

# Evolution of DS-1-like human G2P[4] rotaviruses assessed by complete genome analyses

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Group A rotaviruses (RVAs) are a leading cause of viral gastroenteritis in children, with G2P[4] RVA being one of the most common human strains worldwide. The complete genome sequences of nine G2P[4] RVA strains, selected from a 26-year archival collection (1985–2011) established in Palermo, Italy, were determined. A strain associated with a peak of G2P[4] RVA activity in 1996 resembled a reassortant strain identified in Kenya in 1982 and differed completely in genomic make up from more recent strains that circulated during 2004–2011. Conversely, the 2004–2011 G2P[4] RVAs were genetically more similar to contemporary RVA strains circulating globally. Recent G2P[4] strains possessed either single or multiple genome segments (VP1, VP3 and/or NSP4) likely derived from ruminant viruses through intra-genotype reassortment. Amino acid substitutions were selected and maintained over time in the VP7 and VP8\* antigenic proteins, allowing the circulation of two contemporary G2P[4] variants to be distinguished. Altogether, these findings suggest that major changes in the genomic composition of recent G2P[4] RVAs occurred in the early 2000s, leading to the appearance of a novel variant of the DS-1-like genotype constellation. Whether the modifications observed in the neutralizing antigens and in the genome composition of modern G2P[4] RVAs may affect the long-term effectiveness of the vaccination programmes remains to be explored.

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## INTRODUCTION

Group A rotaviruses (RVAs), family *Reoviridae*, are major pathogens causing gastroenteritis in young children, with a mortality rate of 453 000 deaths annually (Tate *et al.*, 2012). RVA has a double-stranded RNA genome made of 11 segments which encode six structural proteins (VP1–VP4, VP6 and VP7) and six non-structural proteins (NSP1–NSP6) (Estes & Kapikian, 2007). Based on the constellation

of segments composing the genome, three human genotypes have been established: Wa-like (genotype constellation 1), DS-1-like (genotype constellation 2) and AU-1-like (genotype constellation 3) (Matthijnsens *et al.*, 2008a; Nakagomi & Nakagomi, 1989). RVA strains are classified into G- and P-types based on genetic and antigenic diversity of the two outer capsid proteins, VP7 (G-genotypes) and VP4 (P-genotypes), respectively (Estes & Kapikian, 2007). To date, 27 different G- and 37 P-genotypes have been described in humans and animals (Matthijnsens *et al.*, 2011; Trojnar *et al.*, 2013). The majority of RVAs infecting humans are: G1P[8], G2P[4], G3P[8], G4P[8], G9P[8] and G12P[8] (Bányai *et al.*, 2012; Santos & Hoshino, 2005). The segmented nature of the RVA genome enables assortment of different individual gene segments of parental virus strains during mixed infections, generating reassortant virus strains. In 2008, the Rotavirus Classification Working Group (RCWG)

The GenBank/EMBL/DDBJ accession numbers for the nine G2P[4] RVA complete genome sequences determined in this work are KC178763–KC178771 (for VP1), KC178772–KC178780 (VP2), KC178781–KC178789 (VP3), KC178790–KC178798 (VP4), KC178799–KC178807 (VP6), KC178808–KC178816 (VP7), KC178718–KC178726 (NSP1), KC178727–KC178735 (NSP2), KC178736–KC178744 (NSP3), KC178745–KC178753 (NSP4) and KC178754–KC178762 (NSP5).

A supplementary figure is available with the online version of this paper.

proposed a full genome-based classification scheme allowing the differentiation of genotypes for the 11 RVA RNA segments, providing an ideal platform for deciphering the complex genetic diversity of RVAs (Matthijssens *et al.*, 2008a,b).

Although the global prevalence rate of G2P[4] strains is lower than that of G1P[8] strains, G2P[4] RVAs are recognized as a common cause of viral diarrhoea in humans, frequently reaching an epidemiological predominance in some geographical areas (Antunes *et al.*, 2009; Chouikha *et al.*, 2011; Dey *et al.*, 2009; Doan *et al.*, 2011; Iturriza-Gómara *et al.*, 2011; Kirkwood *et al.*, 2009; Paul *et al.*, 2008; Santos & Hoshino, 2005; Todd *et al.*, 2010). Notably, increased circulation of G2P[4] strains has been signalled after the introduction of RVA vaccination in several countries (Gurgel *et al.*, 2007; Nakagomi *et al.*, 2008; Zeller *et al.*, 2010), although epidemiological investigations in South America have revealed that an increase in the circulation of G2P[4] RVA strains also occurred in the same time frames in countries in which no universal RVA vaccination programmes were established (Gurgel *et al.*, 2007; Matthijssens *et al.*, 2009, 2012). Although it is very likely that vaccination will have some effect on the genotype distribution of circulating RVA strains, it is unclear what the long-term effects of this vaccine-derived selective pressure will be compared with other factors shaping natural genotype fluctuations (Matthijssens *et al.*, 2012).

VP7 sequence analysis of G2 RVAs detected over a 35-year period revealed that G2 VP7 lineages follow temporal patterns of evolution. The oldest G2 strains, sharing the same lineage (G2-I) as the prototype strain DS-1, were not detected after 1990, while a lineage of G2 RVAs (G2-II) circulated for almost 15 years between 1991 and 2005. In the 1990s a novel variant of a distinct lineage emerged (IVa-1) and spread all over the world, followed by another lineage IV variant (IVa-3) in the 2000s. Virtually all the G2 RVAs detected in the last decade

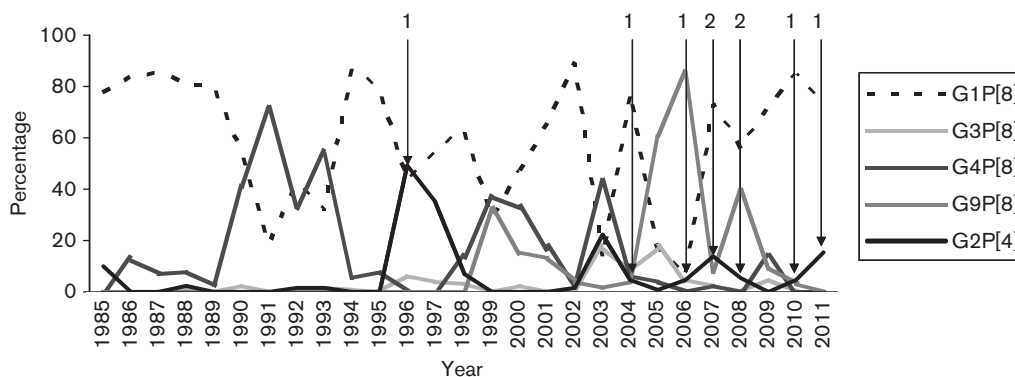
(2001–2009) belong to those two variants (Doan *et al.*, 2011).

RVA surveillance in Palermo, Italy, has been conducted uninterruptedly since 1985, revealing yearly prevalence rates of RVAs ranging from 20% to 50% among children aged <5 years hospitalized with gastroenteritis (Arista *et al.*, 1986, 1990, 1997, 2003, 2004, 2005; De Grazia *et al.*, 2007). In the 26-year period 1985–2011, the prevalence of G2P[4] RVAs in the paediatric population of Palermo showed yearly fluctuations (Arista *et al.*, 2005; De Grazia *et al.*, 2007), with peaks in 1985, 1996 to 1998, 2003 and 2004, 2006 to 2008, 2010 and 2011 (Fig. 1). Interestingly, the G2 strains identified between 2002 and 2004 did not react with the G2 type-specific monoclonal antibody (mAb) RV5:3, which was related to a change (D to N) in antigenic region A at residue 96 (Arista *et al.*, 2005) and is a hallmark of all recent G2 RVAs (lineage IVa) (Doan *et al.*, 2011). In order to explore in more detail the observed antigenic/genetic diversity in G2P[4] RVAs and to understand whether these changes were limited to the outer capsid proteins or also affected the overall genetic structure, the complete genomes of nine selected G2P[4] RVA strains isolated in Palermo, Italy, between 1996 and 2011 were determined.

## METHODS

**Sample collection.** Uninterrupted surveillance on RVA circulation has been conducted in Palermo, southern Italy, for 26 years (1985–2011). Stool samples were collected from children under 5 years admitted at the ‘G. Di Cristina’ Children’s Hospital of Palermo for acute gastroenteritis. Since the rotavirus season in Palermo is generally delayed with respect to the usual winter peak and includes the end of winter and spring (the peak circulation is in March and April), this paper is written using years instead of seasons.

**Nucleic acid extraction and G- and P-genotyping by semi-nested PCR.** Viral RNA was extracted from 10% faecal suspension using a QIAamp Viral RNA mini kit (Qiagen), according to the manufacturer’s instructions.



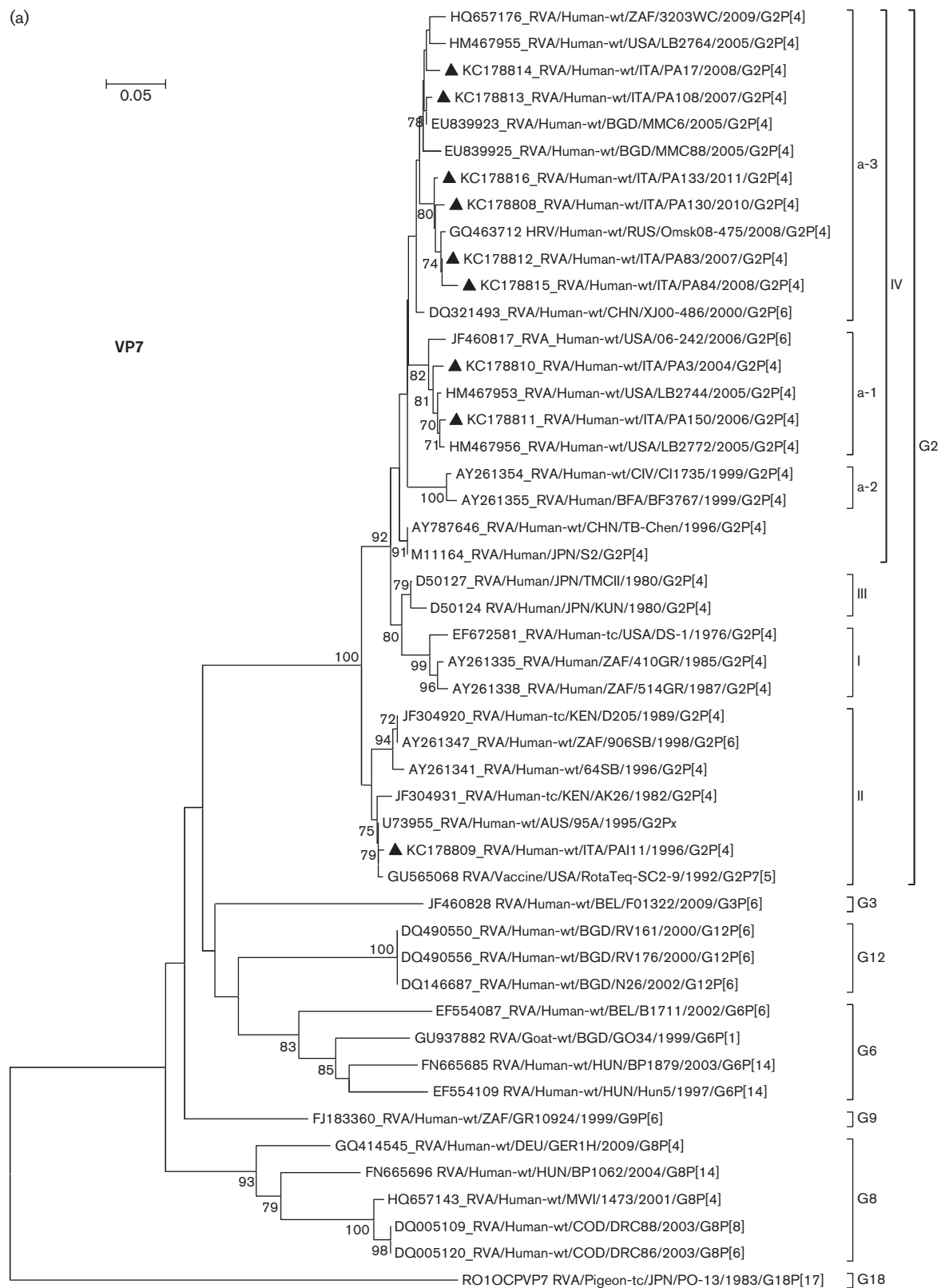
**Fig. 1.** Circulation of RVA genotypes in Palermo, Italy, from 1985 to 2011. Arrows indicate the year of collection and number of G2P[4] isolates selected for whole genome analysis.

**Table 1.** Comparison of the genomic constellations of the Italian G2P[4] RVA strains analysed in this study (in bold) and other RVA strains belonging to the DS-1 genogroup

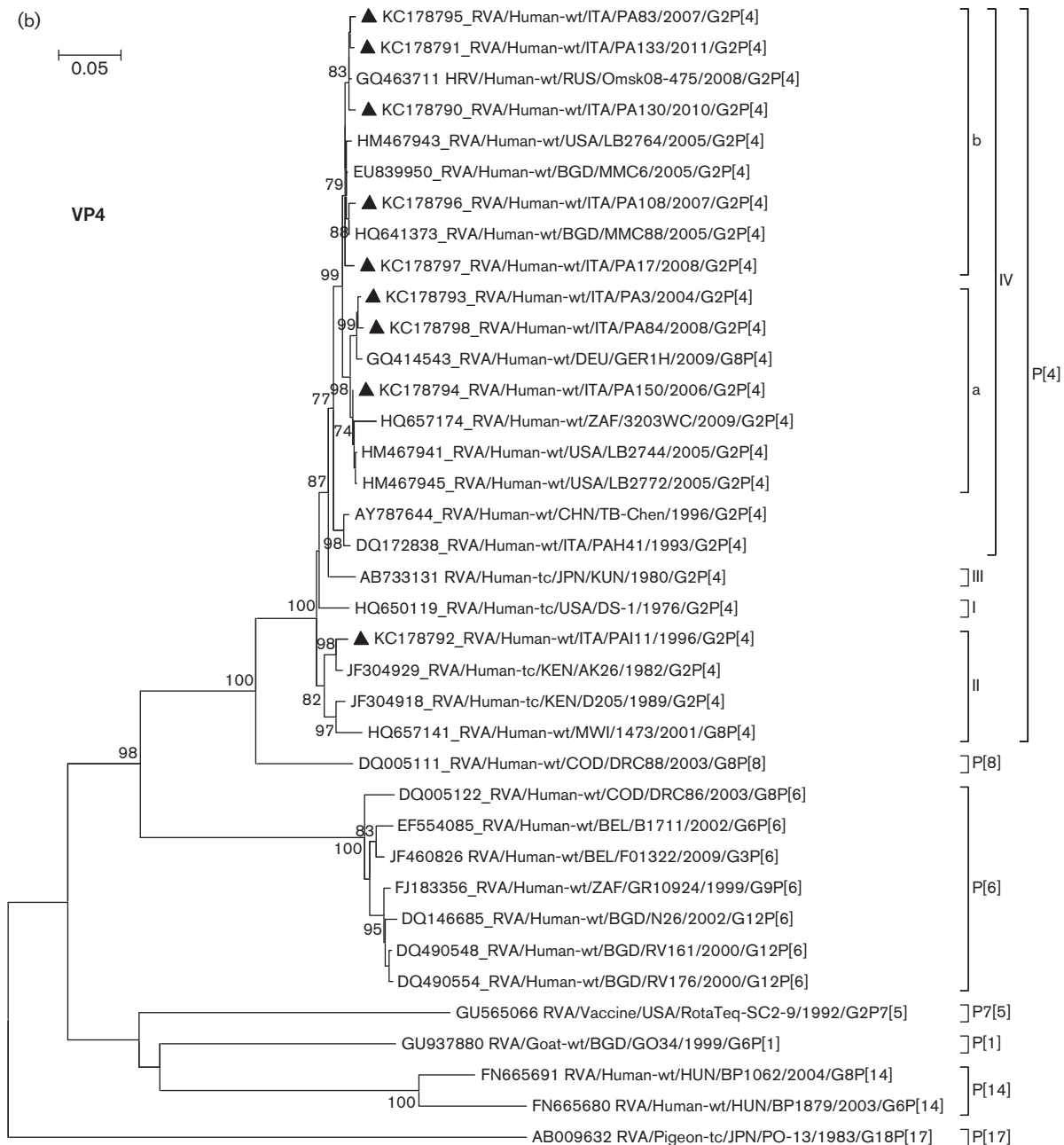
Colour codes indicate lineage attribution as described in Fig. 2. Gold is used for segments belonging to the lineage constellation of the ancestor DS-1 strain, violet for those belonging to the PAI11/1996 lineage constellation and red for those belonging to the lineage constellation of the G2P[4] RVA strains isolated in Palermo in the 2000s. Orange, yellow, purple and pink shades indicate genotype 2 segments belonging to human RVAs but not related to the three previously described lineage constellations. Different blue and green shades indicate segments of animal origin. Grey indicates the genotype 2 segments typical of the vaccine strain SC2-9.

		VP7	VP4	VP6	VP1	VP2	VP3	NSP1	NSP2	NSP3	NSP4	NSP5	
RVA/Human-tc/USA/DS-1/1976/G2P1B[4]	Human	G2	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	<b>Human</b>
RVA/Human-tc/KEN/AK26/1982/G2P[4]	Human	G2	P[4]	I2	R2	C2	M2	A2	N1	T2	E2	H2	
RVA/Human-tc/KEN/D205/1989/G2P[4]	Human	G2	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/AUS/95A/1995/G2Px	Human	G2											
RVA/Human-wt/CHN/TB-Chen/1996/G2P[4]	Human	G2	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/ZAF/64SB/1996/G2P[4]	Human	G2											
<b>RVA/Human-wt/ITA/PAI11/1996/G2P4</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N1</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
RVA/Human-wt/ZAF/906SB/1998/G2P[6]	Human	G2											
RVA/Human-wt/ZAF/GR10924/1999/G9P[6]	Human	G9	P[6]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/BGD/RV161/2000/G12P[6]	Human	G12	P[6]	I2	R2	C2	M2	A2	N2	T2	E1	H2	
RVA/Human-wt/MWI/1473/2001/G8P[4]	Human	G8	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/BEL/B1711/2002/G6P[6]	Human	G6	P[6]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/BGD/N26/2002/G12P[6]	Human	G12	P[6]	I2	R2	C2	M2	A2	N1	T2	E6	H2	
RVA/Human-wt/COD/DRC88/2003/G8P[8]	Human	G8	P[8]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
<b>RVA/Human-wt/ITA/PA3/2004/G2P[4]</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N2</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
RVA/Human-wt/BGD/MMC88/2005/G2P[4]	Human	G2	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/USA/LB2772/2005/G2P[4]	Human	G2	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/BGD/MMC6/2005/G2P[4]	Human	G2	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
<b>RVA/Human-wt/ITA/PA150/2006/G2P[4]</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N2</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
RVA/Human-wt/IND/265/2007/G10P[6]	Human										E2		
<b>RVA/Human-wt/ITA/PA83/2007/G2P[4]</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N2</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
<b>RVA/Human-wt/ITA/PA108/2007/G2P[4]</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N2</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
<b>RVA/Human-wt/ITA/PA84/2008/G2P[4]</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N2</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
<b>RVA/Human-wt/ITA/PA17/2008/G2P[4]</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N2</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
RVA/Human-wt/RUS/Omsk08-475/2008/G2P[4]	Human	G2	P[4]	I2							E2		
RVA/Human-wt/IND/UP-BE-1/2009/GxPx	Human										E2		
RVA/Human-wt/DEU/GER1H/2009/G8P[4]	Human	G8	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/ZAF/3203WC/2009/G2P[4]	Human	G2	P[4]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
RVA/Human-wt/BEL/F01322/2009/G3P[6]	Human	G3	P[6]	I2	R2	C2	M2	A2	N2	T2	E2	H2	
<b>RVA/Human-wt/ITA/PA130/2010/G2P[4]</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N2</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
<b>RVA/Human-wt/ITA/PA133/2011/G2P[4]</b>	<b>Human</b>	<b>G2</b>	<b>P[4]</b>	<b>I2</b>	<b>R2</b>	<b>C2</b>	<b>M2</b>	<b>A2</b>	<b>N2</b>	<b>T2</b>	<b>E2</b>	<b>H2</b>	
RVA/Human-wt/HUN/Hun5/1997/G6P[14]	Human	G6	P[14]	I2	R2	C2	M2	A11	N2	T6	E2	H3	<b>Animal or animal-derived</b>
RVA/Goat-tc/BGD/GO34/1999/G6P[1]	Goat	G6	P[1]	I2	R2	C2	M2	A11	N2	T6	E2	H3	
RVA/Human-wt/HUN/BP1879/2003/G6P[14]	Human	G6	P[14]	I2	R2	C2	M2	A11	N2	T6	E2	H3	
RVA/Human-wt/HUN/BP1062/2004/G8P[14]	Human	G8	P[14]	I2	R2	C2	M2	A11	N2	T6	E2	H3	
RVA/Cow-wt/IND/MP/B-47/2008/GxPx	Cow										E2		
RVA/Cow-wt/IND/MP-B-100/2008/GxPx	Cow										E2		
RVA/Vaccine/USA/RotaTeq-SC2-9/1992/G2P7[5]	Human/cow	G2	P7[5]	I2	R2	C2	M1	A3	N2	T6	E2	H3	<b>Vaccine</b>

G2P[4] rotaviruses complete genome analyses



**Fig. 2.** Phylogenetic trees based on the full-length nucleotide sequences of RVA VP7 (a), VP4 (b), VP6 (c), VP1 to VP3 (d–f), and NSP1 to NSP5 (g–k) genes. Bootstrap values (1000 replicates) above 69% are shown at branch nodes. The number of nucleotide substitutions per site is indicated by the scale bars. The strains analysed in this study are marked with a triangle. VP7 lineages and sublineages of G2 strains and VP4 lineages of P[4] strains are indicated according to Doan *et al.* (2011, 2012). NSP2 lineages within the N1 genotype are indicated according to Donker *et al.* (2011).



**Fig. 2.** (continued)

The extracted RNA was reverse transcribed as previously described (Iturriza-Gomara *et al.*, 1999). Rotavirus G- and P-genotyping was performed using semi-nested multiplex PCRs (Gentsch *et al.*, 1992; Gouvea *et al.*, 1990; Iturriza-Gómara *et al.*, 2004; Martella *et al.*, 2004). All strains typable by reverse transcription-polymerase chain reaction (RT-PCR) were further investigated by direct sequencing of VP7 and VP4 for phylogenetic analysis.

**Whole genome sequencing and phylogenetic analysis.** A selection of G2P[4] strains representative of the surveillance period, for which a sufficient amount of the original faecal sample was available, was subjected to whole genome analysis (Fig. 1). The complete genome

sequencing, including the 5' and 3' termini of each genome segment, of the selected RVA strains was performed as described elsewhere (Matthijnssens *et al.*, 2006). The genotypes of the G2P[4] RVA strains were determined according to the genotyping recommendations of the RCWG using the RotaC online classification tool (<http://rotac.regatools.be>) (Maes *et al.*, 2009). Cognate genome sequences were individually compared with whole genome RVA sequences available in GenBank. When reassortment was suspected, the most similar sequences available in GenBank were also added to the corresponding tree even if sequences of other gene segments were not available. Phylogenetic analysis was performed using the software MEGA5 (Tamura *et al.*, 2011). Phylogenetic trees were constructed using the neighbour-joining method and

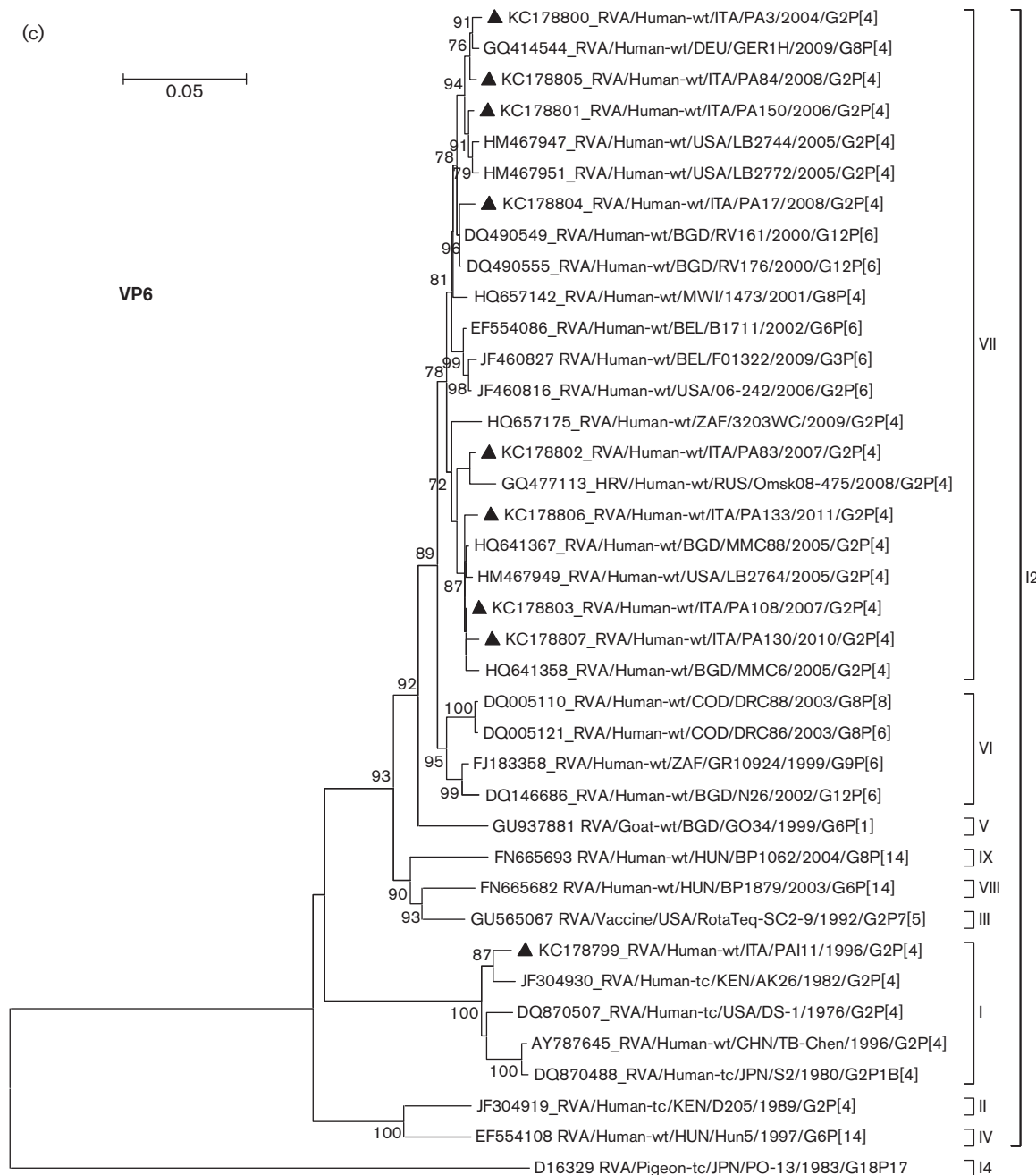


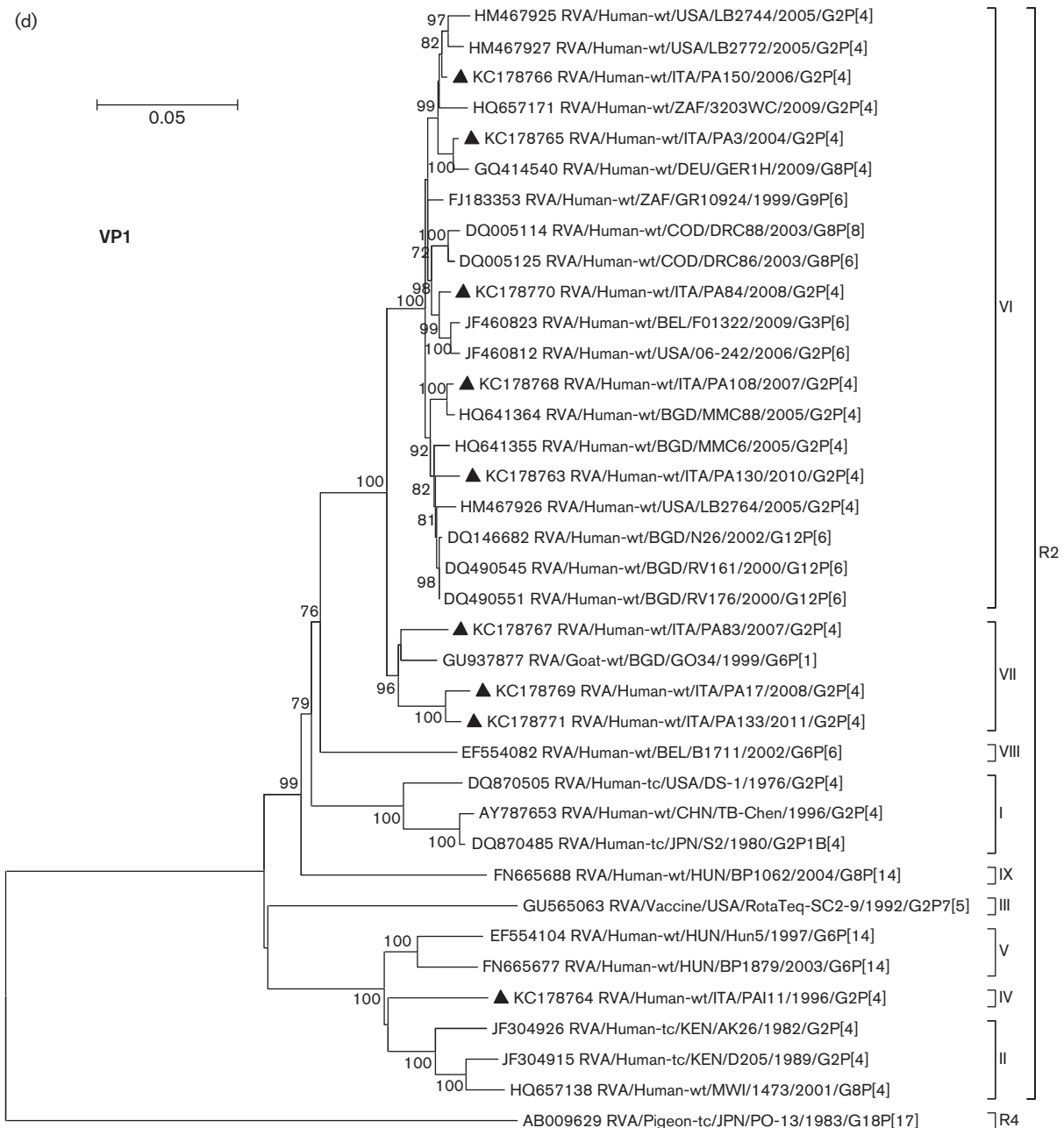
Fig. 2. (continued)

Kimura's two-parameter model with 1000 bootstrap replicates. Lineage designation for the various genome segments followed the outlines of the existing literature and was substantiated by sequence comparison and phylogenetic analysis of our sequence datasets.

**Accession numbers.** GenBank accession numbers for each individual genomic segment are: KC178763–KC178771 (VP1); KC178772–KC178780 (VP2); KC178781–KC178789 (VP3); KC178790–KC178798 (VP4); KC178799–KC178807 (VP6); KC178808–KC178816 (VP7); KC178718–KC178726 (NSP1); KC178727–KC178735 (NSP2); KC178736–KC178744 (NSP3); KC178745–KC178753 (NSP4); KC178754–KC178762 (NSP5).

## RESULTS

Representative G2P[4] strains were selected on the basis of preliminary phylogenetic analysis of 38 partial VP7 sequences (>456 nt in length) detected between 1993 and 2011. We were not able to produce sequences from strains from before 1993 (Fig. S1, available in JGV Online). Nine G2 RVA strains, detected in seven different years of increased G2P[4] circulation, 1996 (PAI11/1996), 2004 (PA3/2004), 2006 (PA150/2006), 2007 (PA83/2007 and PA108/2007), 2008 (PA17/2008 and PA84/2008), 2010



**Fig. 2.** (continued)

(PA130/2010) and 2011 (PA133/2011), were chosen for complete genome analysis. Although the same VP7 gene can be found in different genetic backgrounds, the nine strains were selected as representative of the various VP7 lineages, and of clusters within different lineages, as detected over the 1993–2011 surveillance period (Fig. S1). Once complete genome sequences were obtained, the genotype for each of the 11 gene segments was defined using the online RVA classification tool RotaC at <http://rotac.regatools.be> (Maes *et al.*, 2009). This analysis

allowed unambiguous genotype assignment of all Italian G2P[4] strains as G2-P[4]-I2-R2-C2-M2-A2-N2-T2-E2-H2, except for strain PA11/1996, which possessed an N1 NSP2 genotype. Complete genotype assignments of the Italian strains, together with those of other representative G2P[4] RVA strains, including reference strain DS-1, are shown in Table 1.

Phylogenetic trees of all 11 genome segments were constructed (Fig. 2). Overall, the VP1–VP4, VP6–VP7

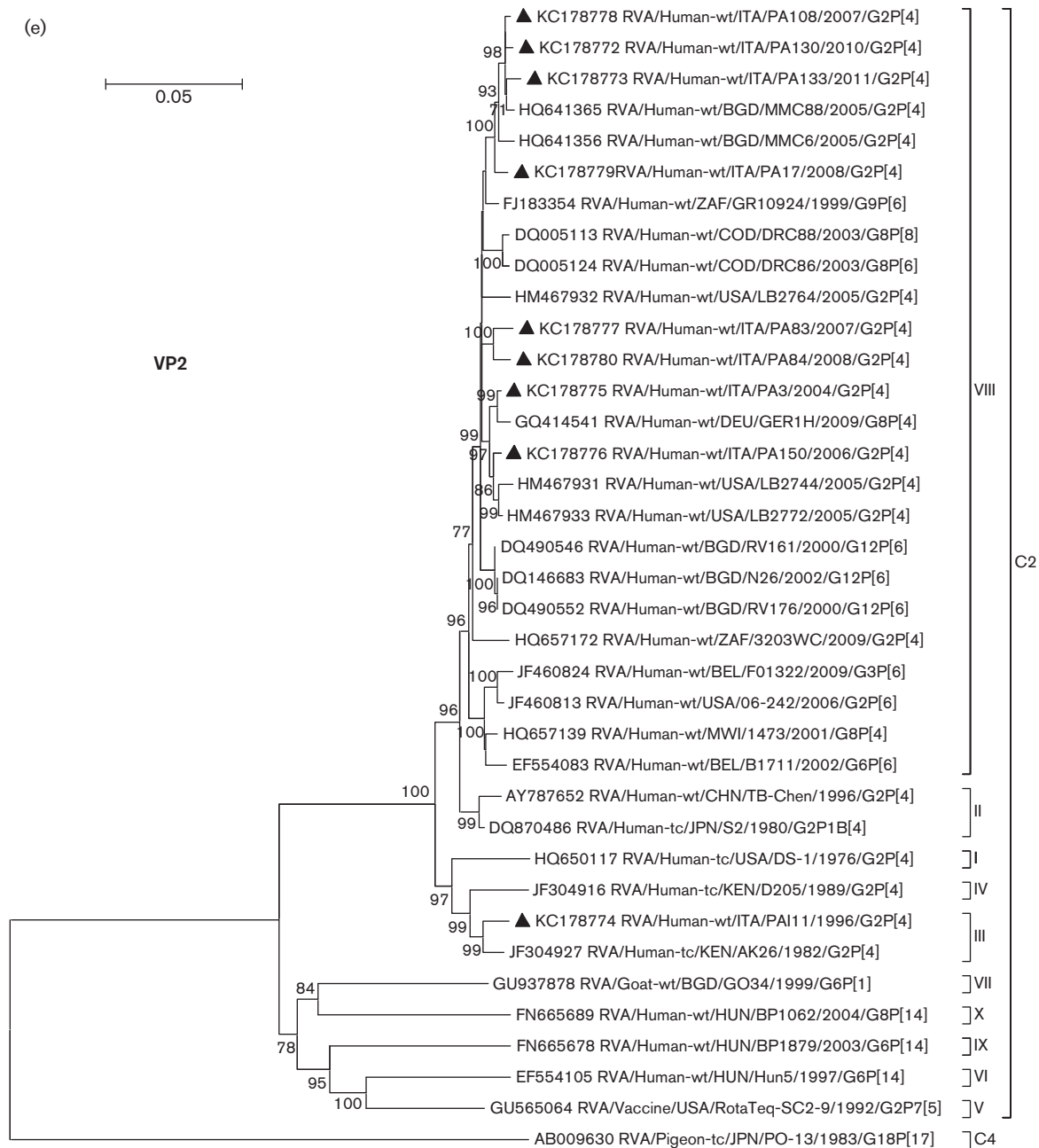
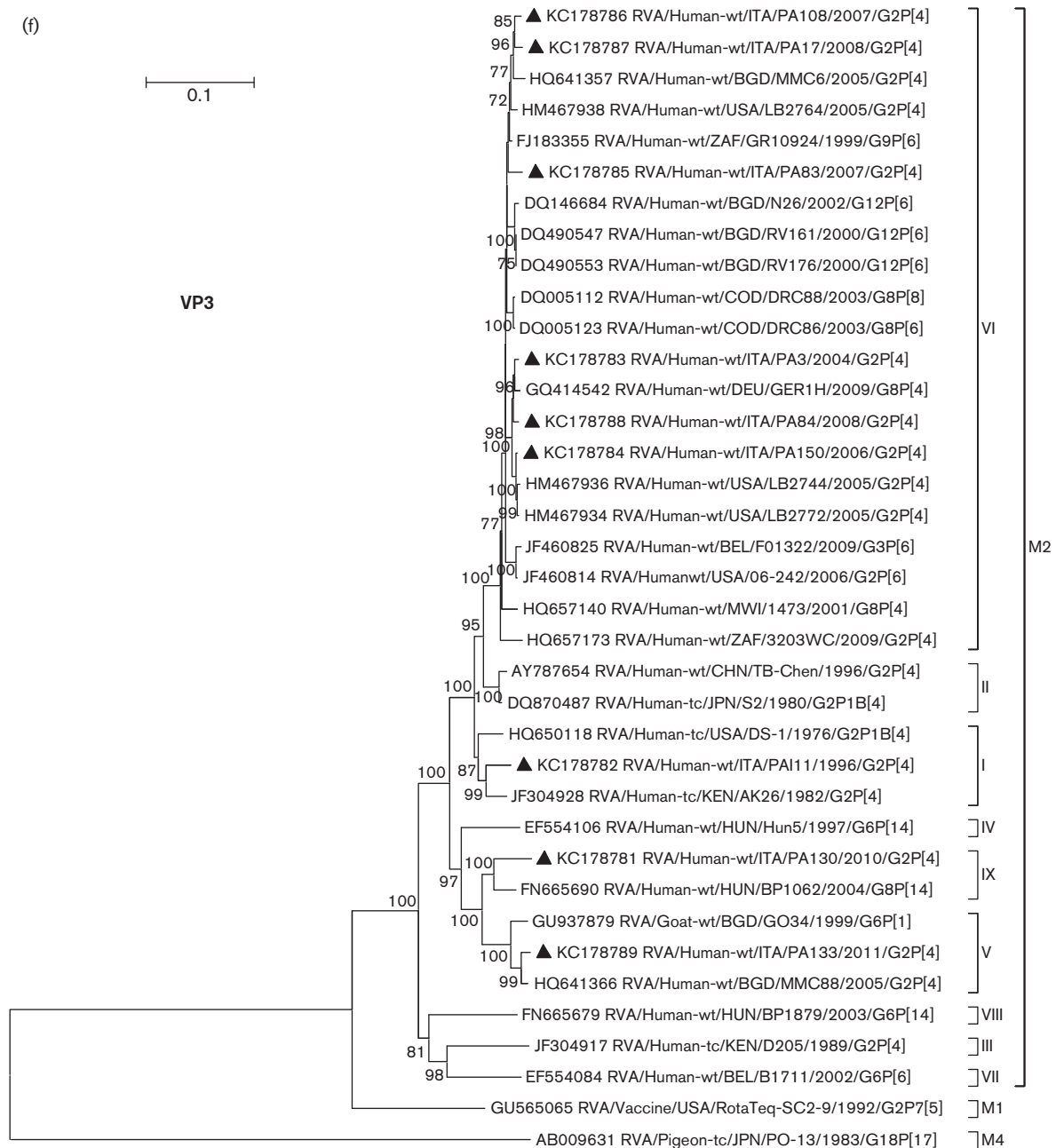


Fig. 2. (continued)

and NSP1–NSP5 gene segments of the Italian G2P[4] strains showed a variable degree of sequence identity, ranging from >81.6% (at nt level) for NSP2 to >95% for NSP3. As a general rule, for each genome, segment lineages were defined ad hoc based on observed clustering patterns in the phylogeny and indicated with roman numerals, but for VP7, VP4 and NSP2 N1 genotypes the same lineage designations were used as defined by Doan *et al.* (2011, 2012) and Donker *et al.* (2011), respectively (Fig. 2). Lineage attribution is shown in Table 1 using colour coding to distinguish segments belonging to different lineages

from each other and indicating the main lineage constellations. According to the proposed lineage segregation, none of the Italian strains matched the genetic pattern of the prototype DS-1 strain isolated in the 1970s. Also, in the VP7 tree all the analysed viruses, except for strain PAI11/1996, were only distantly related to the G2-reassortant vaccine strain SC2-9. Strain PAI11/1996 showed a close phylogenetic clustering with AK26 Kenyan strain in all genome segments with the exception of VP1, NSP2 and NSP4 (Fig. 2). Several genome segments of PAI11/1996 were also related to another Kenyan strain, D205 (VP2,





**Fig. 2.** (continued)

VP4, VP7 and NSP5), to the Malawian strain MWI/1473 (VP4) and, in VP7, to G2 strains from South Africa (64SB and 906SB) and Australia (95A) (Table 1). For the other eight Italian G2P[4] strains, detected between 2004 and 2011, all genome segments clustered separately from those of strain PAI11/1996. Strains PA3/2004 and PA150/2006 formed a novel lineage constellation, which was shared also by RVA strains detected in different continents – MMC6 from Bangladesh, 3203WC from South Africa and LB2772 from the USA – suggesting a worldwide circulation of this lineage constellation within the DS-1-like RVA strains. The

most recent Italian strains, PA83/2007, PA108/2007, PA17/2008, PA84/2008, PA130/2010 and PA133/2011, also retained a lineage constellation very similar to the former, although exhibiting one or more segments of different origin. Strains PA83/2007, PA17/2008 and PA133/2011 share their VP1 and VP3 reassortant segments with the caprine strain GO34 and the reassortant human strain MMC88, both from Bangladesh (Fig. 2d, f, Table 1). Strain PA130/2010 displayed a VP3 similar to the Hungarian animal-like human RVA strain BP1062 (Fig. 2f, Table 1) (Bányai *et al.*, 2010). The 2004–2011 G2P[4] strains

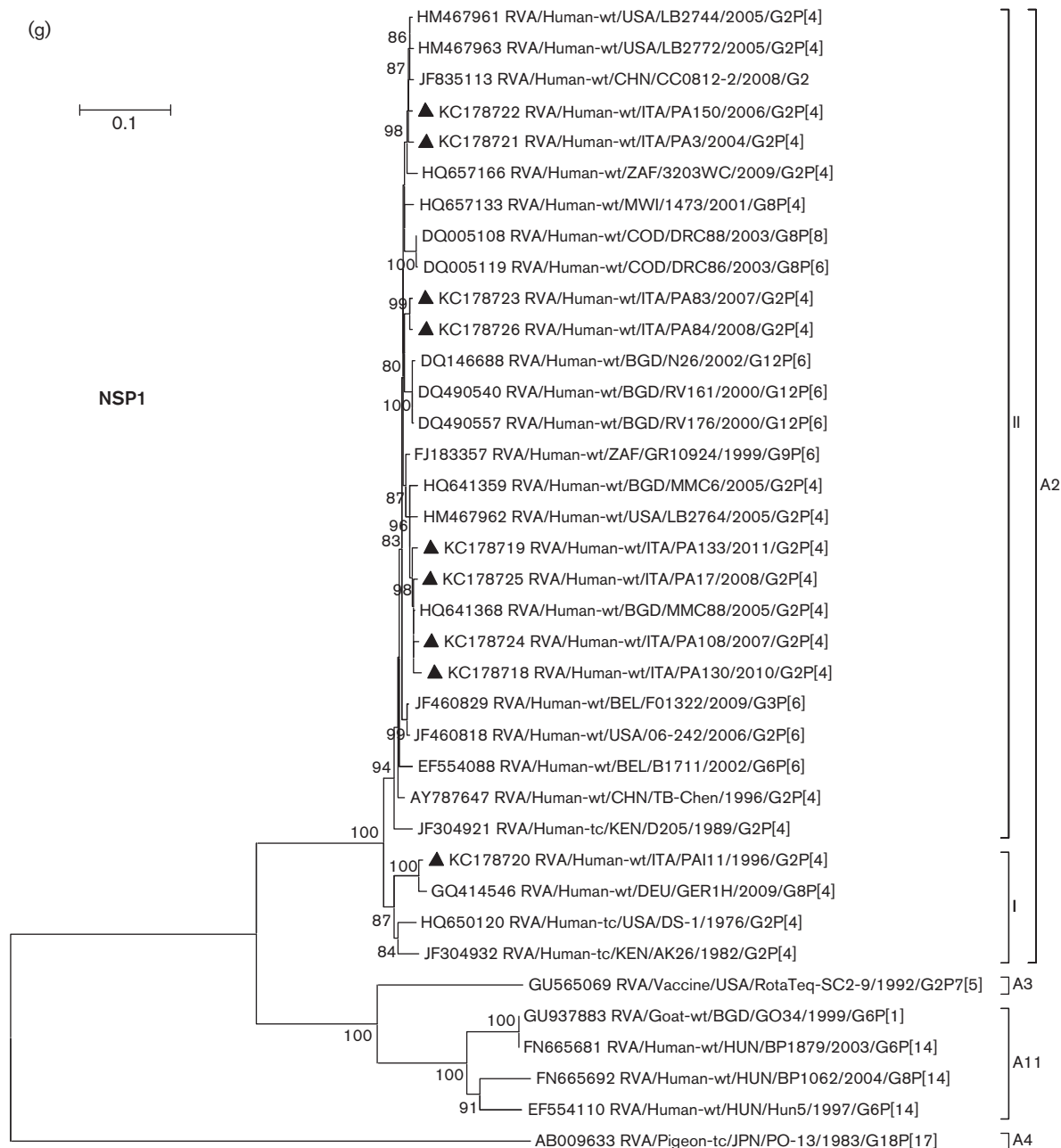
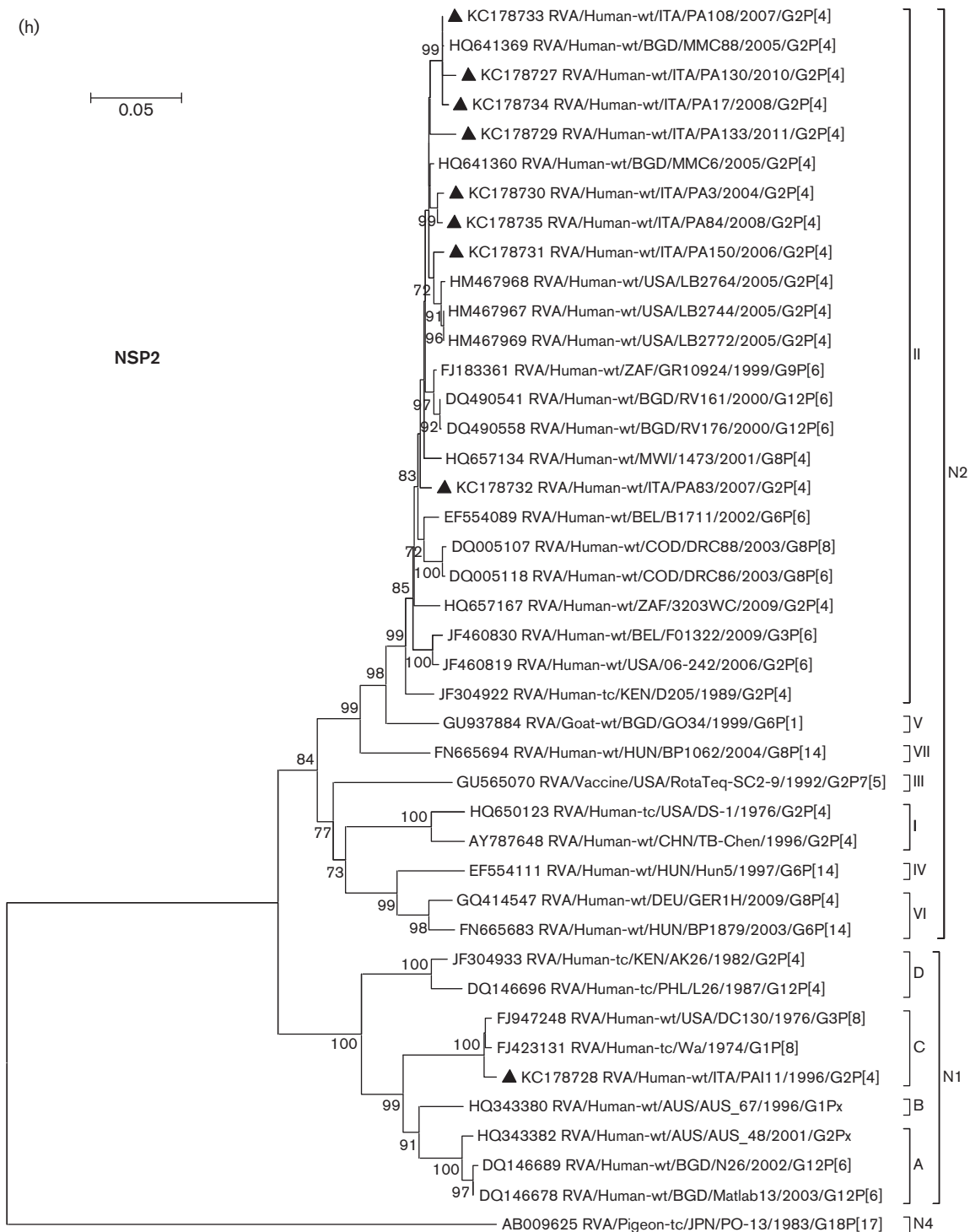


Fig. 2. (continued)

segregated into three different lineages within the NSP4 E2 genotype (Fig. 2j, Table 1), with strains PA3/2004, PA150/2006 and PA83/2007 clustering together with contemporary G2P[4] RVA strains detected all over the world and with TB-Chen strain isolated in China in 1996, strain PA84/2008 and PA133/2011 clustering with human RVAs from central and eastern Europe, and strains PA108/2007, PA17/2008 and PA130/2010 segregating with human and ruminant RVAs from India.

In the VP7 tree, strain PAI11/1996 clustered in the G2-II lineage, while the 2004–2011 RVAs clustered into two

G2-IV sublineages, diverging by at least 3.1% (nt) from each other, and including Italian viruses detected in 2004–2006 (sublineage IVa-1) and in 2007–2011 (sublineage IVa-3), respectively (Fig. 2a), suggesting temporally related patterns of segregation. When the VP7 amino acid alignment (Fig. 3) was inspected in detail, strain PAI11/1996 was seen to differ only in three residues from the G2 vaccine strain SC2-9 (lineage II) and in 16 residues from the prototype G2 strain DS-1 (lineage I), with most changes (3/3 and 10/16, respectively) accumulating within the variable regions (VRs). The 2004–2011 G2 Italian RVAs



**Fig. 2.** (continued)

(lineage IV) possessed conserved amino acid differences at 11 residues (seven in the VRs) when compared with strain PA11/1996 and 10 conserved different residues (seven in VRs) when compared with DS-1. In addition, several scattered variable amino acid mutations were observed.

The two G2-IV sublineages (IVa-1 and IVa-3) possessed conserved differences at residues in VR1 (position 15) and VR9 (position 242). Four amino acid changes were observed throughout the VP7 antigenic epitopes (Aoki *et al.*, 2009) among the Italian G2P[4] strains and with respect to DS-1

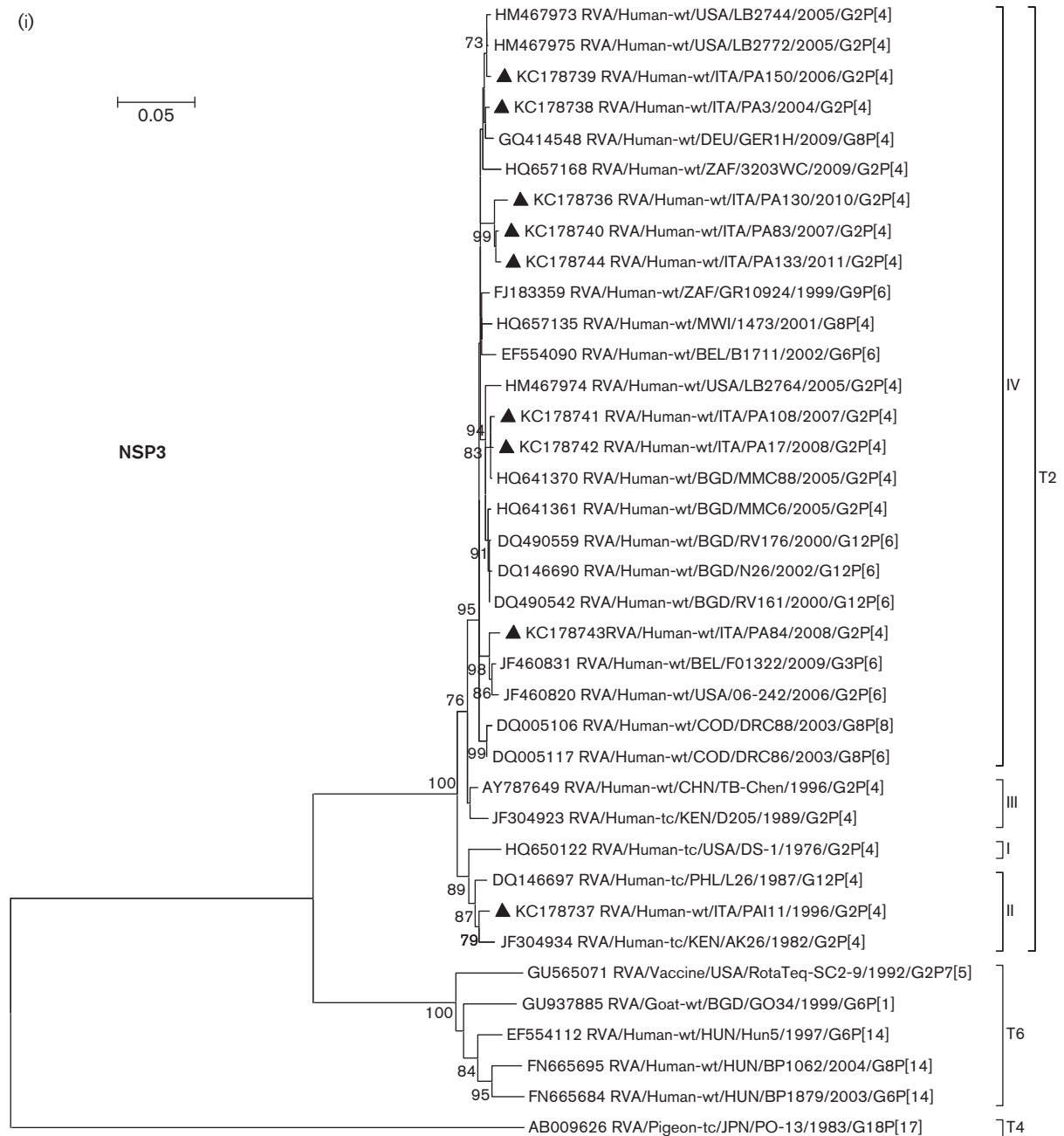
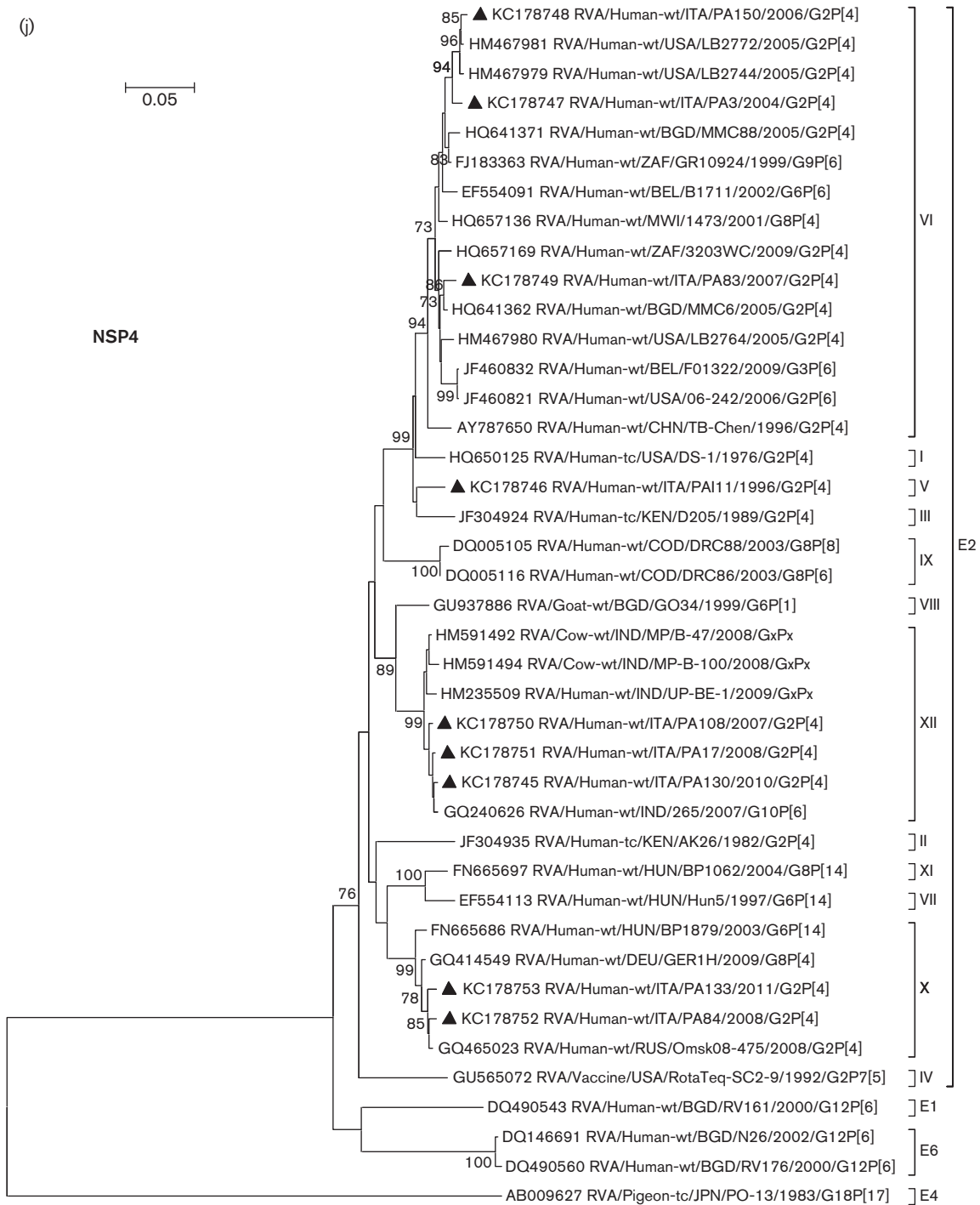


Fig. 2. (continued)

and SC2-9 strains, specifically at positions 87 and 96 in domain 7-1a, 213 in 7-1b and 242 in 7-2, corresponding to the four amino acids previously recognized as potential epitopes for G2 strains to be distinguished in lineages and sublineages (Fig. 3) (Doan *et al.*, 2011).

In the VP4-based tree, strain PA111/1996 clustered in lineage P[4]-II, while the 2004–2011 RVAs clustered into two distinct P[4]-IV sublineages. Strains PA3/2004, PA150/2006 and PA84/2008 segregated into sublineage IVa, while

the majority of the more recent Italian strains (PA83/2007, PA108/2007, PA17/2008, PA130/2010 and PA133/2011) segregated into sublineage IVb (Fig. 2b). When the VP4 amino acid sequences were inspected, overall the G2P[4] strains isolated in Palermo during the surveillance period accumulated mutations at seven different amino acid positions in the antigenic epitopes of VP8\* and VP5\* with respect to the ancestor DS-1 strain (lineage I) (Figs 4 and 5). Lineage II strain PA111/1996 showed two differences



**Fig. 2.** (continued)

from DS-1 in the 8-1 epitope and one in the 8-4 epitope, and three changes in VP5\* epitope 5-1. Five of the amino acid differences detected in PAI11/1996 were maintained in sublineage IVa and IVb strains from Palermo which also differed from DS-1 at residue 133 in epitope 8-3 (N133S), while position 192 reverted to A as in DS-1. A single

conserved difference was observed between the amino acid sequences of the IVa and IVb sublineages at position 607 (V607I), outside the VP8\* variable region (aa 72–203) and VP5\* epitopes. However, in the VP8\* variable region, residue 162 was either R or G in sublineage IVa sequences but either R or K in sublineage IVb.

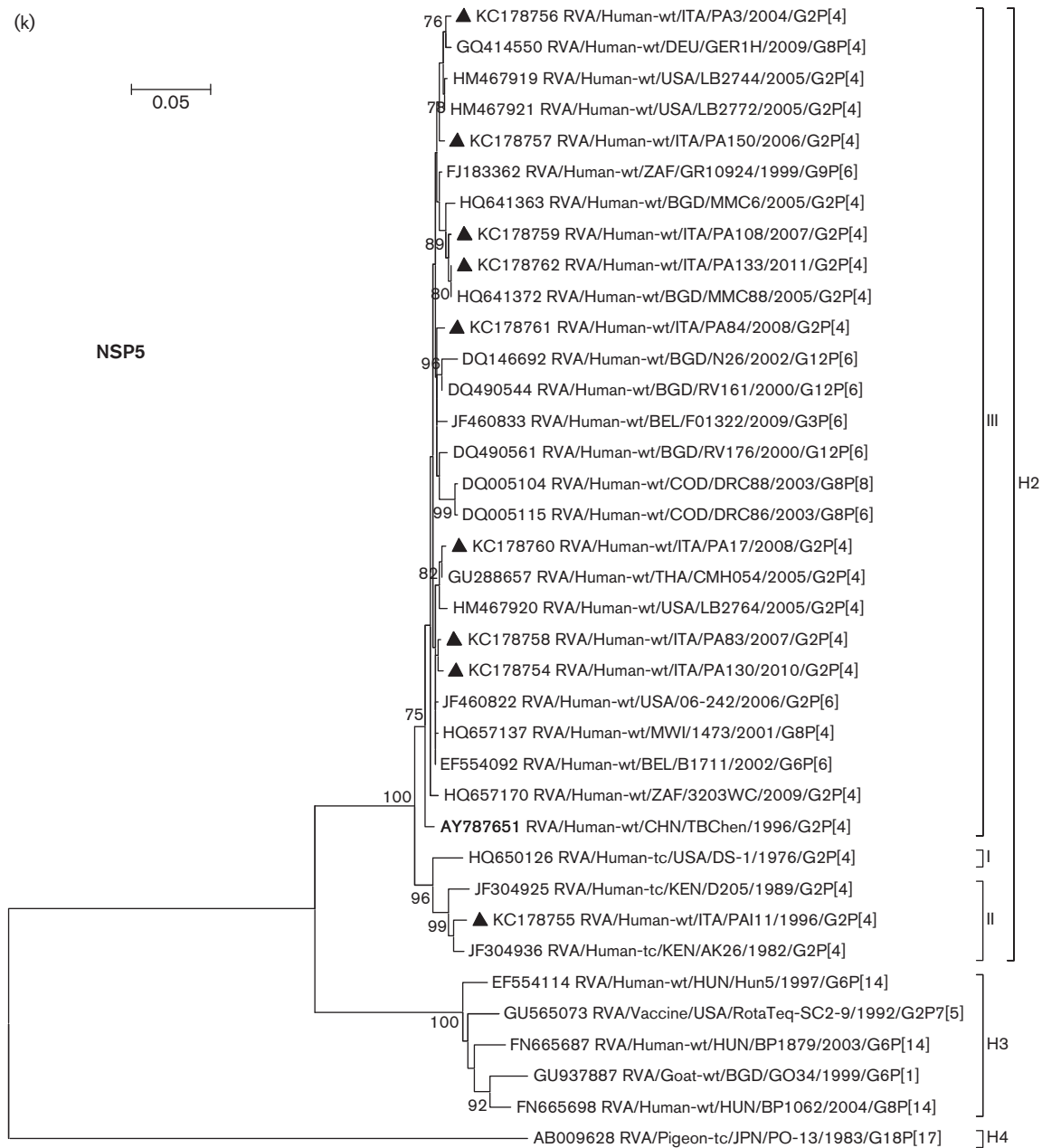
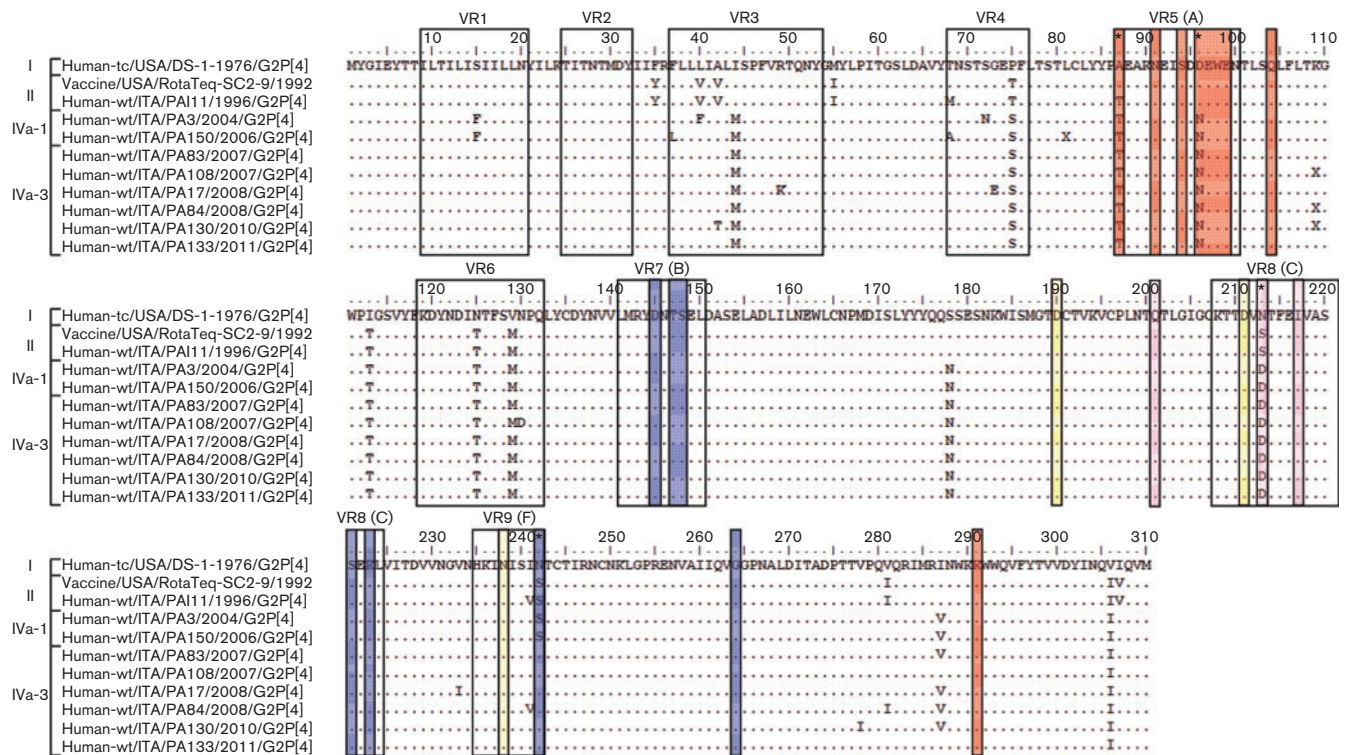


Fig. 2. (continued)

## CONCLUSIONS

During a 26-year (1985–2011) surveillance in Palermo, Italy, the prevalence of G2P[4] RVAs appeared to vary yearly. An increased prevalence of G2P[4] RVAs was monitored in 1985 (9.7%), 1996 to 1998 (49%, 35.3% and 6.7%, respectively), 2003 (22.6%), 2004 (4.9%), 2006 to 2008 (4.3%, 14% and 5.6%, respectively), 2010 (5%) and 2011 (15.4%). The complete genomes of nine G2P[4] RVA strains representative of seven different seasons with increased circulation of G2 RVAs and collected between 1996 and 2011 were used to explore their genetic diversity

in more detail. Based on VP7 phylogeny (Fig. 2a) and sequence analysis of the VP7 antigenic regions (Fig. 3), the old strain PAI11/1996 was assigned to lineage II, while the 2004–2006 strains and the 2007–2011 RVAs clustered in sublineage IVa-1 and sublineage IVa-3, respectively. In-depth data mining of the sequence databases has revealed that RVAs of lineage G2-II were circulating mostly during the 1990s, reaching a global distribution, with the exception of Asia (Doan *et al.*, 2011). RVAs of lineage G2-IV dominated globally in the 2000s, with two distinct sublineages, IVa-1 and IVa-3, emerging consecutively. The two sublineages differ in their VP7 amino acid

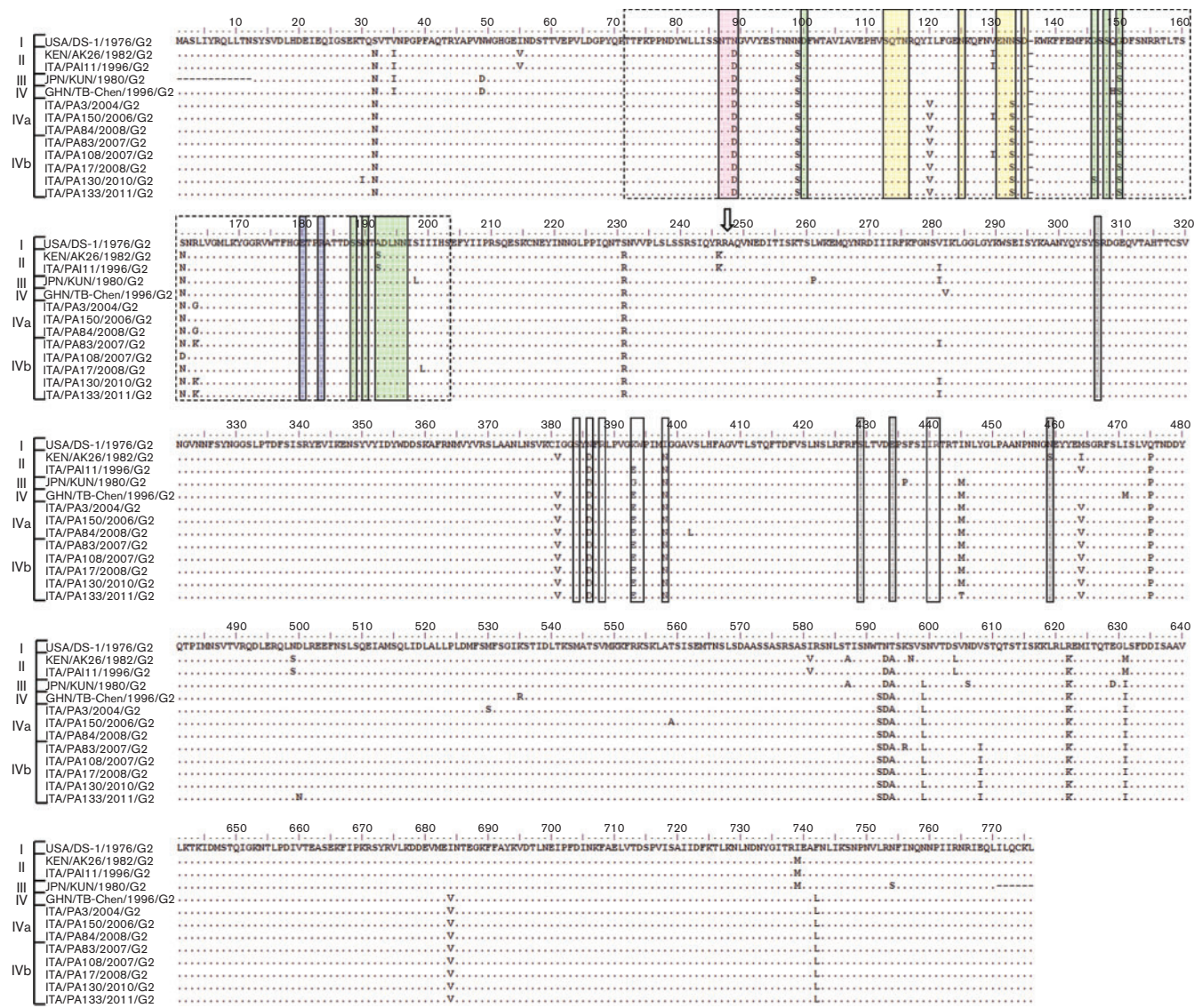


**Fig. 3.** Deduced amino acid sequences of the VP7 genes of Italian G2P[4] strains compared with the DS-1 G2 reference and SC2-9 vaccine strain. Antigenic variable regions (VR) are boxed. Positions of antigenic epitopes 7-1a (red), 7-1b (pink) and 7-2 (blue) and glycosylation sites (yellow) are coloured. Dots indicate conserved positions. Lineages and sublineages within genotype G2 are indicated as defined by Doan *et al.* (2011) based on the amino acids at residues 87, 96, 213 and 242 (indicated by asterisks).

sequence at position 242 but share, at position 96, the asparagine residue that disrupts a G2-specific monoclonal epitope (Arista *et al.*, 2005; Doan *et al.*, 2011). In Italy, G2 RVAs of sublineage IVa-1 circulated from 2004 to 2006 and they were replaced by G2 RVAs of sublineage IVa-3 in the 2007–2011 period. A similar shift from sublineage G2-IVa-1 to G2-IVa-3 was also observed on a global scale approximately in the same period (Doan *et al.*, 2011; Gómez *et al.*, 2011; Mascarenhas *et al.*, 2010). Similarly, upon phylogenetic analysis of the VP4 gene (Fig. 2b) and sequence analysis of VP4 (Fig. 4), the old Italian strain PAI11/1996 was classified as a distinct lineage, P[4]-II, while the 2000s Italian strains were classified into two different sublineages that emerged consecutively, here named P[4]-IVa and -IVb. The emergence and spread of P[4] RVAs of lineage IV during the 2000s has also been documented elsewhere (Gómez *et al.*, 2011) and, on the basis of database interrogation, seems to have occurred worldwide. Accordingly, there is evidence that in the 2000s the shift from the VP7 sublineage G2-IVa-1 to G2-IVa-3 occurred in parallel with the shift from the VP4 sublineage P[4]-IVa to P[4]-IVb, leading to the global spread of a unique VP7/VP4 combination, G2-IVa-3/P[4]-IVb.

In this study, we tried to understand whether the onset and spread of recent G2P[4] RVAs was also associated with

reassortment events in their genome. Interestingly, upon whole genome analysis, the older strain PAI11/1996 displayed not only VP7/VP4 lineages distinct from contemporary G2P[4] RVAs, but also a distinct genomic lineage constellation. Strain PAI11/1996 was closely related to the ancient African strain AK26 in all genome segments except for NSP2 and NSP4. Strain AK26 was described as a DS-1-like inter-genogroup reassortant strain with a Wa-like NSP2 (N1). Also, the VP1 and NSP4 genes of this African strain appeared to originate from or share a common origin with RVA genes of artiodactyl (ruminant and camelid) origin (Ghosh *et al.*, 2011). However, the NSP2 gene of PAI11/1996 belonged to a different N1 lineage with respect to strain AK26. G2 RVAs with an N1 NSP2 are uncommon but they have been described in Japan in six consecutive rotavirus seasons, from 1985 to 1990 (Doan *et al.*, 2012). The prototype Japanese inter-genogroup reassortant G2P[4] strain (AU605) was closely related to strain TB-Chen, and compared with PAI11/1996 belonged to different lineages in all genome segments except VP6 (data not shown). These findings are consistent with the independent origin of the Italian heterotypic NSP2 mono-reassortant strain. Interestingly, in Japan the circulation of NSP2 mono-reassortant strains was observed in seasons when G2P[4] was not the most prevalent genotype



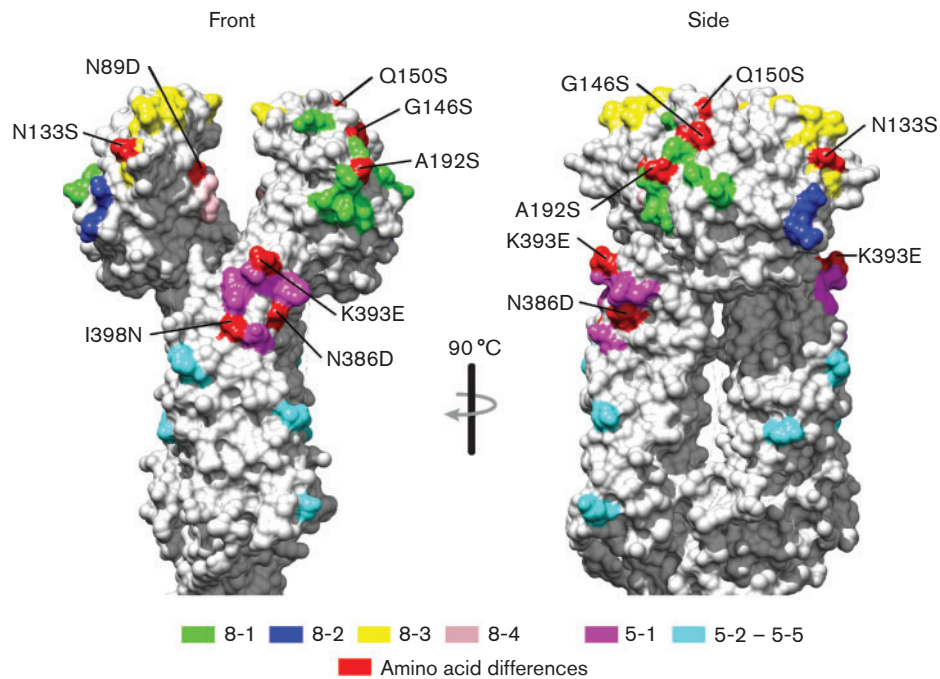
**Fig. 4.** Deduced amino acid sequences of the VP4 protein of P[4] strains belonging to different lineages and sublineages. Positions boxed are situated inside the VP8\* variable region (aa 72–203). Residue numbering is based on the RRV sequence AY033150. Colour coding is used for residues involved in neutralization escape in VP8\* (epitope 8-1, green; 8-2, blue; 8-3, yellow; 8-4, pink) and VP5\* (epitope 5-1, white; 5-2 [aa 434], 5-3 [aa 459], 5-4 [aa 429] and 5-5 [aa 306], grey).

(Doan *et al.*, 2012). Although we have no evidence that all the G2P[4] strains circulating in Palermo in 1996 were NSP2 mono-reassortants, VP7 sequencing data from multiple strains from the 1996 epidemic of G2P[4] suggest that PAI11/1996 represented the epidemic strains that were predominant in Palermo in 1996 (Figs 1 and S1).

All the contemporary G2P[4] strains (circulating from 2004 to 2011) belonged to a novel lineage constellation, distinct from the reference strain DS-1 and from the RVA strains with DS-1-like genotype constellations circulating in the 1990s. This novel 2000s lineage constellation also included strains identified from Bangladesh (MMC6), South Africa (3203WC) and the USA (LB2772), all of which were

circulating in the mid 2000s. Strain LB2772 was closely related to reassortant G3P[6] RVA strains infecting infants in Belgium, and to G2P[6] strains identified in the USA in 2006 (Heylen *et al.*, 2013). Interestingly, several DS1-like non-G2P[4] strains circulating between 1999 and 2003 were already displaying a similar set of genome segments, thus suggesting that these novel 2000s G2P[4] viruses were already circulating at the end of the 1990s. Starting from 2007, several of the Italian G2P[4] strains within this novel 2000s variant of the DS-1-like genotype constellation proved to be reassortant in one or more gene segments (VP1, VP3 and/or NSP4) with RVAs of potential artiodactyl origin. The number of intra-genotype reassortment





**Fig. 5.** Surface representation of the VP4 protein VP8\* and VP5\* antigenic regions. The VP4 model is based on RRV (PDB 3IYU). The right image is rotated 90° compared with the left image. Antigenic epitopes are coloured in green (8-1), blue (8-2), yellow (8-3), pink (8-4), purple (5-1) and cyan (5-2 to 5-5). Surface-exposed residues that differ between strains circulating in Palermo and the DS-1 strain are shown in red.

events in contemporary G2P[4] RVA strains suggests a certain tendency to acquisition of gene segments from animal RVAs when genome segments share the same genotype. From 2007 to 2011, VP1 and VP3 genes closely related with a caprine strain (GO34) from Bangladesh and NSP4 gene segments closely related to those of RVA strains identified from cows in India were detected in G2P[4] RVAs of the novel 2000s variant in Palermo. Similarly, several intra-genogroup reassortment events have been identified within G2P[4] strains co-circulating in the 2005–2006 season in the USA (Bányai *et al.*, 2011). The G2P[4] RVAs circulating in the earlier 2000s appear to be more homogeneous genetically, suggesting that the G2P[4] RVAs detected in the 2000s started to acquire novel gene segments via reassortment only some years after their global spread. Multi-centric investigations analysing a larger number of old and novel G2P[4] RVAs detected in the 2000s from different geographical areas could allow better explanation of the global evolution of G2P[4] RVAs in the last decades and eventually reveal additional gene polymorphisms.

In 2006, two RVA vaccines were licensed and they are now included as part of the routine vaccination schedule for infants in many countries around the world (Ciarlet & Schödel, 2009; Heaton & Ciarlet, 2007; Matthijnsens *et al.*, 2010). None of the currently licensed RVA vaccines contains strains with a complete DS-1-like genotype

constellation (Matthijnsens & Van Ranst, 2012). In addition, the VP7 of the SC2-9 G2-reassortant strain of Rotateq vaccine originates from the tissue culture-adapted parent human RVA strain SC2, dating back to the early 1980s (Matthijnsens *et al.*, 2010). The G2 VP7 of the SC-2 strain belongs to the same lineage as PAI11/1996 (lineage II), but this strain appears to be distantly related to contemporary 2000s G2 strains (Fig. 2), with up to four amino acid differences being observed in the VP7 antigenic epitopes of the most recent Italian strains (Fig. 3). A similar accumulation of mutations in the amino acid sequences of VP7 was observed in Belgian G2 strains post vaccine introduction (Zeller *et al.*, 2012). It is unclear whether the major modifications observed in the neutralizing antigens and in the genome composition of modern G2P[4] RVAs are related, to some extent, to the global introduction of the RVA vaccines. Epidemiological investigations after the introduction of RVA vaccines have reported increased circulation of G2P[4] strains in some countries (Gurgel *et al.*, 2007; Nakagomi *et al.*, 2008; Zeller *et al.*, 2010). However, long-term surveillance studies indicate that large fluctuations in the prevalence of G2P[4] RVAs had also occurred before the introduction of the RVA vaccines (Bányai *et al.*, 2012).

Large-scale surveillance studies and whole genome sequencing are quickly generating a massive amount of data and providing new insights into the mechanisms driving the

evolution of RVAs (Matthijssens *et al.*, 2008a, 2011). It is now clear that human RVAs are subjected to a continual transformation involving accumulation of point mutations, reassortment and repeated intersections with the evolution of animal RVAs (Martella *et al.*, 2011). On the basis of our findings, modern G2P[4] RVAs appear to represent an excellent paradigm of this model.

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