

1 **Title:** Fish assemblages cope with ocean acidification in a shallow volcanic CO₂ vent benefiting from
2 an adjacent recovery area
3

4 **Authors:**

5 Alice Mirasole (1, 2), Geraldina Signa (3), Paola Gianguzza (1), Chiara Bonaviri (1), Antonio
6 Mazzola (1, 3), Salvatrice Vizzini (1, 3).
7

8 **Affiliation:**

9 (1) University of Palermo, Department of Earth and Marine Sciences, via Archirafi 18, 90123
10 Palermo, Italy

11 (2) Stazione Zoologica Anton Dohrn, Department of Integrative Marine Ecology, Villa Dohrn-
12 Benthic Ecology Center, Punta San Pietro s/n, 80077, Ischia, Napoli, Italy

13 (3) CoNISMa, National Inter-University Consortium for Marine Science, Piazzale Flaminio 9, 00196
14 Roma, Italy
15

16 **Authors ORCID iD:**

17 Alice Mirasole - 0000-0003-2517-9548

18 Geraldina Signa - 0000-0003-2171-3299

19 Paola Gianguzza - 0000-0002-7715-8637

20 Chiara Bonaviri - 0000-0002-3897-2361

21 Antonio Mazzola - 0000-0001-8433-278X

22 Salvatrice Vizzini - 0000-0002-7127-7555
23

24 Corresponding authors: Alice Mirasole

25 E-mail address: alice.mirasole@szn.it
26

27 **Abstract**

28 Shallow CO₂ vents are used to test ecological hypotheses about the effects of ocean acidification
29 (OA). Here, we study fish assemblages associated to *Cymodocea nodosa* meadows, long-term
30 exposed to high pCO₂/low pH conditions. Using underwater visual census, we assessed fish
31 community structure and biodiversity in a low pH site, a close control site and a far control site,
32 hypothesising a decline in biodiversity and a homogenisation of fish assemblages under OA
33 conditions. Our findings revealed that fish diversity did not show a univocal spatial pattern, or even
34 significant relationships with pH, but correlated with seagrass leaf canopy. Among-site similarity was
35 found in juveniles' abundance, contrary to the forecasted impacts of OA on early life stages. Despite
36 this, pH was an important driver in structuring fish assemblages, which however showed high
37 similarity between the low pH site and the close control. This unexpected pattern may represent a
38 combined response of fish mobility, enhanced food resources in the acidified site, and a 'recovery
39 area' effect of the adjacent control site.

40

41 **Keywords:** Mediterranean Sea, CO₂ seeps, pH, Fish, Seagrass, *Cymodocea nodosa*, Biodiversity,
42 Community structure, Underwater visual census, Juveniles

43

44 **Introduction**

45 The increase in anthropogenic CO₂ absorbed by the ocean and the consequent pH reduction, known
46 as ocean acidification (OA), are causing changes in the carbon chemistry balance with direct and
47 indirect effects on marine ecosystems and biodiversity (Barry et al. 2011, Cheung et al. 2009,
48 Fabricius et al. 2011, Foo et al., 2018, Nagelkerken & Connell, 2015). OA can exert negative direct
49 effects on organisms, especially calcifying species (i.e. algae, corals, mollusks and echinoderms)
50 impairing survival, calcification, growth, development, reproduction and abundance (Kroeker et al.
51 2013). More difficult is to disentangle the indirect effects of OA, which are as important as direct
52 ones to forecast the overall ecosystem responses (Riebesell & Gattuso, 2015). Indeed, OA may affect
53 indirectly the marine realm by reducing habitat complexity through the decline in habitat-forming
54 species that support high biodiversity (Barry et al. 2011). A recent study gathering data from naturally
55 acidified sites (i.e., CO₂ vents) projected indirect negative effects of OA on biodiversity due to the
56 reduction in structural complexity of biogenic habitat (i.e. coral reefs, mussel beds and some
57 macroalgal habitats) (Sunday et al. 2017). Nevertheless, there is a lack of evidence that biodiversity
58 will increase in systems where habitat-forming species could benefit from OA (i.e., seagrasses)
59 (Sunday et al. 2017).

60 Direct and indirect effects of OA have been documented on fish, although they are considered able
61 to cope with low pH thanks to their capacity of acid-base regulation (Ishimatsu et al. 2008). For
62 instance, OA can directly affect olfactory and auditory functions (Dixson et al. 2010, Simpson et al.
63 2011), behaviour and neurosensory functions (Munday et al. 2014, Nilsson et al. 2012), reproduction
64 (Miller et al. 2013), as well as calcareous structures (Bignami et al. 2013, Mirasole et al. 2017, Di
65 Franco et al. 2019). On the other hand, indirect effects can be driven by habitat alteration and
66 interactions with other species (Nagelkerken et al. 2015, 2017). Moreover, early life stages of fish are
67 considered particularly vulnerable to OA, with consequences on fish population replenishment
68 (Baumann et al. 2011, Munday et al. 2010, Rossi et al. 2016). Accordingly, it has been hypothesized

69 that fish biodiversity will be negatively affected by OA in a near future (Cheung et al. 2009) and
70 decline of catches worldwide is expected (Cheung, 2018).

71 CO₂ vents represent a great opportunity to test the effects of OA at high levels of the biological
72 hierarchy (i.e., community and ecosystem), as they are naturally acidified environments and are
73 widely used to fill the gap between laboratory experiments and the real world (Riebesell & Gattuso,
74 2015). Nevertheless, so far, most studies in CO₂ vents mainly focused on calcareous species and
75 ecosystem shifts driven by a decline not only in species richness at low pH conditions (Hall-Spencer
76 et al. 2008, Fabricius et al. 2011, Kroeker et al. 2012, Foo et al. 2018), but also a loss of functional
77 diversity (Teixido et al. 2018). Only recently, a few studies carried out in CO₂ vents addressed the
78 response of fish to OA at species (Nagelkerken et al. 2015, Cattano et al. 2017) and community levels
79 (Munday et al. 2014, Nagelkerken et al. 2017). The latter studies showed contrasting results on
80 changes in community structure and biodiversity, probably influenced by the different habitat use of
81 the fish studied. Indeed, although both studies found fish behavioural abnormalities, Nagelkerken et
82 al. (2017) found a loss in biodiversity of highly resident benthic fish, while Munday et al. (2014)
83 found only minor differences in the community structure of more mobile necto-benthic fish.

84 This study was conducted in the shallow CO₂ vent located in Vulcano Island (Aeolian Archipelago,
85 Italy) where the seagrass *Cymodocea nodosa* (Ucria) Ascherson, 1870, forms large meadows. In this
86 area, previous studies highlighted lower *C. nodosa* density and biomass close to the CO₂ vent area
87 compared with control sites, although overall characterized by a more intense metabolism and
88 photosynthetic activity (Apostolaki et al. 2014). *C. nodosa* has a crucial role in terms of primary
89 production, biodiversity and food web complexity (Cancemi et al. 2002) and is one of the most
90 important habitat-forming species for coastal fish, providing shelter from predators, nursery ground
91 and food (Guidetti & Bussotti, 2000).

92 The main aim of this study was to assess the effect of high *p*CO₂/low pH on fish assemblages in a
93 coastal area characterised by seagrass meadows. We compared fish assemblages long-term exposed
94 to high *p*CO₂/low pH conditions (low pH site) to those in normal pH conditions from two controls in

95 two periods of the year. In all sites and periods, we assessed species composition, fish biodiversity,
96 community and size-class structure, as well as *C. nodosa* meadow complexity in terms of shoot
97 density and leaf canopy height. We hypothesised a decline in fish biodiversity and a homogenization
98 of fish assemblage (over simplification of the community structure through the numerical dominance
99 of only a few species, *sensu* Nagelkerken et al. 2017) due to the combination of the stress effect of
100 low pH and the expected reduction in seagrass structural complexity. We also expect that the
101 detrimental effects are more severe on juveniles due to their higher vulnerability (Dixson et al. 2010,
102 Munday et al. 2010, 2014).

103

104 **Materials and methods**

105 Study area and fieldwork

106 The shallow venting system of Vulcano Island (Aeolian Archipelago, 24 km off the NE coast of
107 Sicily, Italy) is one of the most active and studied vent in the whole Mediterranean Sea. The main
108 venting area occurs at about 1 m depth on the south-western part of Levante Bay, on the eastern side
109 of the Island (Fig. 1) (Boatta et al. 2013). Overall, gas composition is dominated by CO₂ (97-99%
110 vol.), which generates a pH gradient (from 5.5. to 8.1) along the north shore of the bay (Boatta et al.
111 2013, Italiano 2009). The area is characterized by acidic and reducing conditions, causing changes in
112 major and trace element geochemical fluxes at the sediment-seawater interface (Vizzini et al. 2013).
113 Emissions include also small quantities of H₂S (< 2.2%), which rapidly decrease with distance from
114 the vent (Boatta et al. 2013). Water composition in terms of major elements and compounds (Cl, SO₄,
115 Na, K, Ca and Mg) is similar to that of Mediterranean surface waters, while greater variability has
116 been recorded for dissolved Fe concentrations, which showed high values close to the vents (Boatta
117 et al. 2013). Seawater carbonate chemistry parameters in the Levante Bay range between 2.78 and
118 3.17 mmol kg⁻¹ for total alkalinity and 0.02 and 3.64 for aragonite saturation state (Boatta et al. 2013).
119 Fish assemblage was studied through non-destructive underwater visual censuses (UVCs) (Harmelin-
120 Vivien et al. 1985) along 10 m long × 5 m wide strip transects (total area 50 m²), randomly placed

121 within *Cymodocea nodosa* meadows, at a depth between 2 and 5 m in three sites: a low pH site
122 (hereafter low pH) about 250 m far from the primary CO₂ vent in Levante Bay, and two Control sites:
123 Control 1 (Ctrl 1) about 500 m far from the primary vent, and Control 2 (Ctrl 2) in Lipari Island,
124 about 6.5 km far from the primary vent (Fig. 1). Controls were chosen for the same orientation (South-
125 East), depth (2-5 m) and vegetal coverage (*C. nodosa* meadow) as the low pH site. Visual census was
126 carried out in two periods of the year (Period I: from September to November 2014; Period II: from
127 May to July 2015), between 11.00 AM and 3.00 PM (18 replicates in total per site and period). Cryptic
128 fish (i.e. Blenniidae, Gobiidae, Scorpaenidae) were kept out from the surveys to avoid
129 underestimation (Willis 2001). Fish were identified to species level and abundance was estimated by
130 using seven pre-established abundance classes (1, 2-5, 6-10, 11-30, 31-50, 51-100, >100 number of
131 individuals) according to Harmelin-Vivien et al. (1985). Abundance was calculated by taking into
132 account the mid-point of each abundance class and expressed as mean fish abundance (ind. 50 m⁻²).
133 Fish were also assigned to three size-classes: small (S), medium (M) and large (L), corresponding
134 each to one-third of the maximum total length reported in the literature, according to FishBase online
135 database (Froese & Pauly, 2016), and grouped into trophic groups following the classification by
136 Guidetti & Sala (2007). In agreement with literature (Guidetti & Bussotti, 2000; Harmelin-Vivien et
137 al. 1985), we considered small, medium and large size corresponding to different fish life stages,
138 analogues of juveniles, sub-adults and adults.

139 Seawater temperature, salinity, dissolved oxygen (DO), pH and redox potential (Eh) were measured
140 in each site during the entire sampling period using a multiparameter probe (Hanna Instrument,
141 HI98194). Additionally, seagrass (*C. nodosa*) shoot density and leaf canopy height were measured.
142 *C. nodosa* shoot density was estimated by counting shoots (three replicates per transect) within a 400
143 cm² surface through a 20 x 20 cm frame and then converted to square meter. Leaf canopy height was
144 measured by recording the maximum length of the highest leaf in each shoot analysed (five shoots
145 per square, five replicates per transect).

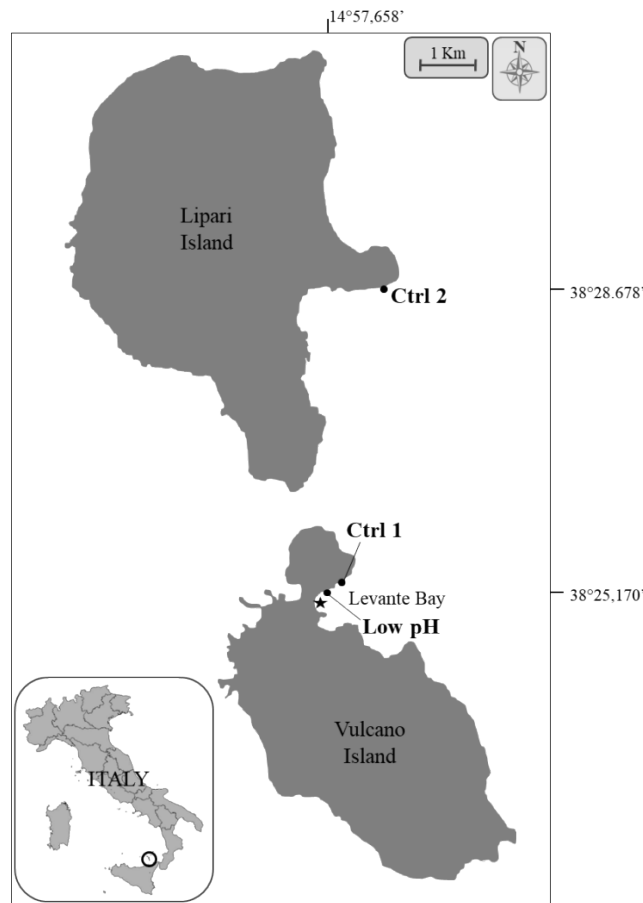


Fig. 1 - Map of the study area showing the sampling sites in Vulcano (Low pH and Ctrl 1) and Lipari Islands (Ctrl 2) (Aeolian Archipelago, Italy). The primary vent is located in Levante Bay (Vulcano) and is indicated by the star.

Data analysis and statistics

Univariate permutational analysis of variance (ANOVA) was used to test for differences among sites (fixed factor with 3 levels: low pH, Ctrl 1 and Ctrl 2) and between periods (fixed factor and orthogonal to Site, with 2 levels: PI and PII) for *Cymodocea nodosa* features (shoot density and leaf canopy height), fish diversity (species richness S, total abundance N, and size-class abundance N: N_{small} , N_{medium} , N_{large}). Data were previously transformed using the $\log(x + 1)$ notation and resembled using Bray Curtis similarity matrices (Primer 6/PERMANOVA+ software, Plymouth Marine Laboratory). Linear regression analysis was run to test the relationship between fish diversity (S, N, N_{small} , N_{medium} , N_{large}) and both *C. nodosa* features (shoot density and leaf canopy height) and pH values, by using Statistica software (Stat Soft V. 10).

160 Permutational ANOVA was used also to test for differences among sites and between periods in
 161 abundance of the most abundant species ($N > 2\%$ in at least one sampling site) taken individually
 162 (univariate analysis), all together and grouped in size-class (N_{small} , N_{medium} , N_{large}) (multivariate
 163 analysis). The analysis was carried out after transformation using the $\log(x + 1)$ notation and
 164 resemblance using Bray Curtis similarity matrices (Primer 6/PERMANOVA+ software, Plymouth
 165 Marine Laboratory). Only in the analysis of size-class abundance, a dummy variable = 1 was added
 166 to improve interpretability of displays and significance of tests when denuded samples were found
 167 (Clarke et al. 2006). To visualize the differences in fish abundance (total and size-class abundance)
 168 among sites and between periods, a constrained ordination (with the interaction “site x period” as
 169 constrained factor) was performed using canonical analysis of principal coordinates (CAP, Anderson
 170 & Willis 2003). A similarity percentage procedure (SIMPER) was used also to identify fish species
 171 mostly contributing to dissimilarities among sites.

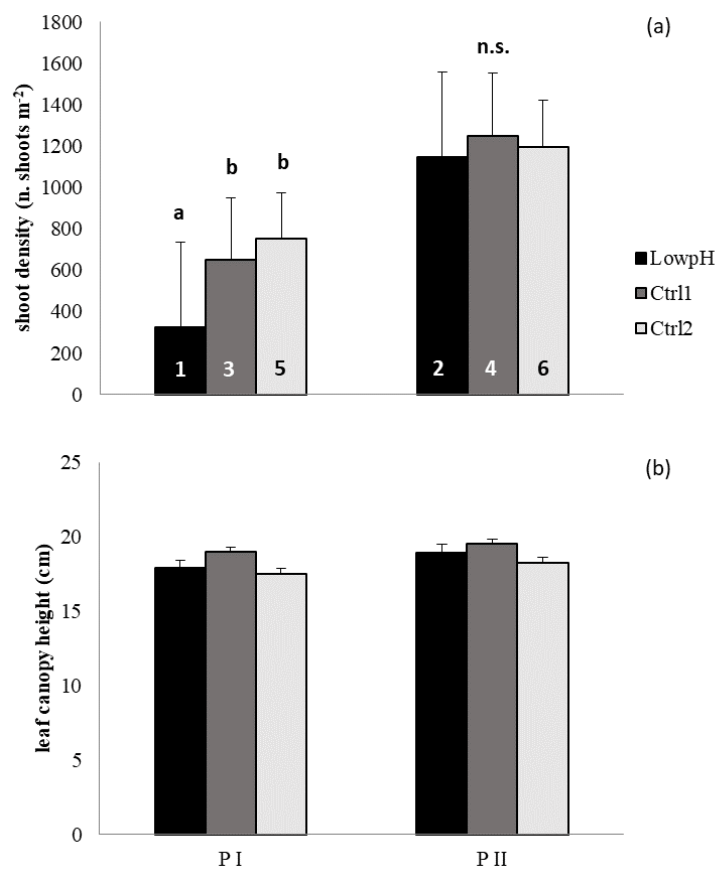
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173 **Results**

174 Environmental features

175 Temperature did not show large among-site wide variability, but increased from Period I (mean $\pm$ s.d.
 176 22.3 ± 2.4 °C; range: 20.1-25.5 °C) to Period II (23.9 ± 3.0 °C; 19.0-29.5 °C). Salinity (38.7 ± 0.4
 177 psu; 38.2-39.2 psu) and DO (7.3 ± 0.3 mg l⁻¹; 6.2-8.6 mg l⁻¹) varied little among sites and between
 178 periods. On the other hand, pH showed among-site variability, being on average 7.80 ± 0.09 in the
 179 Low pH site, 8.19 ± 0.03 in Ctrl 1 and 8.22 ± 0.02 in Ctrl 2. Consistently, mean Eh was 79.3 ± 5.6
 180 mV in the Low pH site, 85.6 ± 8.3 mV in Ctrl 1 and 111.6 ± 13.7 mV in Ctrl 2.

181 *C. nodosa* shoot density showed significant differences for the interaction ‘site x period’ (Pseudo-F
 182 $(2, 318) = 25.76$, $p < 0.001$). Pair-wise tests highlighted significantly lower densities in the Low pH site
 183 than in both Controls in Period I ($p < 0.001$), and also in Period I than in Period II at all sites (Fig.
 184 2a). No among-site and between-period differences in *C. nodosa* leaf canopy height were detected
 185 (Fig. 2b).



187

188 **Fig. 2** - Mean value ($\pm$ SE) of a) *Cymodocea nodosa* shoot density (n. shoots m⁻²) and b) leaf canopy height (cm) in the
189 three sites (Low pH, Ctrl 1 and Ctrl 2) and the two periods (PI and PII). Letters and numbers indicate homogeneous
190 groups within sites and periods, respectively. n.s. indicates not significant differences among sites.

191

192 Fish assemblages

193 Overall, nineteen fish species belonging to six different families were recorded. Assemblages were
194 dominated by Labridae (10 species) and Sparidae (5 species), followed by Mullidae, Mugilidae,
195 Pomacentridae and Serranidae (Annex I). Most species belonged to the invertivorous trophic group
196 with the exception of one herbivore (*Sarpa salpa*), one detritivore (*Mugil cephalus*), one piscivore
197 (*Serranus scriba*) and two planktivores (*Chromis chromis* and *Oblada melanura*). In general, the
198 most abundant species belonged to the families Labridae (i.e. *Coris julis* and *Symphodus ocellatus*),
199 Pomacentridae (*C. chromis*) and Sparidae (i.e. *Diplodus vulgaris* and *S. salpa*).

200 The most abundant fish species (N> 2% in at least one site) showed significant differences among the
201 three sites, except for *M. surmuletus* (Table 1). In particular, three species showed comparable

202 abundances in Low pH and Ctrl 1 sites, and lower (*D. vulgaris*) or higher (*C. chromis* and *C. julis*)
203 than in Ctrl 2. Three species (*S. ocellatus*, *S. salpa* and *S. tinca*) showed similar abundances in Low
204 pH and Ctrl 2, while only one species showed lower abundance in Low pH than both Controls (*O.*
205 *melanura*), and another species differed among the three sites with a marked increase from Low pH
206 to Ctrl 1 and Ctrl 2 (*T. pavo*). By comparing periods, only three species (*C. julis*, *S. tinca* and *T. pavo*)
207 showed a significantly higher abundance in Period I than in Period II, while one species (*S. salpa*)
208 showed the opposite trend.
209

Source of variation		<i>C. chromis</i>				<i>C. julis</i>				<i>D. vulgaris</i>			
	<i>df</i>	MS	Pseudo-F	p	Pair-wise	MS	Pseudo-F	p	Pair-wise	MS	Pseudo-F	p	Pair-wise
Site	2	6895.3	13.0	**	C2 < Low = C1	1622.8	8.8	***	C2 < Low = C1	5783.3	19.2	***	C1 = Low < C2
Period	1	1318.9	2.5	n.s.		4858.9	26.4	***	PII < PI	494.3	1.6	n.s.	
Site*Period	2	493.7	0.9	n.s.		411.1	2.2	n.s.		701.0	2.3	n.s.	
Residuals	102	528.9				183.8				300.8			

Source		<i>M. surmuletus</i>				<i>O. melanura</i>				<i>S. ocellatus</i>			
	<i>df</i>	MS	Pseudo-F	p	Pair-wise	MS	Pseudo-F	p	Pair-wise	MS	Pseudo-F	p	Pair-wise
Site	2	120.0	0.4	n.s.		2714.9	8.0	**	Low < C2 = C1	1516.6	3.0	*	C2 = Low < C1
Period	1	187.9	0.6	n.s.		93.5	0.3	n.s.		252.6	0.5	n.s.	
Site*Period	2	657.3	2.3	n.s.		395.8	1.2	n.s.		758.4	1.5	n.s.	
Residuals	102	285.2				338.6				498.7			

Source		<i>S. salpa</i>				<i>S. tinca</i>				<i>T. pavo</i>			
	<i>df</i>	MS	Pseudo-F	p	Pair-wise	MS	Pseudo-F	p	Pair-wise	MS	Pseudo-F	p	Pair-wise
Site	2	2175.1	4.0	*	C1 < C2 = Low	1799.1	5.4	**	C2 = Low < C1	4387.1	16.8	***	Low < C1 < C2
Period	1	3040.8	5.6	*	PI < PII	2061.3	6.2	*	PII < PI	1011.4	3.9	*	PII < PI
Site*Period	2	840.9	1.5	n.s.		387.3	1.2	n.s.		587.8	2.2	n.s.	
Residuals	102	542.8				332.5				260.7			

210 **Table 1** - Results of univariate permutational ANOVA testing for differences among Sites and between Periods in
211 abundance of the most abundant species (> 2%). Probability levels: n.s. = not significant- p > 0.05; * p ≤ 0.05; ** p ≤
212 0.01; *** p ≤ 0.001.

213

At multivariate level, fish assemblage showed significant differences for the interaction ‘site x period’ (Pseudo-F_(2,102) = 1.87, p < 0.05). Pair-wise tests revealed that the three sites differed each other in both sampling periods, and, in turns, the two periods were significantly different in all sites. Nevertheless, site seems the main factor discriminating the three groups in the CAP ordination (Fig. 3). In particular, Low pH is located on the down-right side of the graph, Ctrl 2 on the up-left side and Ctrl 1 interspersed between the other two sites, with no evident separation between periods within each site. Vectors superimposed onto the graph, showed that the main species contributing to the ordination were *C. chromis*, *C. julis* and *S. tinca* for samples from Low pH and Ctrl 1, and *D. vulgaris* and *T. pavo* mainly for samples from Ctrl 2. Furthermore, SIMPER analysis highlighted the lowest dissimilarity between Low pH and Ctrl 1, and the highest dissimilarity between Low pH and Ctrl 2, due to the species *C. chromis*, *S. salpa*, *D. vulgaris* and *T. pavo*. On the other hand, the dissimilarity between the two controls was due mainly to *C. chromis*, *D. vulgaris* and *S. ocellatus* (Table 2).

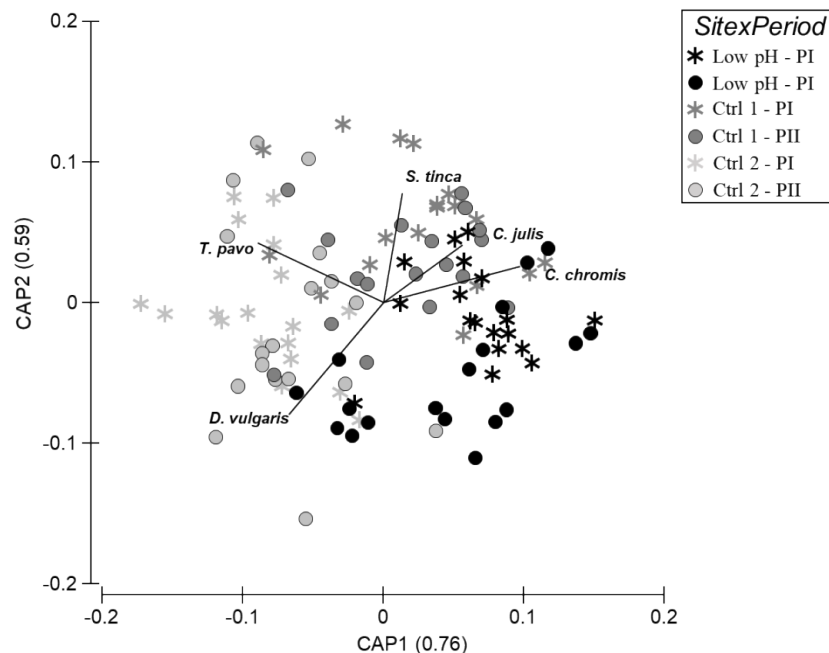


Fig. 3 - Canonical analysis of principal coordinates (CAP) of abundance of the most abundant species (> 2%) in the three sites (Low pH, Ctrl 1 and Ctrl 2) and the two periods (PI and PII). Vectors of the species contributing most to the ordination (Pearson correlation > 0.4) are superimposed. See the annex I for the whole scientific name of the species. The correlation of each axis with the resulted ordination is also indicated.

Species	Contr. %	Av. Ab.		Contr. %	Av. Ab.		Contr. %	Av. Ab.	
		Low pH	Ctrl 1		Low pH	Ctrl 2		Ctrl 1	Ctrl 2
<i>C. chromis</i>	17.32	1.35	1.33	15.79	1.35	0.27	14.8	1.33	0.27
<i>C. julis</i>	8.53	1.27	1.25	9.07	1.27	0.79	9.29	1.25	0.79
<i>D. vulgaris</i>	7.21	0.49	0.30	13.38	0.49	1.19	12.95	0.30	1.19
<i>M. surmuletus</i>	6.76	0.39	0.32	6.68	0.39	0.37	6.07	0.32	0.37
<i>O. melanura</i>	8.20	0.02	0.71	5.92	0.02	0.50	9.84	0.71	0.50
<i>S. ocellatus</i>	12.23	0.47	0.92	8.01	0.47	0.40	12.00	0.92	0.40
<i>S. salpa</i>	12.77	1.08	0.27	14.00	1.08	0.77	9.69	0.27	0.77
<i>S. tinca</i>	10.70	0.58	1.03	8.78	0.58	0.57	9.69	1.03	0.57
<i>T. pavo</i>	7.30	0.18	0.56	11.02	0.18	0.98	9.13	0.56	0.98
	Av. Dis. = 61.59			Av. Dis. = 67.41			Av. Dis. = 66.42		

Table 2 - SIMPER analysis showing fish species contributing most to the dissimilarity among sites and average abundance at each site (Low pH, Ctrl 1 and Ctrl 2). Av. Ab.: Average abundances; Contr. %: dissimilarity contribution; Av. Dis.: Average dissimilarity.

Fish grouped in size-classes differed among sites (N_{small} : Pseudo-F_(2,102) = 3.20, $p < 0.01$; N_{medium} : Pseudo-F_(2,102) = 4.58, $p < 0.001$; N_{large} : Pseudo-F_(2,102) = 5.20, $p < 0.001$) and between periods (N_{small} : Pseudo-F_(1,102) = 7.88, $p < 0.001$; N_{medium} : Pseudo-F_(1,102) = 2.61, $p < 0.05$; N_{large} : Pseudo-F_(1,102) = 2.50, $p < 0.05$). Pair wise tests showed that each class differed among the three sites, in both periods. Likewise CAP ordination of the whole assemblage, CAP of size-class abundance did not highlight any temporal separation, but partial spatial separations. In more detail, small and medium-sized fish from Low pH and Ctrl 1 showed a partial overlap in the centre-right side of the graphs, while Ctrl 2 laid in the centre-left side (Fig. 4a, b). Large fish from the three sites were even more distinct along the first axis (Fig. 4c). Moreover, the main species contributing to discriminate the sites were *C. julis* and *D. vulgaris* for small-sized fish, *D. vulgaris* and *S. ocellatus* for medium-sized fish, and *C. chromis* and *C. julis* for large-sized fish.

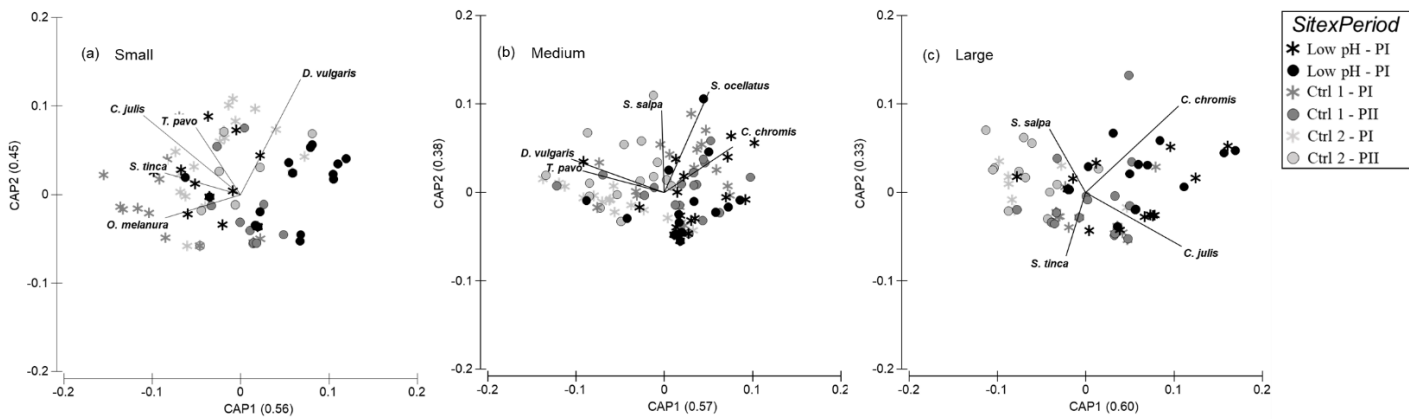


Fig. 4 - Canonical analysis of principal coordinates (CAP) of density of the most abundant species (> 2%) grouped per size-class: a) N_{small} , b) N_{medium} and c) N_{large} in the three sites (Low pH, Ctrl 1 and Ctrl 2) and the two periods (PI and PII). Vectors of the species contributing most to the ordination (Pearson correlation > 0.4) are superimposed. See the annex I for the whole scientific name of the species. The correlation of each axis with the resulted ordination is also indicated.

Fish biodiversity, in terms of species richness (S) and total abundance (N), showed different patterns when comparing sites and periods. Overall, S differed among sites (Pseudo- $F_{(2,102)} = 4.71$, $p < 0.05$) and periods (Pseudo- $F_{(1,102)} = 5.66$, $p < 0.05$), but not for their interaction. In more detail, significantly higher S values were observed in Ctrl 1 than in the other sites, and in Period I than in Period II (Fig. 5a). Despite the high spatial variability of N in Period I, any significant difference was detected, nor clear spatial or temporal patterns (Fig. 5b). Results of the regression analysis between fish diversity (S and N) and environmental variables (*C. nodosa* shoot density, leaf canopy height, pH) revealed a significant positive relationship only between both S and N and leaf canopy height (S: $n = 108$, $r = 0.22$, $p < 0.05$; N: $n = 108$, $r = 0.33$, $p < 0.01$).

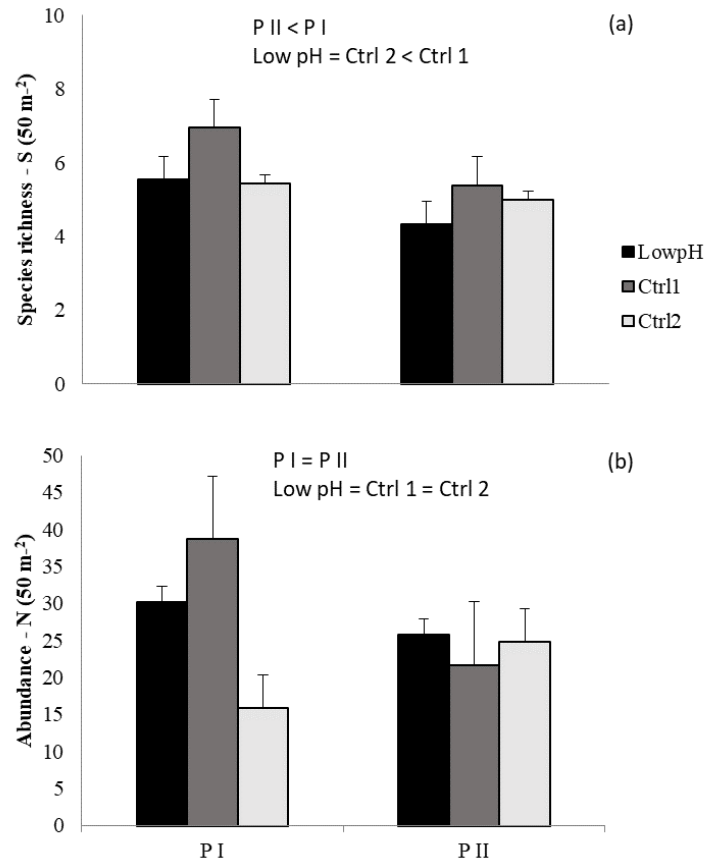


Fig. 5 - Mean value ($\pm$ SE) of a) species richness (S) and b) abundance (N) of fish in the three sites (Low pH, Ctrl 1 and Ctrl 2) and the two periods (P I and P II). Results of univariate permutational ANOVA pair-wise tests are also showed.

As regards size-class abundance (N_{small} , N_{medium} , N_{large}), small- and large-sized individuals showed contrasting temporal trends (Fig. 6a, c): N_{small} decreased significantly from Period I to Period II (Pseudo- $F_{(1,102)} = 4.88$; $p < 0.05$), when N_{large} increased (Pseudo- $F_{(1,102)} = 5.07$; $p < 0.05$). Abundance of medium-sized fish, N_{medium} , did not differ between periods, but it was higher in Ctrl 1 than in the other sites (Pseudo- $F_{(2,102)} = 5.43$; $p < 0.01$) (Fig. 6b). Moreover, likewise total abundance N, N_{small} was positively correlated with *C. nodosa* leaf canopy height ($n = 108$, $r = 0.37$; $p < 0.001$)

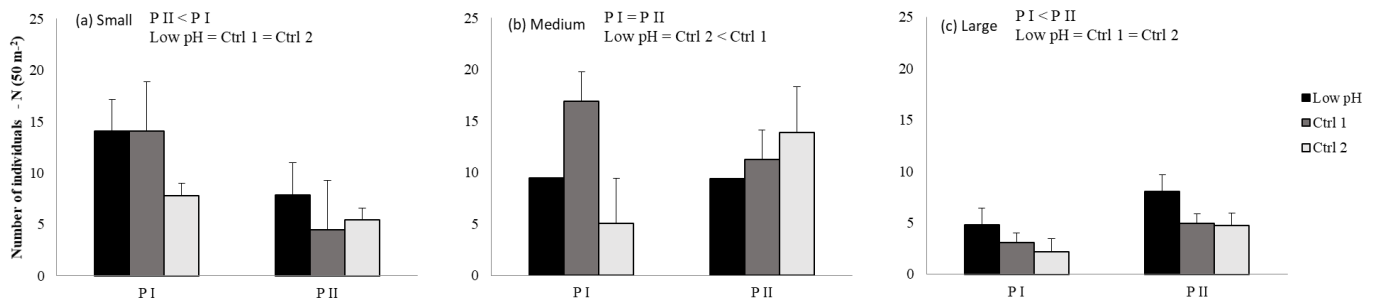


Fig. 6 – Size-class abundance (N) ($\pm$ SE) of a) small (N_{small}), b) medium (N_{medium}) and c) large (N_{large}) fish in the three sites (Low pH, Ctrl 1 and Ctrl 2) and the two periods (P I and P II). Results of univariate permutational ANOVA pair-wise tests are also showed.

Discussion

The forecasted decrease in ocean pH over the next decades (IPCC, 2014) is predicted to negatively affect marine ecosystems and biodiversity through direct and indirect effects (Cheung et al. 2009, Sunday et al. 2017). Even though calcareous organisms are considered more susceptible (Doney et al. 2009), also fish are expected to be directly and/or indirectly affected by ocean acidification (OA), with consequences on their distribution and abundance (Perry et al. 2005, Cattano et al. 2018).

In the present study, we investigated fish communities in a shallow volcanic CO₂ vent to test the hypothesis of a decline in fish biodiversity and the homogenization of fish assemblage in the low pH site due to the combination of the stress effect of acidic conditions and the expected reduction in seagrass structural complexity. We also hypothesised that juveniles are more affected by low pH due to their higher vulnerability to OA (Dixon et al. 2010, Munday et al. 2010, 2014).

Fish diversity, in terms of species richness and abundance, did not show a univocal spatial pattern and significant relationships with pH. However, fish assemblages changed among sites with *Chromis chromis* and *Coris julis* higher in both the acidified site and the close control than in the far control, while *Diplodus vulgaris* and *Thalassoma pavo* showed an opposite trend. Moreover, *C. chromis* dominated the assemblages in both low pH site and the adjacent control, together with *Sarpa salpa* in the former site, and *Symphodus ocellatus* in the latter. In contrast, no species clearly dominated the fish assemblage in the far control, except for *S. salpa* in only one of the two investigated periods.

301 These results suggest that pH played a role in shaping fish assemblage, with a certain species overlap
302 between the low pH site and the close control, although the latter was characterised by boosted species
303 richness.

304 *S. salpa*, which was one of the most influent species in discriminating sites (above all low pH vs. the
305 two controls) is one of the most important herbivorous fish in the Mediterranean Sea. The minor
306 content of phenolic substances, which are deterrent to herbivory (Arnold et al. 2012) and boost the
307 palatability of *Cymodocea nodosa* long-term exposed to high $p\text{CO}_2$ (Apostolaki et al. 2014), may
308 have led to a higher attractiveness of the acidified site for the high mobile *S. salpa*, which indeed was
309 more abundant than in the close control. This result opens interesting scenarios about the role that
310 herbivores may have in future acidified oceans, because of their control on primary producers through
311 top-down forces (Poore et al. 2012). Nowadays, new insights are emerging about the role of
312 herbivores and their compensatory effect under high CO_2 conditions to maintain communities'
313 resistance to disturbance (Ghedini et al. 2015). Indeed, compensatory dynamics are important
314 stabilising mechanisms through which communities respond to environmental changes (Ghedini et
315 al. 2015).

316 Similarly to *S. salpa*, the spatial patterns observed for the damselfish *C. chromis*, which showed a
317 clear dominance and a significantly higher abundance in both the low pH and close control, than in
318 the further control site, suggest its high tolerance to OA. Indeed, although a high species-specific
319 sensitivity to elevated CO_2 was observed in other damselfish species (Ferrari et al. 2011), more recent
320 studies highlighted a high resistance of damselfish to OA (Kwan et al. 2017), as well as an improved
321 aerobic performance when exposed to high $p\text{CO}_2$ (Rummer et al. 2013) and an unaffected gregarious
322 behavior (group cohesion) under high $p\text{CO}_2$ levels (Cattano et al. 2019).

323 On the other hand, among the species that mainly contributed to discriminate sites, particularly
324 interesting is also the case of the two sympatric Labridae, the rainbow wrasse *C. julis* and the ornate
325 wrasse *T. pavo*, which are widespread in the Mediterranean Sea. Here, their different spatial
326 distribution was striking: *C. julis* abundance was similar in the Low pH and the adjacent control site,

327 and higher than in the far control, while *T. pavo* showed the highest abundance in the far control and
328 the lowest in the acidified site. These findings let hypothesize that *C. julis* is more tolerant to acidified
329 conditions compared to the sympatric species and open a new question regarding the combined effects
330 of warming and acidification. Indeed, recent studies showed that the interaction of these species is
331 potentially exacerbated by seawater warming and that their distribution differs according to water
332 temperature (Milazzo et al. 2013, 2016).

333 Looking at the whole assemblage, the close control site had a fish community structure partially
334 overlapping both the low pH site and the far control, and showed the lowest dissimilarity with the
335 low pH site. This result, together with the among-site similarity in fish total abundance, provides
336 evidence that fish did not avoid the low pH site, and this is possibly because the boosted food
337 resources (low-order invertebrates; Vizzini et al. 2017) counteract the low pH-induced stress. At the
338 same time, fish may take advantage from their mobility to escape chronic exposure to high $p\text{CO}_2$ /low
339 pH values and take shelter in the close control, which may represent an adjacent ‘recovery area’. This
340 hypothesis is supported also by the high percentage of invertivorous fish (~ 75 %) in the low pH site
341 and the similar isotopic ($\delta^{13}\text{C}$, $\delta^{15}\text{N}$) composition and niches of fish (Mirasole, unpublished data)
342 between the low pH site and the close control site. The high cost for the acid-base regulation under
343 acidified conditions may be indeed compensated by the higher prey availability associated to
344 macrophytes in the acidified area (Vizzini et al. 2017), corroborating the results found by
345 Nagelkerken et al. (2015; 2017) where the increase in fish density was driven by higher resource
346 abundances.

347 When explaining the overall similarity of fish assemblage between the low pH and adjacent control
348 site, natural fluctuations of pH must be also taken into account. Due to the geo-morphological settings
349 of the Levante Bay, acidified water masses mostly run parallel to the northern shoreline of the bay,
350 when predominant winds belong to north-western sectors (Boatta et al. 2013). However, the pH
351 gradient created by the primary vent along the northern cost of the bay is reduced or deleted when
352 wind direction is from south-east (Sirocco wind). As suggested by Kroeker et al. (2012), it is possible

353 that natural fluctuations in carbonate chemistry in vent areas allow organisms to better tolerate, or
354 even adapt to, these particular conditions. Indeed, laboratory studies found negative effects on
355 organisms mainly when there is a sharp and constant reduction in pH, without time of adaptation
356 (Ishimatsu et al. 2008). Evolutionary mechanisms may help organisms to adapt from one generation
357 to another, but this mechanism implies genetic variation (Crozier & Hutchings, 2014).
358 Transgenerational acclimation occurs when the environment experienced by parents influences the
359 performance of offspring (Allan et al. 2014, Sunday et al. 2014, Cattano et al. 2016) showing that
360 adaptive plasticity mechanisms can have a crucial role for fish, improving their resilience to
361 environmental change (Munday 2014, Fox et al. 2019). However, on the other hand, it has been
362 already suggested that population replenishment may occur from “non-acidified populations”
363 (Kroeker et al. 2012, Munday et al. 2014). Indeed, most fish species analysed in the present study
364 have a long pelagic larval stage ranging between 10 and 30 days (Raventos & MacPherson, 2001). In
365 this case, plastic responses in physiology, morphology, or behaviour may help maintaining fitness in
366 a new stressful environment and take less time (days to months, or within a life stage). Moreover, a
367 meta-analysis on different taxonomic groups highlighted that highly mobile organisms (i.e. fish or
368 crustaceans) with developed intracellular/extracellular pH regulatory mechanisms may be more
369 resilient to OA (Kroeker et al. 2013).

370 Considering fish size-class, only sub-adults showed among-site differences, with significantly higher
371 abundance in the close control site than in the acidified and in the far control site. In contrast, the
372 similar abundance of juveniles across sites rules out the expected negative effects of low pH values
373 on fish early life stages, as previously found both in laboratory experiments (Baumann et al. 2011)
374 and in field studies (Rossi et al. 2016). The contrasting results with literature may be due to a species-
375 specific vulnerability (Ferrari et al. 2011) and/or constrained experimental conditions, such as limited
376 exposure time and high $p\text{CO}_2$ levels (Ishimatsu et al. 2008), and suggest that, contrarily to what
377 expected, juveniles may cope with OA natural conditions. Recent studies conducted in the same area

378 of Vulcano island, highlighted bigger sizes (Cattano et al. 2017) and larger otoliths (Di Franco et al.
379 2019) in *S. ocellatus* juveniles, probably resulting in higher survival rates.

380 As regards the relation between fish assemblage and seagrass meadows, species richness, total and
381 juvenile abundances correlated positively only with *C. nodosa* leaf canopy height, which however
382 did not change between low pH site and controls. This pattern confirms the role of seagrass meadow
383 structure in supporting fish diversity, and, in particular, juveniles (Guidetti & Bussotti, 2000, 2002,
384 Bell & Westoby, 1986, Hori et al. 2009), regardless of pH conditions. On the other hand, consistently
385 with Apostolaki et al. (2014), *C. nodosa* shoot density was overall lower in the Low pH site, and in
386 the first sampling period (i.e. autumn) when fish species richness reached higher values. These results
387 may indicate a stronger effect of low pH conditions on *C. nodosa* density in autumn/winter when the
388 seagrass is featured by low structural complexity (Guidetti & Bussotti, 2000; Cancemi et al. 2002),
389 and suggests a lower effect of density than leaf canopy height in shaping fish diversity, accordingly
390 with the literature (Bell & Westoby, 1986, Hori et al. 2009).

391 Overall, the patterns in species distribution and fish diversity observed in this study were not
392 consistent with those found in another shallow CO₂ vent in the southern hemisphere (New Zealand),
393 where species richness dropped in the acidified site compared with the control site, due to a strong
394 homogenization of community structure (Nagelkerken et al. 2017). Nagelkerken et al. (2017) found
395 a reduction of predatory pressure coupled with an increase of behaviourally dominant fish and a loss
396 of local biodiversity. This pattern was also attributed to the OA-mediated enhancement of food
397 resources, which balanced the stressful conditions at the vent site. Here we do not find a loss in species
398 richness, nor a decrease of fish abundance, although a few species were prevalent in the low pH site.
399 The dissimilar results may be due to the different habitat use of the fish assemblages. Indeed, the
400 different mobility and habitat use of the fish addressed, highly territorial benthic fish in Nagelkerken
401 et al. (2017) vs. nekto-benthic fish in this study, may explain the contrasting results. On the other
402 hand, in another CO₂ vent (Papua New Guinea), consistently with the present study, diversity and
403 community structure of the reef nekton-benthic fish differed little between acidified and control sites

404 (Munday et al. 2014). If Munday et al. (2014) attributed this pattern to a different habitat complexity
405 and composition in corals, rather than to a direct effect of high CO₂, here pH was an important driver
406 of fish assemblage, as its spatial and temporal structure was independent on that of seagrass meadows.
407 In conclusion, our findings suggest that near-future OA predictions (pH = 7.8 in the year 2100, IPCC
408 2014) may influence fish assemblages. In general, the nekto-benthic fish studied were able to cope
409 with OA, form well-structured fish assemblages, despite dominated by high-tolerant species, and
410 seem to benefit from an adjacent ‘recovery area’. Therefore, because of their behavioural and
411 physiological features (mobile habitus, capacity of acid-base regulation) and indirect effects of high
412 pCO₂/low pH in CO₂ vents (i.e. greater food availability), fish can balance the higher energetic cost
413 to live under high pCO₂ environments without major changes in community structure. Being habitat
414 use crucial in influencing fish response to environmental stressors, further investigations are needed
415 to assess if this pattern is exacerbated in highly territorial benthic fish exposed to high pCO₂/low pH
416 conditions. Also, further data are needed to forecast the species-specific responses of other seagrass
417 systems and the resulting impact on associated fauna. Finally, this study confirms the need to use *in*
418 *situ* experiments and naturally acidified environments to evaluate the responses of whole
419 communities long-term exposed to low pH conditions.

420

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426

427 Conflict of interest

428 The authors declare that they have no conflict of interest.

429

430 Ethical approval

431 All applicable international, national, and/or institutional guidelines for the care and use of animals

432 were followed.

References

1. Allan BJM, Miller GM, McCormick MI, Domenici P, Munday PL (2014) Parental effects improve escape performance of juvenile reef fish in a high-CO₂ world. *Proc R Soc B* 281: 20132179.
2. Anderson MJ, Willis TJ. (2003) Canonical analysis of principal coordinates: A useful method of constrained ordination for ecology. *Ecology* 84: 511-525.
3. Apostolaki ET, Vizzini S, Hendriks I, Olsen YS (2014) Seagrass ecosystem response to long-term high CO₂ in a Mediterranean volcanic vent. *Mar Environ Res* 99: 9-15.
4. Arnold T, Mealey C, Leahey H, Miller AW, Hall-Spencer JM., Milazzo M, Maers K (2012) Ocean acidification and the loss of phenolic substances in marine plants. *PLoS ONE*. 7: e35107.
5. Barry JP, Widdicombe S, Hall-Spencer JM (2011) Effects of ocean acidification on marine biodiversity and ecosystem function. *Ocean acidification*. pp 192-209.
6. Baumann H, Talmage SC, Gobler CJ (2011) Reduced early life growth and survival in a fish in direct response to increased carbon dioxide. *Nat Clim Change* 2: 38-41.
7. Bell JD, Westoby M (1986) Importance of local changes in leaf height and density to fish and decapods associated with seagrasses. *J Exp Mar Biol Ecol* 104(1-3): 249-274.
8. Bignami S, Enochs IC, Manzello DP, Sponaugle S, Cowen RK (2013) Ocean acidification alters the otoliths of a pantropical fish species with implications for sensory function. *PNAS* 110: 7366-7370.
9. Boatta F, D'Alessandro W, Gagliano AL, Liotta M, Milazzo M, Rodolfo-Metalpa R, Hall-Spencer JM, Parello F (2013) Geochemical survey of Levante Bay, Vulcano Island (Italy), a natural laboratory for the study of ocean acidification. *Mar Pollut Bull* 73: 485-494.
10. Cancemi G, Buia MC, Mazzella L (2002) Structure and growth dynamics of *Cymodocea nodosa* meadows. *Sci Mar* 66: 365-373.
11. Cattano C, Giomi F, Milazzo M (2016) Effects of ocean acidification on embryonic respiration and development of a temperate wrasse living along a natural CO₂ gradient. *Conserv Physiol* 4: 1-10.
12. Cattano C, Calò, A, Di Franco A, Firmamento R, Quattrocchi F, Sdiri K, Guidetti P, Milazzo M (2017) Ocean acidification does not impair predator recognition but increases juvenile growth in a temperate wrasse off CO₂ seeps. *Mar Environ Res* 132: 33-40.
13. Cattano C, Claudet J, Domenici P, Milazzo M (2018) Living in a high CO₂ world: a global meta-analysis shows multiple trait-mediated fish responses to ocean acidification. *Ecol Monogr* 88(3): 320-335.

14. Cattano C, Fine M, Quattrocchi F, Holzman R, Milazzo M (2019) Behavioural responses of fish groups exposed to a predatory threat under elevated CO₂. *Mar Environ Res*. doi: <https://doi.org/10.1016/j.marenvres.2019.04.011>.
15. Cheung WWL, Lam VW, Sarmiento JL, Kearney K, Watson R, Pauly D (2009) Projecting global marine biodiversity impacts under climate change scenarios. *Fish Fish* 10: 235-251.
16. Cheung WWL (2018) The future of fishes and fisheries in the changing oceans. *J Fish Biol*. 92(3): 790-803.
17. Clarke KR, Somerfield PJ, Chapman MG (2006) On resemblance measures for ecological studies, including taxonomic dissimilarities and a zero-adjusted Bray–Curtis coefficient for denuded assemblages. *J Exp Mar Bio Ecol* 330(1): 55-80.
18. Crozier LG, Hutchings JA (2014) Plastic and evolutionary responses to climate change in fish. *Evol Applications* 7: 68-87.
19. Di Franco A, Calò A, Sdiri K, Cattano C, Milazzo M, Guidetti P (2019) Ocean acidification affects somatic and otolith growth relationship in fish: evidence from an in situ study. *Biol Lett* 15: 20180662. <http://dx.doi.org/10.1098/rsbl.2018.0662>
20. Dixon DL, Munday PL, Jones GP (2010) Ocean acidification disrupts the innate ability of fish to detect predator olfactory cues. *Ecol Lett* 13: 68-75.
21. Doney SC, Fabry VJ, Feely RA, Kleypas JA (2009) Ocean acidification: The other CO₂ problem. *Annu Rev Mar Sci* 1: 169-192.
22. Fabricius KE, Langdon C, Uthicke S, Humphrey C, Noonan S, De'ath G, Okazaki R, Muehllehner N, Glas MS, Lough JM (2011) Losers and winners in coral reefs acclimatized to elevated carbon dioxide concentrations. *Nat Clim Change* 1: 165-169.
23. Ferrari MC, Dixon DL, Munday PL, McCormick MI, Meekan MG, Sih A, Chivers DP (2011) Intrageneric variation in antipredator responses of coral reef fishes affected by ocean acidification: Implications for climate change projections on marine communities. *Glob Change Biol* 17: 2980–2986.
24. Foo SA, Byrne M, Ricevuto E, Gambi MC (2018) The carbon dioxide vents of Ischia, Italy, a natural system to assess impacts of ocean acidification on marine Ecosystems: an overview of research and Comparisons with other vent systems. *Oceanogr Mar Biol* 56: 237-310.
25. Fox RJ, Donelson JM, Schunter C, Ravasi T, Gaitán-Espitia JD (2019) Beyond buying time: the role of plasticity in phenotypic adaptation to rapid environmental change. *Phil Trans R Soc B* 374: 20180174. <http://dx.doi.org/10.1098/rstb.2018.0174>.
26. Froese R, Pauly D (2016) FishBase. World Wide Web electronic publication. www.fishbase.org, version (06/2016).

27. Ghedini G, Russell BD, Connell SD (2015) Trophic compensation reinforces resistance: herbivory absorbs the increasing effects of multiple disturbances. *Ecol Lett* 18: 182-187.
28. Guidetti P, Bussotti S (2000) Fish Fauna of a Mixed Meadow Composed by the Seagrasses *Cymodocea nodosa* and *Zostera noltii* in the Western Mediterranean. *Oceanol Acta* 23: 759-770.
29. Guidetti P, Bussotti S (2002) Effects of seagrass canopy removal on fish in shallow Mediterranean seagrass (*Cymodocea nodosa* and *Zostera noltii*) meadows: a local-scale approach. *Mar Biol* 140: 445-453.
30. Guidetti P, Sala E (2007) Community-wide effects of marine reserves in the Mediterranean Sea. *Mar Ecol Prog Ser* 335: 43-56.
31. Hall-Spencer JM, Rodolfo-Metalpa R, Martin S, Ransome E, Fine M, Turner SM, Rowley SJ, Tedesco D, Buia MC (2008) Volcanic carbon dioxide vents show ecosystem effects of ocean acidification. *Nature* 454: 96-99.
32. Harmelin-Vivien ML, Harmelin JG, Chauvet C, Duval C, Galzin R, Lejeune P, Barnabé G, Blanc F, Chevalier R, Duclerc J, Lassere G (1985) Evaluation visuelle des peuplements et populations de poissons: méthodes et problèmes. *Rev Ecol (Terre Vie)* 40: 467–539.
33. Hori M, Suzuki T, Monthum Y, Srisombat T, Tanaka Y, Nakaoka M, Mukai H (2009) High seagrass diversity and canopy-height increase associated fish diversity and abundance. *Mar Biol* 156(7): 1447-1458.
34. IPCC (2014) Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change [Core Writing Team, R.K. Pachauri and L.A. Meyer (eds.)]. IPCC, Geneva, Switzerland, pp 151.
35. Ishimatsu A, Hayashi M, Kikkawa T (2008) Fishes in high-CO₂ acidified oceans. *Mar Ecol Prog Ser* 373: 295–302.
36. Italiano F (2009) Hydrothermal fluids vented at shallow depths at the Aeolian Islands: relationships with volcanic and geothermal systems. *FOG – Freiberg Online Geology* 22: 55-60.
37. Kroeker KJ, Micheli F, Gambi MC (2012) Ocean acidification causes ecosystem shifts via altered competitive interactions. *Nat Clim Change* 3: 156-159.
38. Kroeker KJ, Kordas RL, Crim R, Hendriks IE, Ramajo L, Singh GS, Duarte CM, Gattuso JP (2013) Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. *Glob Change Biol* 19: 1884–1896.

39. Kwan GT, Hamilton TJ, Tresguerres M (2017) CO₂-induced ocean acidification does not affect individual or group behaviour in a temperate damselfish. *R Soc Open Sci* 4: 170283. <http://dx.doi.org/10.1098/rsos.170283>.
40. Milazzo M, Mirto S, Domenici P, Gristina M (2013) Climate change exacerbates interspecific interactions in sympatric coastal fishes. *J Animal Ecol* 82: 468–477.
41. Milazzo M, Quattrocchi F, Azzurro E, Palmeri A, Chemello R, Di Franco A, Guidetti P, Sala E, Sciandra M, Badalamenti F, García-Charton JA (2016) Warming-related shifts in the distribution of two competing coastal wrasses. *Mar Environ Res* 120: 55-67
42. Miller GM, Watson SA, McCormick MI, Munday PL (2013) Increased CO₂ stimulates reproduction in a coral reef fish. *Glob Change Biol* 19: 3037–3045.
43. Mirasole A, Gillanders BG, Reis-Santos P, Grassa F, Capasso G, Scopelliti G, Mazzola A, Vizzini S (2017) The influence of high *p*CO₂ on otolith shape, chemical and carbon isotope composition of six coastal fish species in a Mediterranean shallow CO₂ vent. *Mar Biol* 164(9): 191.
44. Munday PL, Dixon DL, McCormick MI, Meekan M, Ferrari MCO, Chiversd DP (2010) Replenishment of fish populations is threatened by ocean acidification. *PNAS* 107:12930-12934.
45. Munday PL, Cheal AJ, Dixon DL, Rummer JL, Fabricius KE (2014) Behavioural impairment in reef fishes caused by ocean acidification at CO₂ seeps. *Nat Clim Change* 4: 487-492.
46. Munday PL (2014) Transgenerational acclimation of fishes to climate change and ocean acidification. *F1000prime reports* 6.
47. Nagelkerken I, Connell SD (2015) Global alteration of ocean ecosystem functioning due to increasing human CO₂ emissions. *Proc Natl Acad Sci USA* 112: 13272-13277.
48. Nagelkerken I, Russell BD, Gillanders BM, Connell SD (2015) Ocean acidification alters fish populations indirectly through habitat modification. *Nat Clim Change* doi: 10.1038/nclimate2757.
49. Nagelkerken I, Goldenberg S, Ferreira C, Russell BD, Connell SD (2017) Species interactions drive fish biodiversity loss in a high-CO₂ world. *Curr Biol*. <http://dx.doi.org/10.1016/j.cub.2017.06.023>.
50. Nilsson GE, Dixon DL, Domenici P, McCormick MI, Sørensen C, Watson S, Munday PL (2012) Near-future carbon dioxide levels alter fish behaviour by interfering with neurotransmitter function. *Nat Clim Change* 2: 201-204.
51. Perry AL, Low PJ, Ellis JR, Reynolds JD (2005) Climate change and distribution shifts in marine fishes. *Science* 308: 1912-1915.

52. Poore AGB, Campbell AH, Coleman RA, Edgar GJ, Jormalainen V, Reynolds PL, Sotka EE, Stachowicz JJ, Taylor RB, Vanderklift MA, Duffy JE (2012) Global patterns in the impact of marine herbivores on benthic primary producers. *Ecol Lett* 15: 912-922.
53. Raventos N, Macpherson E (2001) Planktonic larval duration and settlement marks on the otoliths of Mediterranean littoral fishes. *Mar Biol* 138: 1115-1120.
54. Riebesell U, Gattuso JP (2015) Lessons learned from ocean acidification research. *Nat Clim Change* 5: 12-14.
55. Rossi T, Nagelkerken I, Pistevos JCA, Connell SD (2016) Lost at sea: ocean acidification undermines larval fish orientation via altered hearing and marine soundscape modification. *Biol Lett* 12: 20150937.
56. Rummer JL, Stecyk JAW, Couturier CS, Watson S-A, Nilsson GE, Munday PL. (2013) Elevated CO₂ enhances aerobic scope of a coral reef fish. *Conserv Physiol* 1: doi:10.1093/conphys/cot023.
57. Simpson SD, Munday PL, Wittenrich ML, Manassa R, Dixson DL, Gagliano M, Yan HY (2011) Ocean acidification erodes crucial auditory behaviour in a marine fish. *Biol Lett* 7: 917-920.
58. Sunday JM, Calosi P, Dupont S, Munday PL, Stillman JH, Reusch TB (2014) Evolution in an acidifying ocean. *Trends Ecol Evol* 29: 117–125.
59. Sunday JM, Fabricius KE, Kroeker KJ, Anderson KM, Brown NE, Barry JP, Connell SD, Dupont S, Gaylord B, Hall-Spencer JM, Klinger T, Milazzo M, Munday PL, Russell BD, Sanford E, Thiyagarajan V, Vaughan MLH, Widdicombe S, Harley CDG (2017) Ocean acidification can mediate biodiversity shifts by changing biogenic habitat. *Nat Clim Change* 7: 81–85.
60. Teixido N, Gambi MC, Parravicini V, Kroeker K, Micheli F, Villeger S, Ballesteros E (2018) Functional biodiversity loss along natural CO₂ gradients. *Nat Commun.* 9 (1). DOI.10.1038/s41467-018-07592-1.
61. Vizzini S, Di Leonardo R, Costa V, Tramati CD, Luzzu F, Mazzola A (2013) Trace element bias in the use of CO₂ vents as analogues for low pH environments: Implications for contamination levels in acidified oceans. *Estuar Coast Shelf. S* 134: 19-30.
62. Vizzini S, Martínez-Crego B, Andolina C, Massa-Gallucci A, Connel SD, Gambi MC (2017) Ocean acidification as a driver of community simplification via the collapse of higher-order and rise of lower-order consumers. *Sci Rep.* 7:4018.
63. Willis (2001) Visual census methods underestimate density and diversity of cryptic reef fishes. *J Fish Biol* 59: 1408-1411.

602 **Annex I** - List of all fish species censused in the three sites (Low pH, Ctrl 1 and Ctrl 2). Trophic group and total relative
603 abundance (%) of each species in the two periods (PI and PII) are indicated. Trophic groups: Invertivore (INV);
604 Detritivore (DET); Planktivore (PLK); Small piscivore (PISC); Herbivore (HER).

Family	Species	Trophic Group	Low pH		Ctrl 1		Ctrl 2	
			PI	PII	PI	PII	PI	PII
Labridae	<i>Coris julis</i>	INV	7.7	3.5	6.8	4.5	6.3	1.9
	<i>Labrus viridis</i>	INV	<2	<2	<2	<2	<2	<2
	<i>Symphodus cinereus</i>	INV	<2	<2	<2	<2	<2	<2
	<i>Symphodus doderleini</i>	INV	<2	<2	<2	<2	<2	<2
	<i>Symphodus ocellatus</i>	INV	7.2	<2	15.6	2.5	<2	7.2
	<i>Symphodus mediterraneus</i>	INV	<2	<2	<2	<2	<2	<2
	<i>Symphodus roissali</i>	INV	<2	<2	<2	<2	<2	<2
	<i>Symphodus rostratus</i>	INV	<2	<2	<2	<2	<2	<2
	<i>Symphodus tinca</i>	INV	3.1	<2	6.1	2.7	3.5	2.3
	<i>Thalassoma pavo</i>	INV	<2	<2	2.2	<2	8.4	4.0
Mugilidae	<i>Mugil cephalus</i>	DET	<2	<2	<2	<2	<2	<2
Mullidae	<i>Mullus surmuletus</i>	INV	<2	4.5	<2	<2	2.3	<2
Pomacentridae	<i>Chromis chromis</i>	PLK	12.1	9.5	12.0	12.2	<2	4.7
Serranidae	<i>Serranus scriba</i>	PISC	<2	<2	<2	<2	<2	<2
Sparidae	<i>Oblada melanura</i>	PLK	<2	<2	9.6	<2	3.6	3.6
	<i>Diplodus annularis</i>	INV	<2	<2	<2	<2	<2	<2
	<i>Diplodus sargus</i>	INV	<2	<2	<2	<2	<2	<2
	<i>Diplodus vulgaris</i>	INV	<2	3.1	<2	<2	8.0	8.0
	<i>Sarpa salpa</i>	HER	16.4	21.4	<2	4.7	<2	26.2

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