the solid state was
20 also assessed by UV-*vis* spectroscopy, circular dichroism, and ESI-MS.

21 Finally, as proof of concept, the toxic effects of the Hal/PE obtained, were evaluated *in vivo* on
22 selected target insects, i.e., adults of the yellow mealworm *Tenebrio molitor* L. and young larvae of
23 the greater wax moth *Galleria mellonella* (L.) in terms of mortality and sub-lethal effects induced in
24 the target insects. The two insects are respectively known as pests of stored grains and foodstuff
25 (Hill, 1990) and of *Apis mellifera* L. hives (Kwadha et al., 2017), respectively. Moreover, *G.*
26 *mellonella* larvae are widely used as model insects in laboratory trials, since they may be

1 maintained quite easily in continuous colonies, with relatively low production costs, they show a
2 quite rapid life cycle (about 6 weeks), survival at 30°C - 35°C, and generally do not require ethical
3 permits (Dindo et al., 2003; Ignasiak and Maxwell, 2017; Kay et al., 2019). The results support the
4 potential use of nanomaterials based on Hal and pyrethrins as natural pesticides, as well as the
5 safety nature of the resulting organic/inorganic nanomaterial ensures new environmentally friendly
6 pesticides.

7 **2. Materials and Methods**

8 The pyrethrum extract (PE), PESTANALTM (extract botanical insecticide, CAS 8003-34-7 -
9 analytical standard), was purchased from Sigma-Aldrich Chemical Co, and used without further
10 purification. Depending on the batch used it contains 40.7 wt% (Batch number: BCBZ5050) or ca.
11 54 wt% (Batch number: BCCD5535), of pyrethrins content. In particular, the first batch was used
12 for the photodegradation studies, and the second one for the preparation and characterization of
13 Hal/PE nanomaterial. Halloysite (Merck) was used as purchased without further purification. It
14 presented an average tube diameter of 50 nm and an inner lumen diameter of 15 nm. Typical
15 specific surface area is 65 m² g⁻¹; pore volume of ~1.25 cm³ g⁻¹; refractive index 1.54 and specific
16 gravity 2.53 g cm⁻³.

17 Commercial pyrethrum used as a positive control in *in vivo* studies was from Bayer (Pyrethrum
18 Actigreen).

19 FT-IR spectra (KBr) were recorded with an Agilent Technologies Cary 630 FT-IR spectrometer.
20 Specimens for these measurements were prepared by mixing 5 mg of the sample powder with 100
21 mg of KBr.

22 GC-MS used was on a single quadrupole Shimadzu GC-MS-QP2010 Plus equipped with an AOC-
23 20i autoinjector (Shimadzu, Kyoto, Japan) and a Supelcowax 10 capillary column (30 m long, 0.25
24 mm i.d., 0.25 µm film thickness). One µl of each sample was injected at 250°C in the splitless
25 mode, and the column flow (carrier gas: helium) was set at 1 mL/min. The GC oven temperature
26 was held for 5 min at 40°C, then increased by 2°C/min to 250°C, held for 15 min, and finally raised

1 to 270°C at 10°C/min. The MS interface worked at 280°C, and the ion source at 250°C. Mass
2 spectra were taken at 70 eV (in EI mode) from m/z 30 to 500. The GC/MS data were analysed using
3 the GCMSolution package, Version 2.7.

4 For Transmission Electron Microscope (TEM) observation the samples were prepared using a drop
5 of water suspension on a formvar coated copper grid (400 mesh) and allowing the drop to dry
6 completely in a vacuum desiccator. The TEM images of the samples were obtained using a Philips
7 TEM CM 100 transmission electron microscope at accelerating voltage = 80 kV.

8 The thermogravimetric analysis (TGA) of the material was performed in a TGA/DSC1 STAR
9 System from Mettler Toledo Inc. The sample (15 mg) was subjected to a pre-treatment in air flow
10 (30 mL/min) from 25°C to 100°C with a heating rate of 10°C/min and holding time at 100°C for 30
11 min, to remove any eventual physisorbed water. Then, the temperature was increased from 100 to
12 1000°C under air flow (30 mL/min) and the weight loss occurring during this step was considered
13 to calculate the organic weight content of the Hal/PE nanomaterial.

14 The various components of the aqueous pyrethrum extract were analysed by a UPLC
15 chromatographic system, Waters ACQUITY H-CLASS model, coupled with a Waters Xevo G2-XS
16 QToF (Waters) mass spectrometer, with an ESI-APCI type source. UPLC conditions: ramp rate
17 accelerated to 2 mL/min in 0.45 min, mobile phase A was water, B was acetonitrile.

18 Xevo G2-XS QToF conditions: Positive mode, Cone Voltage: 20 V, Capillary: 0.8 kV, Collision
19 Energy: 6.00 eV, Source Temperature: 120°C, Desolvation Temperature: 600°C, Cone Gas Flow:
20 50 L/h, Desolvation Gas Flow: 1000 L/h.

21 UV-*vis* spectra were acquired on a Jasco V-550 spectrophotometer.

22 Circular Dichroism (CD)/UV measurements were performed at room temperature on a Jasco J-715
23 spectropolarimeter. Spectra were recorded at a scanning speed of 100 nm/min and five consecutive
24 scans in the 400-205 nm range were averaged for each sample. Electrospray ionization mass
25 spectrometry (ESI-MS) analysis was carried out on a Micromass ZQ-4000 single quadrupole mass
26 spectrometer operating at 4000 m/z. An infusion syringe pump was used for direct injection of the

1 sample at a rate of 10 $\mu\text{l}/\text{min}$. Operating parameters were: capillary voltage 3 kV, cone voltage 30
2 V, capillary temperature 120°C, desolvation temperature 220°C. Mass analysis was carried on in
3 the 100-1300 amu range (positive ionization mode).

4 **2.1 Synthesis of the Hal/PE nanomaterial**

5 To a dispersion of Hal (100 mg) in water (5 mL), pyrethrum extract (50 mg) was added. The
6 suspension was sonicated for 5 min, at an ultrasound power of 200 W and at 25°C, and was left
7 under stirring for 18 h at room temperature. After this time, the dispersion was centrifuged and the
8 obtained powder was washed with water, until the unreacted pyrethrins were removed and, then
9 dried at r.t. The loading and loading efficiencies (LD% and LE%, respectively) were calculated as
10 follows by using Eq. 1 and 2.

11 Ca. 2 mg of Hal/PE nanomaterial were carefully weighed and washed with four portions of hexane
12 (2 mL each). The organic solution obtained was then analyzed by GC-MS.

$$13 \quad \text{LD\%} = \frac{m_{\text{pyrethrins_Hal}}}{m_{\text{Hal}} + m_{\text{pyrethrins_Hal}}} \times 100 \quad (\text{Eq. 1})$$

$$14 \quad \text{LE\%} = \frac{m_{\text{pyrethrins_Hal}}}{m_{\text{pyrethrins_TOT}}} \times 100 \quad (\text{Eq. 2})$$

15 where $m_{\text{pyrethrins_Hal}}$ and $m_{\text{pyrethrins_TOT}}$ are the amount (express in mg) of the pyrethrins loaded on Hal
16 and the total feed one, respectively and m_{Hal} is the mass of Hal (express in mg).

17 **2.2 Photodegradation studies in solution**

18 Different aliquots of an aqueous suspension of PESTANALTM (0.03 mg/mL) or aqueous dispersion
19 of Hal/PE (0.25 mg/mL) were placed under UV lamp (254 nm, 32 W, with a distance of 15 cm, at
20 room temperature). The sample into a closed UV quartz cuvette (total volume 2 mL) was exposed
21 to the lamp. For PESTANALTM, after predetermined times, the aqueous solutions were analysed
22 and the percentage of decomposed pyrethrins was determined by spectrophotometric measurements
23 estimating the disappearance of the maximum absorption band of pyrethrum (226 nm) and by
24 UPLC-ESI-MS.

1 In the case of Hal/PE dispersions, 2 samples were irradiated at 254 nm for 1 h and 4 h respectively.
2 After the set time, both samples were stirred vigorously for four minutes with 2 mL of hexane, to
3 extract residual pyrethrins. UV spectra were recorded on the obtained hexane solutions. A
4 comparison with a non-irradiated sample was carried out. All experiments were carried out in
5 duplicate at 25°C.

6 **2.3 Photodegradation studies in solid state**

7 Irradiation experiments were performed by using a UV-*vis* medium-pressure Mercury 125 W lamp.
8 Each sample was inserted into a rectangular UV quartz cuvette (1 × 1 cm, total volume 3 mL) by
9 accurate deposition over a selected wall of the cell. In particular, 2.9 mg of Hal/PE nanomaterial
10 were uniformly distributed, while a thin film of PESTANALTM was prepared upon deposition of 4
11 × 25 µL of a 3 mg/mL hexane solution quickly drying at room temperature. Each cuvette,
12 appropriately closed, was placed with the selected wall exposed to the lamp at a distance of 7 cm
13 and inclined by 45°. The temperature of samples during irradiation did not exceed 22°C. After the
14 desired irradiation time the PESTANALTM film was dissolved in 6.5 mL of hexane, while the
15 Hal/PE nanomaterial was dispersed in 6.5 mL of hexane and vigorously stirred for 30 minutes for
16 pyrethrin extraction: the supernatant was recovered upon ultracentrifugation at 5000 rpm for 10
17 min. All the obtained hexane solutions were preliminarily analyzed by CD/UV spectroscopy by
18 using a 1 cm quartz cell. For ESI MS analysis a good correlating calibration curve was obtained in
19 methanol for PESTANALTM concentrations ranging from 0.189 to 0.027 mg/mL and by using
20 glucose 0.14 mM as an internal standard. Thus, for ESI MS analysis all hexane samples were dried
21 under nitrogen and dissolved in the appropriate amount of methanol in the presence of glucose 0.14
22 mM. For each experiment, two replications were carried out

23 **2.4 Kinetic Release**

24 The release of pyrethrins from Hal/PE nanomaterial was performed as follows: 9.5 mg of the
25 sample were transferred into a dialysis membrane (Medicell International Ltd MWCO 12–14,000
26 with a diameter of 21.5 mm) and wet with 0.5 mL of water. The membrane was then put in a round

1 bottom flask containing 4.5 mL of water at 25°C under constant stirring. At fixed times, 0.5 mL of
2 the release medium was withdrawn and analyzed. To keep constant the volume of the release
3 medium 0.5 mL of water was added each time to replace the withdrawn one. Each sample was
4 extracted three times with 0.5 mL of hexane, dried under nitrogen and dissolved in 0.5 mL of
5 hexane. The pyrethrins concentration in the solution was determined by UV-vis spectrophotometry
6 using the Lambert-Beer law, measuring the absorbance at 224 nm, by assuming that at each time all
7 pyrethrum components were proportionally released.

8 Total amount of pyrethrins released (F_t) was calculated as follows:

$$9 \quad F_t = V_m C_t + \sum_{i=0}^{t-1} V_a C_i \quad (\text{Eq. 3})$$

10 where V_m and C_t are the volume and concentration of pyrethrum at time t . V_a is the volume of the
11 sample and C_i is pyrethrins concentration at time i ($i < t$).

12 **2.6 Activity Hal/PE on insects**

13 **2.6.1 Insects**

14 *T. molitor* specimens were kept in plastic boxes (30 × 18 × 10 cm) at 25 ± 1°C, 65 ± 5% R.H. and
15 L16:D8 photoperiod. Food consisted of cracker pieces, corn flour, and carrots as a source of
16 hydration. To obtain adults of similar age, *T. molitor* pupae were transferred from the colony to a
17 separate box provided with food. After about one week, the newly emerged adults were collected
18 and used in trials.

19 *G. mellonella* larvae were reared in plastic boxes (24 × 12 × 8 cm) at 30 ± 1°C, 65 ± 5% R.H. and
20 L0:D24 photoperiod and fed on an artificial diet (Dindo et al., 2003; Marchetti et al., 2009). Young
21 larvae (in the 3rd or 4th instar, approximately 1-1.5 cm long) were used in the trials.

22 Laboratory colonies of *T. molitor* and *G. mellonella* were maintained at the Department of
23 Agricultural and Food Sciences of University of Bologna.

24 **2.6.2 Toxic effects of Hal/PESTANAL™ vs. PESTANAL™ and commercial pyrethrum** 25 **extract**

1 The efficacy of Hal/PE nanomaterial (1 mg/mL corresponding to 0.02 mg/mL in pyrethrins) and
2 commercial pesticides (Pyrethrum Actigreen 0.04 mg/mL, PESTANAL™ 0.04 mg/mL) was tested
3 by contact bioassays, separately for *T. molitor* and *G. mellonella*. Distilled water was maintained as
4 control.

5 In each treatment 1 mL liquid product was pipetted onto the bottom of a 6 cm diameter plastic petri
6 dish. Insects (either *T. molitor* adults or *G. mellonella* larvae) were then placed in the dish which
7 was eventually covered with its lid.

8 For each insect species, 6 replicates per treatment were set up, each consisting of one petri dish with
9 5 individuals, with 90 insects in total per species (30 per treatment). All petri dishes were
10 maintained in an incubator at $25 \pm 1^\circ\text{C}$, $65 \pm 5\%$ R.H., and L16:D8 photoperiod.

11 In all treatments, the number of dead individuals was counted at three check times, i.e., 24 h, 72 h,
12 and 168 h (1 week) post insect exposure. Sub-lethal effects (i.e., the number of individuals showing
13 low mobility) were also detected at each check time.

14 *Parameters*

15 In both trials, for each insect species and at each check time, mortality (%) was calculated over the
16 original insect number ($n = 5$ per replicate per treatment). At each check time, the number of
17 survived healthy individuals and of low mobile individuals was also calculated. For each treatment,
18 these data were pooled for all replicates.

19 *Statistical analysis*

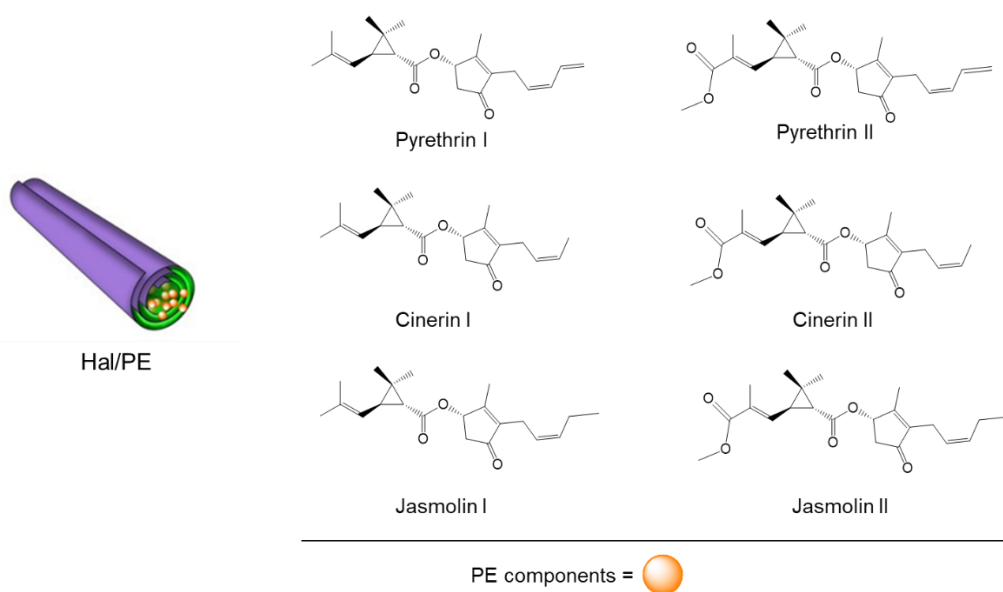
20 Percent mortalities were analyzed by one-way ANOVA or by Kruskal-Wallis test, respectively
21 followed by Tukey's HSD test or non- parametric multi-comparisons when a significant difference
22 occurred. The Kruskal-Wallis test was used in case of variance heterogeneity (p-value > 0.05,
23 Levene test). Prior to analysis, the percentage values were angular transformed by means of
24 Freeman-Tukey double arcsine transformation for small samples ($n < 50$) (MOSTELLER and
25 YOUTZ, 1961). To compare low mobile vs. healthy alive individuals, 2×2 contingency tables

1 were created, using Yates correction for small numbers (<100). Data analysis was performed with
2 the statistical program STATISTICA v.10.0 (StatSoft, Tulsa, OK, USA).

3

4 **3. Results and Discussion**

5 Pyrethrum extract is a mixture of different components, where the main ones are the so-called
6 pyrethrins (Pyrethrin I, Pyrethrin II, Cinerin I, Cinerin II, Jasmolin I, and Jasmolin II). Based on the
7 physico-chemical properties of both Hal and PE components, the interaction between them can
8 occur by electrostatic attraction interactions, hydrophobic effects, and hydrogen bonding occurring
9 between the Al-OH groups present at the inner surface of Hal and the oxygen atoms of the PE
10 components (Figure 1).

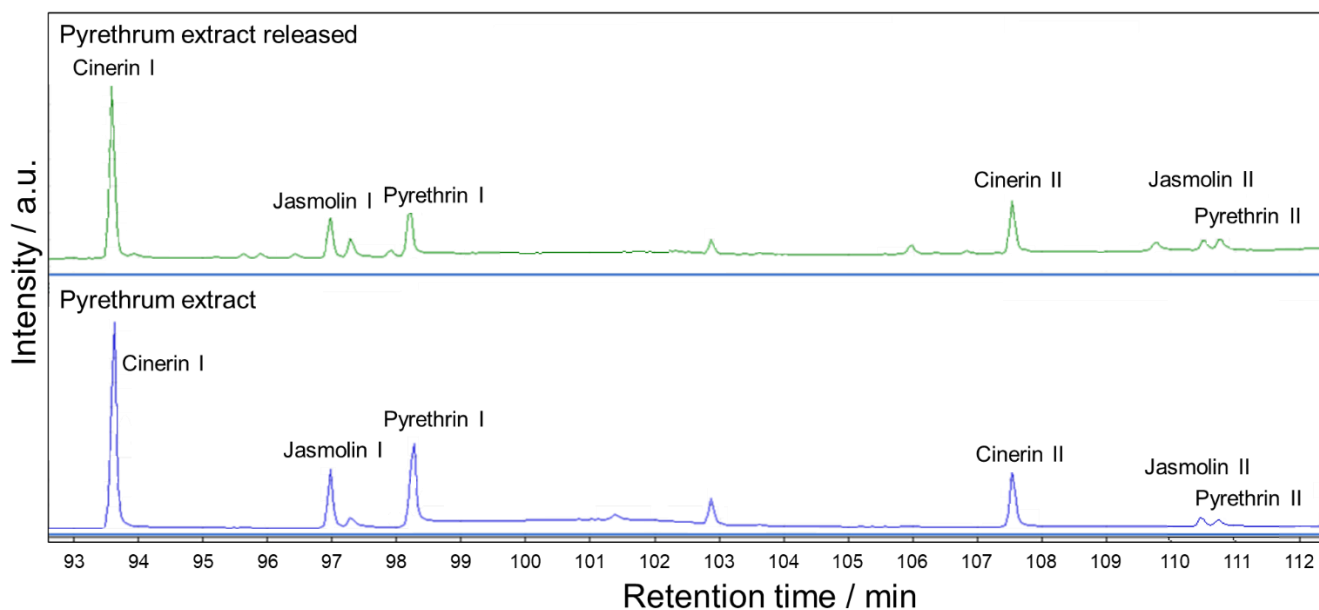


11

12 **Figure 1.** Schematic representation of the Hal/PE nanomaterial synthesized.

13 The main components of pyrethrum extract from PESTANALTM (PE) were loaded into Hal by
14 adopting a procedure reported elsewhere (Massaro et al., 2022b; Riela et al., 2021). After the work-
15 up the amount of extract loaded on Hal, was estimated by GC-MS analysis and was found to be ca.
16 5.5 wt% with an entrapment efficiency of ca. 30%.

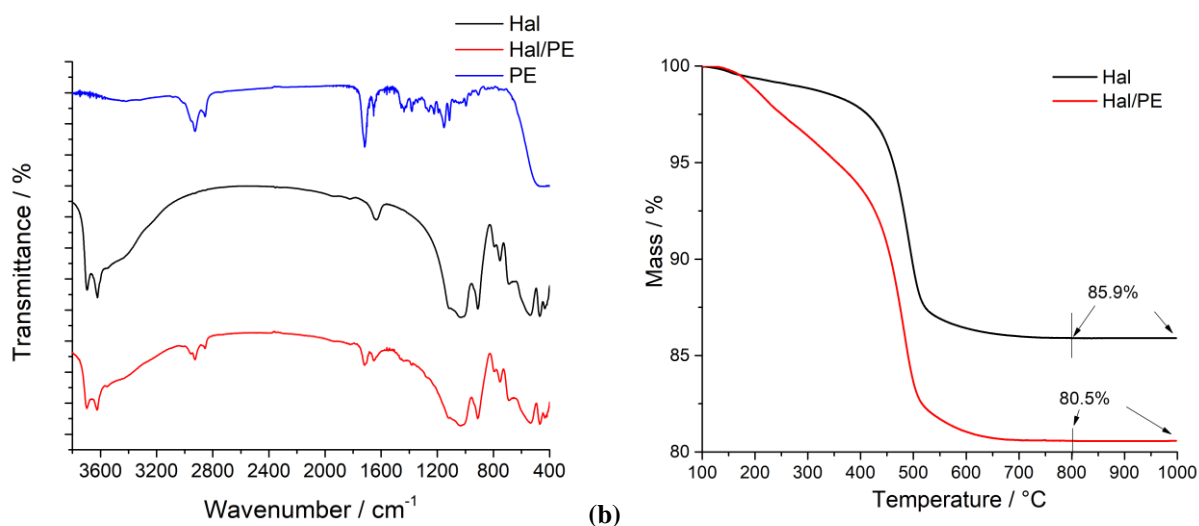
17 Noteworthy, the GC-MS analysis highlighted that the main components of pyrethrum extract are
18 loaded onto Hal in the same molar ratios as present in the extract itself (Figure 2).



1

2 **Figure 2.** GC-MS analysis of the commercial pyrethrum extract (PESTANAL™) and the pyrethrum extracted
 3 from Hal/PE nanomaterial.

4 The successful loading was verified by FT-IR spectroscopy and thermogravimetric analysis. The
 5 FT-IR spectrum of the Hal/PE nanomaterial shows the typical vibration bands of HNTs, namely the
 6 bands at 3622 and 3493 cm^{-1} corresponding to O–H stretching vibrations of buried hydroxyl groups
 7 and inner surface hydroxyl groups, respectively, and the bands at 3484 and 1645 cm^{-1} assigned to
 8 O–H stretching vibrations of adsorbed water. Besides these, some additional bands, related to the
 9 PE, are present corroborating the loading of pyrethrum components (Pajnik et al., 2018) (Figure 3a).
 10 In particular, the bands related to the C–H asymmetric and symmetric stretching vibrations of
 11 pyrethrins at ca. 2900 and 2850 cm^{-1} are clearly observable together with the band at ca. 1715 cm^{-1}
 12 ascribed to the stretching of C=O group, and the band at ca. 1500 cm^{-1} attributable to the stretching
 13 of C=C group of pyrethrins skeleton. Finally, the band at ca. 1200 cm^{-1} was due to the stretching
 14 vibration of C–O groups.



1 (a) (b)

2 **Figure 3.** (a) FT-IR spectra pristine Hal, PE, and Hal/PE nanomaterial and (b) TGA curves of pristine Hal and Hal/PE

3 nanomaterial.

4 Pristine pyrethrum extract totally decomposes in a multi-step process occurring in a temperature

5 range between 200 and 450°C (Figure S1). As results from the comparison of the TGA curves

6 (Figure 3b) the pristine Hal and Hal/PE nanomaterial show different thermal stability under air

7 atmosphere by increasing the temperature from 100°C up to ca 600°C. In both cases, a negligible

8 weight loss occurs between rt up to 100°C due to removal of some physically adsorbed water (this

9 step was omitted in the shown figure) The Hal exhibits relatively good stability up to ca 350°C,

10 then between 350 and 600°C, suddenly lose weight until reaching a stable value corresponding to

11 ca 86 wt% of the initial mass.

12 Accordingly, with the literature (Massaro et al., 2020) a progressive dehydroxylation of the

13 structural Al-OH groups of the Hal occurs until complete structural modification, at around 550-

14 600°C. The Hal/PE nanomaterial according to the presence of pyrethrum, an organic ester, starts to

15 be degraded earlier than the pristine Hal, starting from ~ 200°C, losing weight increasingly by

16 increasing the temperature up to 600°C, up to stable mass corresponding to 80.5 wt%.

17 The amount of PE loaded into Hal was calculated by using the rule of the mixture by considering

18 the mass losses at 25–150°C (ML₁₅₀) and the residual masses at 800°C (MR₈₀₀) for the pristine

19 components (Hal and PE) and the Hal/PE nanomaterial (Lisuzzo et al., 2020; Lisuzzo et al., 2019).

1 In Table 1 the ML_{150} and MR_{800} values of PE, Hal and Hal/PE nanomaterial are reported. From
2 these data it was possible to calculate the loading efficiency that was of ca. 5.5 wt% in agreement
3 with GC-MS analysis.

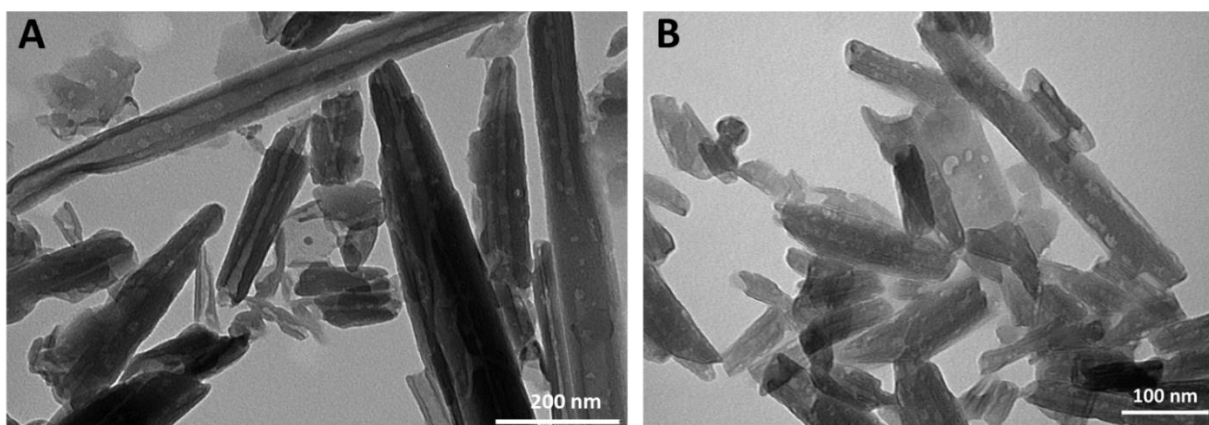
4 Details on the calculation of the loading amounts of PE into Hal are shown in SI.

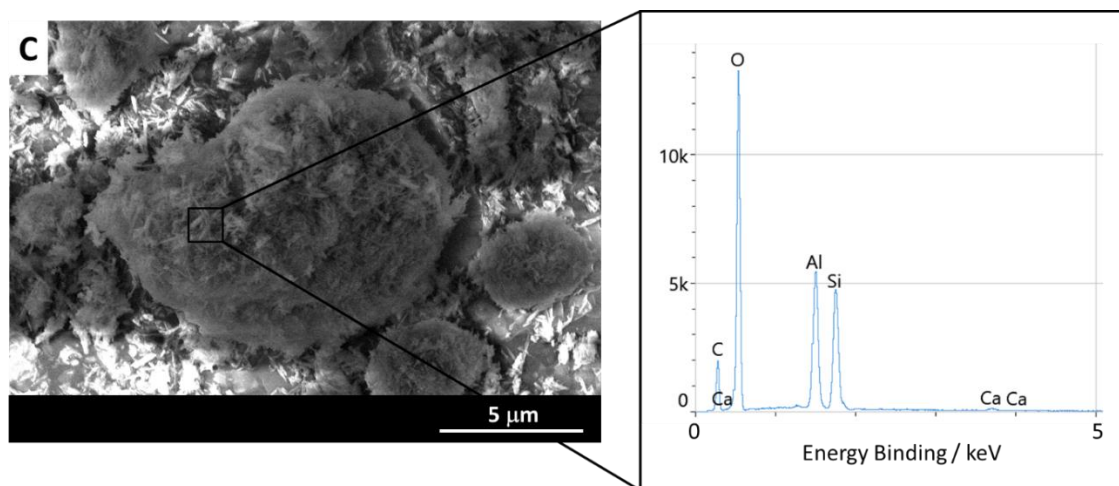
5 **Table 1.** Thermogravimetric parameters for PE, Hal and Hal/PE nanomaterial.

	ML_{150} / wt%	MR_{800} / wt%	MD_{800} / wt% ^a
PE	0	0	100
Hal	3.0	85.9	11.2
Hal/PE	3.0	80.5	16.5

6 ^a for the calculation of MD_{800} (degraded matter) values see SI.

7 The morphology of the Hal/PE nanomaterial was imaged by TEM and SEM measurements. As it is
8 possible to note, after the extract loading the tubular morphology of Hal is preserved (Figure 4A).
9 However, TEM image shows that the Hal lumen is not apparent in all its length confirming the
10 successful loading (Figure 4B). Furthermore, the EDX analysis showed that beside the presence of
11 the Al, Si and O atoms related to the inorganic carrier, C atoms are present in the nanomaterial,
12 further confirming the loading (Figure 4C).





1

2 **Figure 4.** (A-B) TEM images of (A) raw Hal and (B) Hal/PE nanomaterial, (C) SEM image of Hal/PE nanomaterial,
 3 the insert shows the EDX analysis on the selected area.

4 3.3 Kinetic release

5 To evaluate the performances of the Hal/PE nanomaterial for potential application as a pesticide,
 6 the kinetic release of the pyrethrins, constituents of PE extract, from Hal/PE nanomaterial was
 7 evaluated by the dialysis bag method, using the same conditions adopted for the *in vivo* experiments
 8 (water, 25°C) and the obtained kinetic data are shown in Figure 5. In the same conditions, the PE
 9 spreading through the membrane is hampered by its very low solubility in water.

10 As it is possible to notice from Figure 5, a slow release occurred in the first 300 minutes after which
 11 a plateau is observed. Indeed, only 14 wt% of the total amount of pyrethrins loaded into Hal was
 12 released after 48 h indicating that the Hal could act as a reservoir for a slow release of pesticides.

13 To better understand the release mechanism, the kinetic data were analyzed by the first order and
 14 power fit models. The mathematical analysis showed that the kinetic data are better fitted by the
 15 first order model ($M_{\infty} = 9.7 \pm 0.3$ mg/mL, $k = 0.014 \pm 0.001$ min⁻¹, $R^2 = 0.9887$), indicating a simple
 16 diffusion of the pyrethrins from Hal lumen.

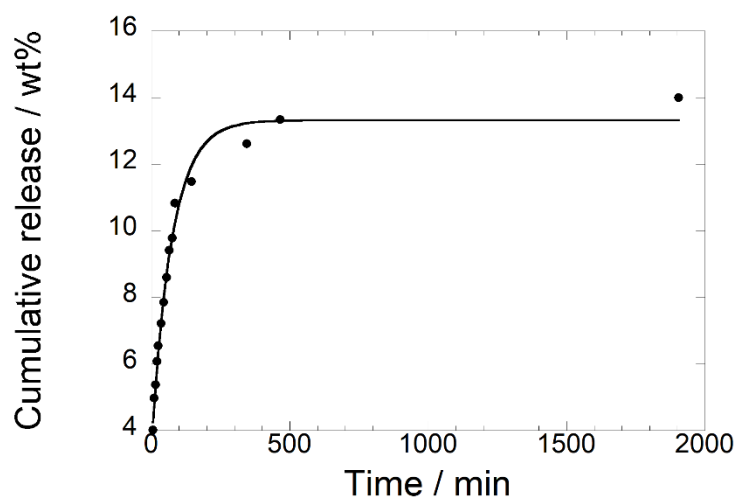


Figure 5. Kinetic release of pyrethrins from Hal/PE nanomaterial at 25°C in water.

3.4 Photodegradation studies

In the last years, pyrethrins were largely replaced by their synthetic derivatives, pyrethroids. However, due to their harmful effects on various species, the use of pyrethrins should again take the lead. Pyrethrins indeed possess, among other interesting properties, less negative effects on the environment compared to synthetic ones (Chrustek et al., 2018).

One of the major drawbacks of the agricultural use of pyrethrum extract is its rapid photodegradation under UV light. Therefore, the assessment of the photostability of the obtained nanomaterial in solution and in solid state is relevant for its possible future use as a pesticide indoors and outdoors. In this section, we report on the photostability of the Hal/PE nanomaterial in comparison with the pyrethrum extract both in solution and in a solid state.

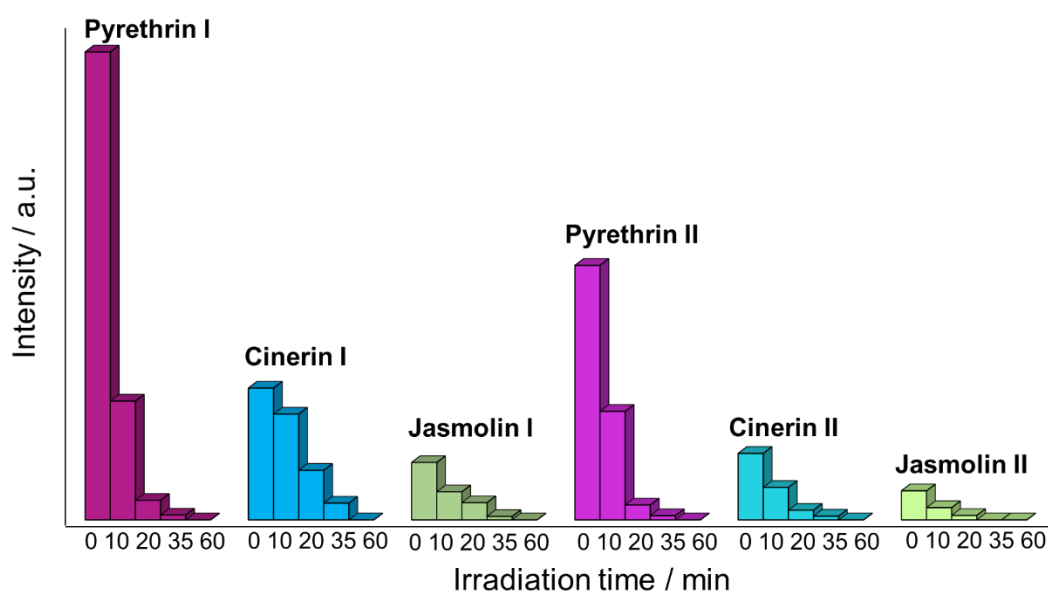
3.4.1 Photostability in solution

Inspired by recent photochemical studies performed on pyrethrin formulations (Zhang et al., 2019), we firstly tested the photostability of PESTANALTM and Hal/PE nanomaterial in water under UV irradiation at 254 nm. The amount of pyrethrins photodegraded as a function of time was determined by UV-*vis* measurements. As far as is regarding the pyrethrum extract, pyrethrin degradation followed a pseudo-first-order model ($R^2 = 0.994$), the process in aqueous solution was

1 very rapid ($k_{obs} = 0.07 \text{ min}^{-1}$), with a degradation rate higher than 50% after about 10 min (Figure
2 S2).

3 The decay of each component of the PESTANAL™ was also assessed by UPLC-ESI-MS. As found
4 by UV-vis measurements, pyrethrins were almost totally photodegraded within 20 min of irradiation
5 (Figure 6).

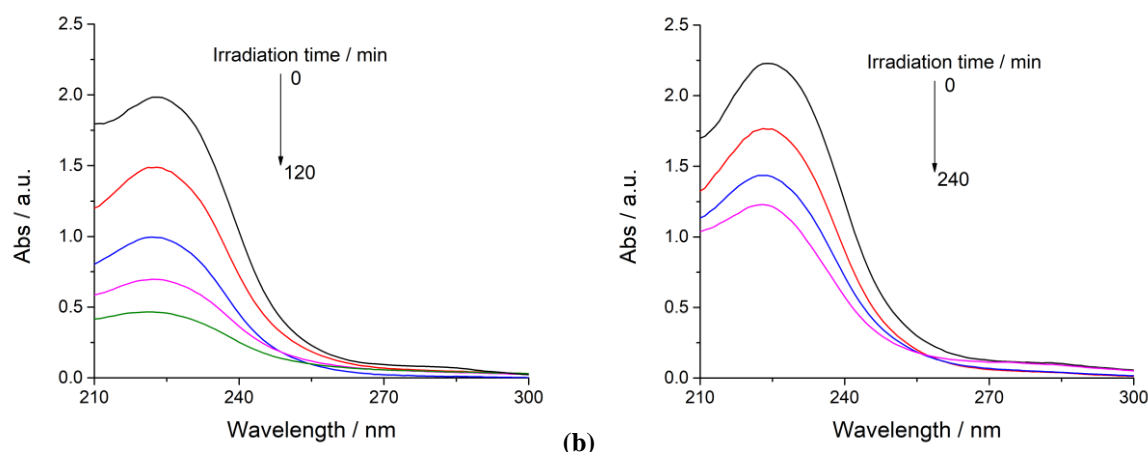
6 Noteworthy, conversely to experimental results obtained for the PESTANAL™, after the pyrethrins
7 loading into Hal, no photodegradation occurred in the first 60 min. Indeed, UV spectra obtained by
8 a Hal/PE water dispersion before and after irradiation (Figure S3) show that signal intensity, related
9 to pyrethrin content, reduced less than 10% after 60 min and about 15% after 4 h, presumably due
10 to molecular release in water and consequent photodegradation. On the other hand, almost 90% of
11 initial pyrethrins were found active after 60 min, as presumably still included in Hal/PE and then
12 photoprotected. This result agrees with the very slow release of pyrethrins from Hal/PE
13 nanomaterial in aqueous medium described above, and in parallel highlights the shielding ability of
14 halloysite against the short-wavelength UV radiations (UV-C range). Similar results were found by
15 Liu et al. which demonstrated that the UV reflection by the Hal layered silicate structures greatly
16 improved the effective UV protection time of the pesticide chlorpyrifos (Qin et al., 2022).



17 **Figure 6.** Photodegradation of PE components (Pyrethrin I and II, Cinerin I and II, Jasmolin I and II) in water as a
18 function of time.
19

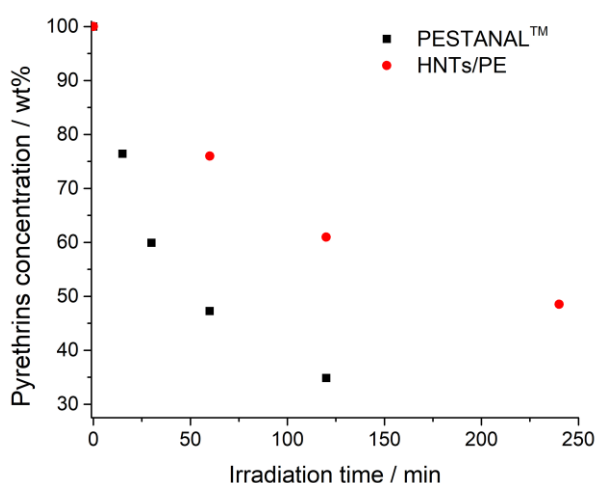
1 3.4.2 Photostability in solid state

2 A deeper investigation on photostability of Hal/PE was performed on the solid state. In this case a
3 mercury lamp, that includes UV-B and UV-A emissions, was used for irradiation experiments to
4 better mimic natural UV light. The Hal/PE nanomaterial and a PESTANALTM film, used as control,
5 were subjected to UV light irradiation, and consequent photodegradation was monitored by UV/CD
6 spectroscopy and ESI-MS. In accordance with pyrethrin maximum absorbance (Head, 1973) UV
7 spectra exhibited a maximum at *ca.* 225 nm (Figure 7) and due to the chirality of the six
8 components - related to their insecticidal activity (Kawamoto et al., 2020), the same spectral region
9 was dominated by a negative CD band. A positive CD signal was also detected between 300 and
10 350 nm (Figure S4) (Crombie, 1999). Upon UV irradiation of PESTANALTM a gradual lowering of
11 either UV or CD signals diagnostic of progressive PE components photodecomposition was
12 observed. In particular, after two hours UV/CD absorption maxima were reduced to *ca.* 30%.
13 Conversely, in the case of Hal/PE nanomaterial more than 60% of both UV and CD signal
14 intensities were preserved after 4 hours of irradiation (Figures 7 and S4). These preliminary results
15 suggested that PE lifetime under ultraviolet light exposition was significantly enhanced by loading
16 into Hal.



17 (a) (b)
18 **Figure 7.** UV spectra recorded on solutions obtained by treating with a volume of hexane (a) the PESTANALTM film
19 and (b) the Hal/PE nanomaterial after different irradiation times with ultraviolet light.
20

1 UV/CD data were corroborated by ESI MS analysis that provided identification and quantification
2 of the individual components (Figures S5 and S6). The obtained plots showing the percentage of
3 remaining pyrethrins vs the irradiation time are depicted in Figure 8. For the PESTANAL™ sample,
4 the initial pyrethrin content reduced to 47% after one hour and to 35% after two hours. Conversely,
5 after 2 h irradiation of the Hal/PE nanomaterial, ca. 61% of the total amount of PE into the lumen
6 was recovered and noteworthy, more than 50% of the initial amount was still present after four
7 hours, as testified by the analysis of hexane solution of the PE extracted from Hal/PE nanomaterial
8 (Figure 8). Collected data highlighted for the Hal/PE nanomaterial a significant improved
9 photostability in respect to PESTANAL™. Weakening of UV radiation interacting with the Hal
10 walls accompanied by possible absorption of the lowest wavelengths shall be responsible for the
11 observed protective effect (Cavallaro et al., 2020).



12
13 **Figure 8.** Percentage of remaining pyrethrins (wt%) after different UV irradiation times of the Hal/PE nanomaterial and
14 the PESTANAL™ film.

15 **3.5 Toxic activity of Hal/PE nanomaterial vs *T. molitor* and *G. mellonella*.**

16 It is known that pyrethrins pass through the exoskeleton of insect chitin by contact diffusion and
17 cause depolarization by preventing the closure of the sodium channels of the cell membrane in
18 nerve and muscle cells (Kaya, 2020).

19 As proof of concept, the toxic effects of the developed Hal/PE nanomaterial were tested *in vivo* on
20 selected target insects, i.e., adults of the yellow mealworm *T. molitor* and young larvae of the
21 greater wax moth *G. mellonella* in terms of mortality and sub-lethal effects induced in the target

1 insects. Hal did not show any toxic effects on the target insects in the experimental conditions used.
2 Firstly, the Hal/PE nanomaterial and commercial pesticides were used at the concentration
3 recommended for field application (2 mg/mL, corresponding to 0.04 mg/mL of PE components and
4 0.04 mg/mL for Hal/PE nanomaterial and commercial pyrethrins, respectively). In these conditions,
5 no differences between the two treatments were observed. Therefore, we tested the Hal/PE
6 nanomaterial at a concentration of 1 mg/mL, corresponding to 0.02 mg/mL of PE components.
7 Table 2 reports the results obtained in terms of mortality of the two insects after different exposure
8 times to the Hal/PE nanomaterial and PESTANALTM (similar results were obtained in the case of
9 the Pyrethrum Actigreen[®]).

10 As it is possible to observe, after 24 h of exposure, the mortality of *G. melonella* was ca. 97-100%
11 for both Hal/PE nanomaterial and commercial pesticide. At each check time, mortality in the
12 negative control (distilled water) was extremely low (Table 2). This result was noteworthy, since
13 the presence of Hal allowed to halve the amount of pyrethrins used without differences in the
14 pesticide activity, in comparison to the concentration commonly recommended for this kind of
15 treatments. This finding could be a consequence of the presence of Hal which both protect the
16 pyrethrins molecules from photodegradation and hydrolysis and at the same time, it enhances their
17 aqueous solubility improving the pesticide activity.

18 Regarding the yellow mealworm adults, the results were quite different. In this case, indeed, no
19 significant mortality of the insects was observed in the first 48 h of all treatments. After 1 week of
20 exposure, eventually, ca. 35% of mealworms were found dead without any appreciable difference in
21 each case investigated. These results could be explained by considering the different characteristics
22 of the two insects. Natural pyrethrins, like their synthetic derivatives pyrethroids, do not act as a
23 repellent towards insects, but, rather, as contact insecticide, causing nervous system toxicity,
24 leading to death or “knockdown” of the insect. Therefore, their action on *T. molitor* adults, which
25 are hard-shelled beetle, could be limited by their scarce uptake.

26

1 **Table 2.** Mortality (%) of *T. molitor* adults and *G. mellonella* larvae observed in the first trial. Individuals were
 2 treated by contact with Hal/PE nanomaterial (1 mg/mL, corresponding to a pyrethrin concentration of 0.02
 3 mg/mL), commercial pesticides (0.04 mg/mL) or negative control (water). Mean values (\pm SE) at each check time
 4 (24 h, 72 h, and 1 week post insect exposure) are reported. Number of replicates = 6. At each time, % mortality
 5 was calculated on the original number of individuals (5 per treatment per replicate= 90 individuals per insect
 6 species). Means in a column followed by the same letter are not significantly different ($P < 0.05$) by one-way
 7 analysis of variance followed by Tukey's HSD test (yellow mealworms - 72 h) or Kruskal-Wallis test followed
 8 by non-parametric multi-comparisons (other parameters).

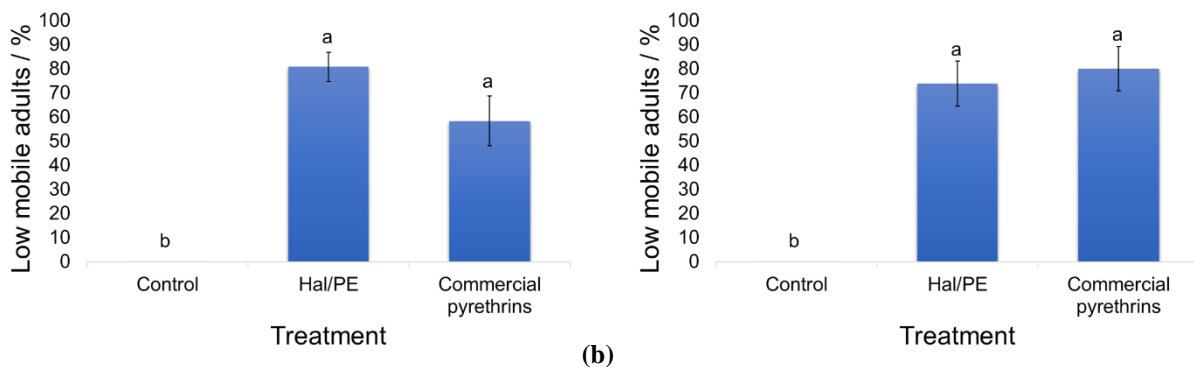
<i>G. mellonella</i>			
Treatment	Mortality (%) (24 h)	Mortality (%) (72 h)	Mortality (%) (1 week)
Hal/PE	100 a	100 a	100 a
Commercial Pesticides*	97 \pm 3 a	100 a	100 a
Distilled water	0 b	0 b	3 \pm 3 b
H (N)	15.6 (18)	17 (18)	16.6 (18)
P	0.0004	0.0002	0.0002

9

<i>T. molitor</i>			
Treatment	Mortality (%) (24 h)	Mortality (%) (72 h)	Mortality (%) (1 week)
Hal/PE	13 \pm 10 a	23 \pm 16 a	29 \pm 10 a
Commercial Pesticides ^a	17 \pm 10 a	33 \pm 13 a	36 \pm 9 a
Distilled water	0 a	3 \pm 3 a	15 \pm 3 b
F (df)		1.7 (2.15)	
H (N)	3.5 (18)		7.0 (18)
P	0.175	0.208	0.03

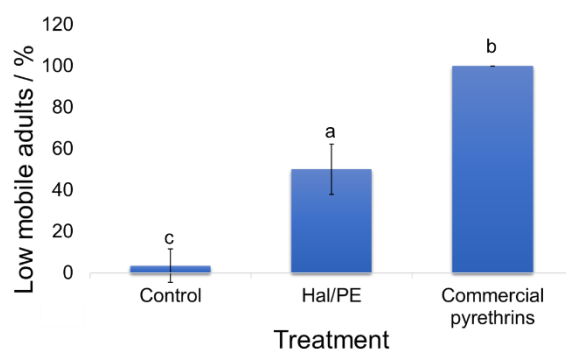
1 *PESTANAL™ or Pyrethrum Actigreen®.

2 However, by checking the mobility of the survived individuals at each exposure time investigated, it
3 was found that the treatment with both Hal/PE and PESTANAL™ resulted in significantly lower
4 mobility of survived individuals, in comparison to the negative control (24 h: $\chi^2 = 35.40$, $df = 1$, P
5 < 0.0001 for Hal/PE nanomaterial; $\chi^2 = 20.69$, $df = 1$, $P < 0.0001$ for PESTANAL™; 72 h: $\chi^2 =$
6 28.58 , $df = 1$, $P < 0.0001$ for Hal/PE nanomaterial; $\chi^2 = 30.91$, $df = 1$, $P < 0.0001$ for
7 PESTANAL™; 168: $\chi^2 = 11.72$, $df = 1$, $P = 0.0006$ for Hal/PE nanomaterial; $\chi^2 = 37.99$, $df = 1$, P
8 < 0.0001 for PESTANAL™) (Figure 9). No significant difference was found between the two
9 treatments both at 24 h ($\chi^2 = 2.02$, $df = 1$, $P = 0.1554$) and 72 h ($\chi^2 = 0.01$, $df = 1$, $P = 0.9129$) after
10 exposure. Surprisingly, after one week of treatment with the Hal/ PE nanomaterial the number of
11 low mobile adults was lower than at 72 h (Figure 9b and 9c). This result suggests that the survived,
12 low mobile, yellow mealworm adults overtime showed some capacity to retrieve. Since it is known
13 that low dosages of pyrethrins led to a recovery of the insects from the exposure (Soderlund, 2010),
14 the experimental results are in agreement with the slow release of PE components from Hal/PE
15 nanomaterial. After 1 week treatment with PESTANAL™, indeed, the number of low mobile adults
16 (100%) was significantly higher compared with Hal/PE nanomaterial ($\chi^2 = 8.97$, $df = 1$, $P = 0.0027$)
17 (Figure 9c).



18 (a)

(b)



(c)

1

2 **Figure 9.** Sub-lethal effects of Hal/PE nanomaterial and PESTANALTM on *T. molitor* adults in contact
 3 bioassays. Columns indicate the percentage (\pm SE) of low mobile adults, calculated over the total number of
 4 alive adults, at three check times: (a) 24 h, (b) 72 h and (c) 1 week after placing the insects in the treated
 5 petri dishes. In each graph, different letters above the columns indicate significant differences in the
 6 percentage of low mobile adults as determined by 2×2 contingency tables, created to test any possible
 7 combination of treatments. At each check time, the numbers of alive adults for Hal/PE nanomaterial,
 8 PESTANALTM and water were 26, 24 and 30 (24 h); 23, 20 and 29 (72 h) and 18, 17 and 29 (1 week),
 9 respectively.

10 4. Conclusions

11 In summary, a novel system for the controlled release and UV protection of pyrethrum extract
 12 components was obtained using Hal as nanocontainer. The combination of the eco-friendly clay
 13 mineral and pyrethrum improves the photochemical stability of the extract, both in solution and in
 14 the solid state, enhances its water solubility, and pesticidal activity in the *in vivo* models considered.
 15 Hal offers some advantages over other nanomaterials already reported for these purposes (Wibowo
 16 et al., 2014; Yan et al., 2019), because of its low-cost, large, and natural availability and its intrinsic
 17 properties such as photoprotective effects. Similarly to other clay minerals (da Rocha et al., 2019),
 18 indeed, Hal chemical and structural features render it a good photoprotective material for different
 19 organic molecules. Specifically, in this study, it was demonstrated that the encapsulation of
 20 pyrethrum extract components into Hal lumen, prevents their photodegradation. Irradiation of
 21 Hal/PE nanomaterial both in solution and in the solid state showed that the clay protected the PE

1 from UV light for at least 4 h, conversely, the pristine PE was fully photodegraded after 1 h of
2 irradiation.

3 *In vivo* tests performed on *G. mellonella* and *T. molitor* showed that the pyrethrins loaded onto Hal
4 possess the same activity as standard pyrethrins, at half dose compared to them, in line with the
5 observations performed in previous studies (Chaud et al., 2021). The result suggests the possibility
6 to reduce the amount of insecticide to control target insect pests with no decrease in efficacy, with
7 an advantage for the environment. Wax moth larvae were more susceptible than yellow mealworm
8 adults. It is, however, well-known that the latter are not very sensitive to pyrethrins (Pedersen et al.,
9 2020), thus they can be used to evaluate the performances of the nanomaterial synthesized on
10 different pests with a different sensibility to the treatment. Therefore, the controlled release
11 nanoformulation described in this work has great potential in the development of low-cost and
12 environmentally friendly formulations of botanical insecticides for sustainable pest control.

13 **Acknowledgments**

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