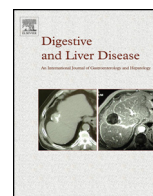




Contents lists available at ScienceDirect

Digestive and Liver Disease

journal homepage: www.elsevier.com/locate/dld



Progress Report

A “systems medicine” approach to the study of non-alcoholic fatty liver disease

Salvatore Petta^a, Luca Valenti^b, Elisabetta Bugianesi^c, Giovanni Targher^d, Stefano Bellentani^{e,f}, Ferruccio Bonino^{g,*}, the Special Interest Group on Personalised Hepatology of the Italian Association for the Study of the Liver (AISF)¹

^a Section of Gastroenterology, Di.Bi.M.I.S Policlinico Paolo Giaccone Hospital, University of Palermo, Italy

^b Internal Medicine, Foundation IRCCS Ca' Granda Ospedale Maggiore Policlinico, Department of Pathophysiology and Transplantation, University of Milan, Italy

^c Gastroenterology and Hepatology, Department of Medical Sciences, Città della Salute e della Scienza di Torino Hospital, University of Turin, Italy

^d Section of Endocrinology, Diabetes and Metabolism, Department of Medicine, University Hospital of Verona, Italy

^e Shrewsbury and Telford NHS Trust, Department of Gastroenterology, Shrewsbury, UK

^f Fondazione Italiana Fegato, Bassovizza, Trieste, Italy

^g General Medicine 2, Department of Clinical and Experimental Medicine, University Hospital of Pisa, Italy

ARTICLE INFO

Article history:

Received 24 May 2015

Accepted 31 October 2015

Available online xxx

Keywords:

Fatty liver

Medicine

NAFLD

NASH

Personalized

Systems medicine

ABSTRACT

The prevalence of fatty liver (steatosis) in the general population is rapidly increasing worldwide. The progress of knowledge in the physiopathology of fatty liver is based on the systems biology approach to studying the complex interactions among different physiological systems. Similarly, translational and clinical research should address the complex interplay between these systems impacting on fatty liver. The clinical needs drive the applications of systems medicine to re-define clinical phenotypes, assessing the multiple nature of disease susceptibility and progression (e.g. the definition of risk, prognosis, diagnosis criteria, and new endpoints of clinical trials). Based on this premise and in light of recent findings, the complex mechanisms involved in the pathology of fatty liver and their impact on the short- and long-term clinical outcomes of cardiovascular, metabolic liver diseases associated with steatosis are presented in this review using a new “systems medicine” approach. A new data set is proposed for studying the impairments of different physiological systems that have an impact on fatty liver in different subsets of subjects and patients.

© 2015 Editrice Gastroenterologica Italiana S.r.l. Published by Elsevier Ltd. All rights reserved.

1. Introduction

The prevalence of non-alcoholic fatty liver disease (NAFLD) in the general population is growing worldwide: 44% in the USA, 33% in Europe and 25% in Italy [1–6]; for non-alcoholic steatohepatitis (NASH), the progressive form of NAFLD, the estimate is 2–3% [3,5]. A large cohort study reported that NAFLD was associated with 26% higher 5-year overall health-care costs, mainly from cardio-metabolic diseases [6].

Progress in the knowledge of NAFLD/NASH pathophysiology was driven by the “systems biology” approach, i.e. the

interdisciplinary study of complex interactions within different biological systems. “Systems pathophysiology” studies the complex interactions between major human vital systems and their interplay. “Systems medicine” combines systems biology and pathophysiological approaches to translational research, integrating various bio-medical tools and using the power of computational and mathematical modelling. This enables the personalization of diagnosis, prognosis and treatment. Systems medicine helps to re-define clinical phenotypes using molecular and dynamic parameters to discover new diagnostic and prognostic biomarkers and to guide the design of new clinical trials. Thus, a systems medicine approach appears mandatory for a “patient-based” classification of the complex interactions between different biological systems and physiological functions involved in NAFLD/NASH, now grouped under the general definition of “metabolic syndrome” (MetS; Table 1). A special interest group (SIG) of the Italian Association for the Study of the Liver (AISF) assembled after the 2013 Single Topic Conference on Personalized Hepatology, held in Pisa

* Corresponding author at: General Medicine 2, Department of Clinical and Experimental Medicine, University of Pisa, Lungarno Bruno Buozzi n. 13, 56125 Pisa, Italy. Tel.: +39 050 543858; fax: +39 050 995457.

E-mail address: ferruccio.bonino@unipi.it (F. Bonino).

¹ See Appendix A.

Table 1
(a) Definition of the metabolic syndrome, according to recent classifications. (b) Quantitative score to estimate the impact of metabolic factors on nonalcoholic fatty liver disease.

(a) Feature	National Cholesterol Education Program, ATP-III	International Diabetes Federation	Joint statement of IDF, NHLBI, AHA, WHF, IAS, IASO
Visceral obesity	>102 cm (males), >88 cm (females)	≥94 cm (males), ≥80 cm (females)(ethnic differences)	≥94 cm (males), ≥80 cm (females)(ethnic differences)
Lipid levels	TG ≥ 150 mg/dL or treated for dyslipidemia HDL-Chol <40 mg/dL (males); <50 mg/dL (females)	TG ≥ 150 mg/dL or treated for dyslipidemia HDL-Chol <40 mg/dL (males); <50 mg/dL (females)	TG ≥ 150 mg/dL or treated for dyslipidemia HDL-Chol <40 mg/dL (males); <50 mg/dL (females)
Arterial pressure	≥130/85 mmHg or treated for Htx	≥130/85 mmHg or treated for Htx	≥130/85 mmHg or treated for Htx
Blood glucose	≥110 mg/dL or treated for DM	≥100 mg/dL or treated for DM	≥100 mg/dL or treated for DM
Notes	3 of the above	Visceral obesity + 2 of the above	3 of the above
Grade			
(b)			
0	No abdominal adiposity and no other features of MetS		
1	Abdominal adiposity		
2	Abdominal adiposity + 1 feature of MetS (i.e. atherogenic dyslipidemia, low HDL cholesterol and/or high TG, hypertension or fasting hyperglycemia/glucose intolerance/diabetes)		
3	Abdominal adiposity + 2 features of MetS		
4	Abdominal adiposity + 3 features of MetS		

Abbreviations: ATP-III, Adult Treatment Panel-III; IDF, International Diabetes Federation; NHLBI, National Heart, Blood and Lung Institute; AHA, American Heart Association; WHF, World Heart Federation; IAS, International Atherosclerosis Society; IASO, International Association for the Study of Obesity; DM, diabetes mellitus; TG, triglycerides; Htx, hypertension; HDL, High Density Lipoprotein; MetS, Metabolic Syndrome.

Notes:

The 10-year risk of having a cardiovascular disease (CVD) can be estimated by the Framingham risk score equation inserting major CVD risk factors (i.e., age, sex, total cholesterol, HDL-cholesterol, smoking history, systolic blood pressure and current use of anti-hypertensive drugs) of the individual patient in the NIH web site: <http://cvdrisk.nhlbi.nih.gov/>.

All patients included in either score 0 or score 1 are in primary prevention of CVD.

Traditional CVD risk factors considered are as follows: family history of premature CVD, age, male sex, cigarette smoking, hypertension (i.e., blood pressure ≥140/90 mmHg or on treatment), LDL-C ≥130 mg/dl (or on treatment), atherogenic dyslipidemia (i.e., triglycerides ≥150 mg/dl and/or HDL-C <50 mg/dl in women <40 mg/dl in men or on treatment), obesity (BMI ≥30 kg/m²) and CKD (estimated glomerular filtration rate <60 ml/min/1.73 m²).

in October 2013. The steering and writing committees of this work included all the experts of the NAFLD/NASH session. Using a rigorous, evidence-based approach, the experts identified and synthesized the literature that forms the evidence regarding specific topics; using their expertise to interpret the evidence, they compiled the specific chapters. The final version of the manuscript was assembled using a step-wise editing process conducted via web-based communication. The steering and writing committees sought additional opinions from an external group of experts who offered their input on an individual basis. The proposed systems medicine approach for the clinico-pathological assessment of the complex abnormalities that have an impact on NAFLD was used as a basis for launching a large prospective cohort study supported by the AISF.

2. Pathophysiology

Obesity and insulin resistance (IR) lead to intrahepatic triglyceride storage, considered the ‘first hit’, that ultimately leads, after a ‘second hit’ [7], to hepatocyte necrosis, inflammation and fibrosis (NASH). More recently, this view was challenged by a ‘multiple hits’ hypothesis, where multiple extra- and intra-hepatic signals are implicated [8–11]. Lipotoxicity is not necessarily linked to triglycerides accumulating within the liver, as some free fatty acids (FFA; palmitate and other lipotoxic intermediates) were shown to be more hepatotoxic than triglycerides [8–11]. Moreover, the liver is targeted by signals from other tissues, including adipose tissue, the gut and its microbiota.

2.1. Lipid partitioning, lipotoxicity and insulin resistance

The increased flux of FFA from an enlarged and insulin-resistant adipose tissue to the liver is considered the major determinant,

with smaller contributions from dietary fat and de novo lipogenesis [12–15]. In the liver, the bulk of absorbed FFA is re-esterified, packaged into very-low density lipoproteins (VLDL) and exported into the bloodstream as triglyceride-rich particles for supplying peripheral tissues. If the ability of insulin to suppress VLDL secretion is impaired, the result is atherogenic dyslipidaemia [13]. Liver steatosis per se does not cause liver necro-inflammation, and could even be seen as a mechanism for preventing lipotoxicity, allowing the storage of FFA in a less harmful form [14,15]. Lipotoxic intermediates and diacylglycerol (DAG) were implicated in hepatic IR, which, in turn, directly contributes to systemic IR and worsens both glucose and lipid metabolism [15].

2.2. Hepatic consequences of deranged metabolism

Different hepatic cell types and intracellular pathways determine the amount of damage and likelihood of progression to advanced fibrosis [16,17]. Kupffer cells play a key role in the pathogenesis of NASH; in mice, depletion of these cells ameliorates steatosis, inflammation, hepatic injury and fibrosis [17]. Kupffer cell activation is mainly dependent on danger signals from steatotic hepatocytes and the accumulation of toxic lipids and bacterial products, and is a critical contributor to the recruitment of monocytes into the liver.

Different signalling pathways regulate intra-hepatic inflammation. Inflammasome was indicated as a pivotal regulator of the interactions with the gut microbiota implicated in the progression of NAFLD and obesity [16–18]. NASH is characterized by hepatocyte apoptosis and apoptotic bodies are pro-inflammatory and pro-fibrogenic [19]; lipotoxic apoptosis is mediated by oxidative stress, considered a ‘second hit’, causing progression to NASH [8,20]. Autophagy is another relevant pathway for NAFLD pathogenesis; its inhibition increases triglyceride storage, but activation favours the progression of fibrosis [21].

2.3. Adipose tissue and NASH

Adipose tissue is a critical site for the development of systemic IR and an altered pattern of adipokine secretion is pivotal for adipose tissue dysfunction [22–25]. Leptin has pro-fibrogenic effects on the liver, via activation of several biological functions of hepatic stellate and Kupffer cells [23]. Adiponectin increases insulin sensitivity and has anti-inflammatory and anti-fibrogenic effects in the liver and in adipose tissue [22,24]. The renin-angiotensin system is another major modulator of IR, critical for liver inflammation and fibrogenesis [25].

2.4. Diet and gut microbiota

The gut microbiota contributes to obesity and fatty liver [26] and the intestinal immune system is critically involved in the regulation of gut microbiota and NALP3 for experimental NASH [18]. The severity of NAFLD histology was associated with higher carbohydrate intake (fructose and sucrose) [27] and accumulating evidence suggests that coffee might be protective on progression of NAFLD and fibrosis [28].

3. Diagnosis

The clinico-epidemiological features of high-risk groups play a major role in guiding the physician to suspect NAFLD and NASH in the single patient (age, sex, ethnicity, body weight and metabolic status) [1–6], whereas serum transaminases do not qualify as markers of NAFLD or NASH [29]. Liver biopsy is a “gold standard” for the diagnosis of NAFLD, even if the number of fat-containing hepatocytes does not correspond exactly to the quantification obtained by magnetic resonance proton spectroscopy (^1H MRS) fat-fraction [30–46]. Its major limitation is the tissue sample size, which corresponds only to 1/50,000th of the liver compared with 1/150th for MRS. Fat accumulation is spatially heterogeneous; a single biopsy may not adequately represent the overall fat content of the liver [31,32]. In addition, liver biopsy is invasive and impractical in monitoring persons at risk of fatty liver disease.

Ultrasound is the most commonly used technique for diagnosis [33,34], based on qualitative features, including echogenicity, echo-texture, beam attenuation, diaphragm and vessel visibility. Fat accentuates scattering; therefore, fatty liver appears hyper-echogenic. Because there is no absolute echogenicity for fat infiltration, a comparison of echogenicity of the kidney is required. These ultrasonographic criteria have a sensitivity range of 60–95% and specificity of 84–100%; in obese patients the sensitivity and specificity are reduced to 49% and 75% respectively [38]. Semi-quantitative ultrasound criteria (mild, moderate or severe steatosis) are affected by subjective interpretation, with poor reproducibility and a low sensitivity for mild steatosis [35]. Recently, several methods were proposed for the quantitative assessment of liver fat content using a combination of computer-assisted measures [39–43]. The computer-aided measurement of the ultrasound hepatic/renal echo intensity ratio and the hepatic echo intensity attenuation rate strongly correlated with liver fat content according to either histology or ^1H MRS [38–43]. Therefore, the combination of ultrasound-based quantitative methods could be used to measure liver fat in clinical practice; however, prospective studies are needed to assess their accuracy and reproducibility.

^1H MRS techniques can measure liver fat, decomposing the liver signal into its fat and water components; the measure is not influenced by fibrosis or obesity and was proven to be highly reliable [38–43]. Its limitations are mainly costs and small spatial coverage with subjective positioning of the volume of interest that may affect accuracy. ^1H MRS is now considered the gold

standard – replacing liver biopsy – as the non-invasive modality for fat liver quantification. Chemical shift imaging (CSI) acquired routinely in liver MRI is also used for liver fat quantification and showed very good correlation with hepatic fat at histology [44].

4. Rationale and methodology of a cohort study

A better understanding of the multiple risk and pathogenetic factors of NAFLD/NASH is mandatory for new diagnostic strategies, and individualized prevention and management. We analysed herein the major pathophysiological systems, factors and co-factors involved in NAFLD/NASH and, using a “systems medicine” approach, we propose new algorithms to stage and/or grade their involvement in the individual.

4.1. Genetics

Heritability plays a major role in the progression of NAFLD towards fibrosing NASH and genome-wide association studies have identified common genetic determinants of steatosis [45–53]. Patatin-like phospholipase-domain-containing-3 (PNPLA3) rs738409 C>G single nucleotide polymorphism, encoding for I148M protein variant is a major determinant [54–59]. The I148M allele frequency explains the inter-ethnic variability of NAFLD: higher in Hispanics (minor allele frequency [MAF] 0.49) than in Europeans (MAF 0.23), and less common in Afro-Americans (MAF 0.17). The PNPLA3-I148M variant hampers triglyceride esterase causing reduced remodelling of lipid droplets in association with excessive intake of carbohydrates or saturated fatty acids and other genetic factors [60–63]. The association between the I148M variant and NAFLD holds true both in adults and in adolescents [64–68]; I148M allele homozygosity predisposes to NASH and hepatic fibrosis [56–58]. The association between the I148M variant and fibrosis is also evident in chronic viral hepatitis and genetic diseases, such as hereditary haemochromatosis [69–71]. In addition, the I148M variant predisposes to hepatocellular carcinoma (HCC), independently of the aetiology of chronic liver disease, and homozygous patients have a worse prognosis [72–76]. The TM6SF2-E167K variant was also associated with NAFLD [53]; it favours progression to NASH and fibrosis by impairing the secretion of very low density lipoproteins in hepatocytes, but protecting from atherosclerosis [59,77]. Therefore, both PNPLA3-I148M and TM6SF2E167K may be useful in identifying NAFLD patients at a higher risk of hepatic than cardiovascular complications and the PNPLA3-based categorisation of NAFLD may have therapeutic implications. Preliminary data suggest that I148M-homozygous subjects might benefit from weight loss after a short-term low-carbohydrate diet [68,78]; however, disease progression is modulated by multiple environmental and genetic factors [48,79–82]. Thus, PNPLA3 I148M and TM6SF2 variants plus/minus a family history for cirrhosis and/or HCC and ethnicity can be used to stratify patients at risk in clinical practice (Table 2).

4.2. Age and gender

4.2.1. Childhood

In children, in whom alcohol abuse, drug consumption and co-morbidities are much less important than in adults, NAFLD is generally considered to be of primary origin [83]. Genetic background, epigenetic programming, intra-uterine environment and post-natal nutrition are major risk factors and single nucleotide polymorphisms (SNPs) may identify children who are at a higher risk of NAFLD [55,56,60,63,65]. In fact, the PNPLA3 rs738409 C>G SNP polymorphism was associated with higher risks of fatty liver, NASH and fibrosis [64,67]. Intrauterine malnutrition causes a foetal

Table 2
Genetic factors score.

	Risk score
Ethnicity	
Afro-American	0
Asiatic or European	0.5
Hispanic	1
Family history for NAFLD/NASH	
Negative	0
Present in one parent	1
In more than one sibling	1.5
In both parents	2
PNPLA3 I148M	
Normal	0
Heterozygous	0.5
Homozygous	1
TM6SF2 E167K	
Absent	0
Present	1

The overall score of genetic factors impact ranges from 0 to 5 resulting from the sum of the values of 3 different categories of genetic factors.

Abbreviations: NAFLD, nonalcoholic fatty liver disease; NASH, nonalcoholic steatohepatitis; PNPLA3 I148M, patatin-like phospholipase-domain gene I148M variant; TM6SF2 E167K, trans-membrane 6 superfamily member 2 gene E167K variant.

adaptive response with a consequent permanent reprogramming of tissue structures and functions [84–86]. NASH and Mets were reported to be higher in children with intrauterine growth retardation, who were overfed after birth, possibly because of discordance between intrauterine and extrauterine environments, whereas breastfeeding seems to protect from NASH [87,88]. These studies suggest that prenatal and postnatal periods are critical for metabolic programming [89,90]. Also, physical inactivity and excessive caloric intake are responsible for the “obese and metabolically dysfunctional” phenotype; diets rich in sugar (soft drinks with fructose-based corn syrup), salt and saturated fats and poor in micronutrients are associated with obesity and NAFLD [91,92].

4.2.2. Gender

The prevalence of NAFLD is higher in men with an “inverted U shaped curve”: it increases from young to middle-aged individuals and declines in the elderly [93]. The “protective” female cardio-metabolic phenotype, present in Caucasians and Asiatics, but not in Hispanics and Blacks, disappears with the menopause [94–98]. Accumulated experimental and clinical evidence suggests that estrogens might exert protective effects on the interplay among brain, endocrine and digestive systems and consequently morbidity and mortality [99–104]. However, the impact of oestrogen replacement in post-menopausal women with NAFLD is still debated [105]. Thus, gender and age-related hormonal changes should be considered for the re-definition of clinical phenotypes, both in a new “patient based” approach and in preventive educational programmes. On this basis, a tentative qualitative score for grading the impact of gender on liver steatosis in relation to age, weight and fat distribution may be applied to stratify individuals with fatty liver disease (Tables 2 and 3).

4.3. Nutrition

The epidemiology of nutrition is a paradox: on the one hand, obesity has doubled in the last 30 years, reaching about one-third of the global population; on the other hand, malnutrition affects about 2 billion people worldwide. Fatty liver occurs in the most severe forms of protein calorie starvation, such as kwashiorkor anorexia, bulimia, cachexia, massive (rapid) weight loss, and uncontrolled inflammatory bowel diseases [106]. However, NASH and fibrosis are highly unusual in these conditions. A typical American diet

Table 3
Hypothesis of a sex-related score for fatty liver disease.

		Females	Males
(a) Effect of age			
<12 years		0	0
>12 <50 years		1	2
>50 years		3	3
Type	Females	Males	Clinical features
(b) Added effect of body weight and fat distribution			
A	0	0	Normal weight, female or male, without either gluteo-femoral or visceral fat accumulation pattern
B	1	3	Female overweight but without gluteo-femoral and or visceral fat accumulation or male overweight without visceral accumulation
C	2	4	Female overweight with gluteo-femoral and or visceral fat accumulation or male with visceral fat accumulation, independently of body weight

(100 g of fat daily) supplies the liver with ~20 g of fat, equivalent to one-half of the total triglyceride (TG) content of an average liver, while the flux of FFA through the bloodstream amounts to ~100 g/day, with 20% being extracted by the liver. Thus, the daily input of TG from the diet (~20 g/day) and FFA from adipose tissue (~20 g/day) approximates the entire TG content of the liver [107]. Under conditions of an acute intake–expenditure imbalance, metabolic tissues store excess nutrients for future use. With a chronic imbalance, the physiological storage capacity is exceeded, activating cellular stress signalling pathways that attempt to stem further nutrient influx by inhibiting insulin signalling and promoting inflammation. In obesity-induced metabolic diseases, the continued nutrient imbalance drives this process forward, leading to chronic inflammation and IR and, ultimately, to type 2 diabetes, cardiovascular disease (CVD) and other overtly pathological consequences. It has been reported that when IR is induced by excess nutrient intake, 59% of hepatic fat is derived from circulating FFA, with lesser contributions from de novo lipogenesis (26%) and diet (15%) [12]. Thus, overfeeding induces subcutaneous and visceral obesity; the latter directs an increased flux of FFA directly to the liver, thereby making a greater contribution to hepatic steatosis. Few studies have evaluated the role of food quantity and quality on NAFLD development and clinical outcomes [108]. The Mediterranean diet is associated with an improvement in health status, as indicated by a significant reduction in overall mortality (9%), mortality from CVD (9%), mortality from cancer (6%) and incidence of Parkinson’s disease and Alzheimer’s disease (13%) [109,110]. Concerning NAFLD and NASH, there are currently a few studies with low numbers of patients. Dietary habits, in particular saturated fat, may promote NASH by modulating hepatic triglyceride accumulation and antioxidant activity, and indirectly by affecting insulin sensitivity and postprandial triglyceride metabolism [111].

The American Heart Association recommends that added sugars should represent less than 5% of total calories (corresponding to 2.5% of calories from fructose), in spite of this the percentage of total energy from added sugars is still approximately 14–15% [112,113]. Substantial relationships have been demonstrated between increased fructose consumption and obesity, dyslipidemia and IR [114]. Compared with glucose, fructose enhances de novo lipogenesis, promotes postprandial hypertriglyceridaemia, induces hepatic and extrahepatic IR, reduces satiety and increases visceral adiposity. The mechanism behind the fructose-induced expansion of ectopic fat is still under intense investigation, but it is hypothesized that fructose might be a strong inducer of de novo lipogenesis, which only produces saturated fatty acids. Fructose consumption may induce hepatic lipid accumulation by activating lipogenic gene expression. Another explanation might be the direct flow of fructose carbon into the glycolytic pathway,

bypassing a key regulatory enzyme of glycolysis, phosphofruktokinase. Through this route, a higher proportion of the carbon from ingested fructose, compared with glucose, is incorporated into triglycerides. Abdelmalek et al. studied the histological severity of NAFLD according to daily fructose intake in a large cohort of adults: although the steatosis grade was lower in those with increased fructose intake, the degree of fibrosis was higher [115]. The daily intake of industrial, not fruit, fructose is a risk factor for severe liver fibrosis, at least in patients with chronic hepatitis C (CHC) [116].

4.3.1. Alcohol intake in NAFLD

There is a great deal of uncertainty regarding the definition of the threshold of alcohol consumption and the duration of abstinence needed to rule out NAFLD [117]. A light-to-moderate alcohol consumption (defined as less than 20 g per day) was associated with a decreased prevalence of NASH and a lesser degree of hepatic fibrosis in a large survey of patients selected from a well-characterized population with biopsy-proven NAFLD, suggesting that light-to-moderate alcohol consumption might have a protective effect on NAFLD progression [118]. Such findings, however, should not encourage physicians to recommend alcohol drinking to teetotalers with NAFLD.

4.4. Metabolic and endocrine systems

4.4.1. Insulin resistance

IR is defined as a decreased sensitivity or responsiveness to the actions of insulin that promote glucose disposal. It plays a major role in type 2 diabetes, and is closely associated with the cluster of metabolic abnormalities that define MetS [119]. IR is a characteristic feature of NAFLD, even in non-obese, non-diabetic subjects [120], and affects both lipid and glucose metabolism in target organs (liver, skeletal muscle, adipose tissue and myocardial muscle). As IR is the hallmark of obesity and several metabolic complications, including NAFLD, quantifying insulin sensitivity/resistance in humans is of great importance. Several direct and indirect methods are currently employed, some of them relying on steady-state analysis of glucose and insulin concentrations, others – such as the gold standard hyperinsulinaemic–euglycaemic clamp – on dynamic testing. Simple surrogates, such as the homeostasis model assessment of IR (HOMA-IR) and the quantitative insulin sensitivity check index (QUICKI) are those most extensively used. Both indices are derived from fasting plasma insulin and glucose concentrations. HOMA-IR is calculated as $\{[\text{fasting insulin (U/ml)}] \times [\text{fasting glucose (mg/dl)}]\} / 22.5$ [121]. HOMA-IR ≥ 3 generally defines a condition of IR, but the coefficient of variation of HOMA-IR varies considerably depending upon the number of samples obtained and the insulin assay used [122]. QUICKI can be calculated from a mathematical transformation of HOMA [123]. Although HOMA-IR and QUICKI are suited for use in large epidemiological or clinical research studies, their specificity in evaluating insulin sensitivity in individual patients is low.

4.4.2. Metabolic syndrome (MetS) and its clinical features

The MetS is a cluster of interrelated metabolic conditions that exponentially increase the risk of developing CVD, type 2 diabetes and NAFLD (Table 1).

The association between NAFLD and the features of MetS is well established. Prevalence of the MetS in NAFLD varies from 18% in normal-weight individuals to 67% in obese subjects [124]. In type 2 diabetes, NAFLD can be diagnosed by ultrasound scan in 69% of cases [125] and 87% of the patients with fatty infiltration who consented to biopsy have histological confirmation of NAFLD [126]. The prevalence of NAFLD in individuals with dyslipidaemia is estimated to be 50% [127].

In this complex interplay, obesity, which has reached epidemic proportions worldwide, is the most common and well-documented risk factor for NAFLD, visceral obesity playing a prominent role in the development of MetS features, including NAFLD [128]. A Chinese study enrolling 5562 subjects who were NAFLD free at baseline, reported that MetS was independently associated with the presence and development of NAFLD during a 5-year follow-up [129]. Along the same lines, a study in the USA reported that “lean NASH” was independently associated with Hispanic ethnicity, younger age and some features of MetS, such as hypertension [130]. Another study enrolling 29,994 adults reported that non-obese patients with NAFLD had a higher prevalence of MetS features than obese patients without NAFLD, especially among women [131]. A large Korean occupational cohort study, involving 2589 subjects who were NAFLD free at baseline, and who were re-examined after a mean of 4.4 years, reported that elevated triglycerides and fasting glucose levels and enlarged waist circumference were independently associated with incident NAFLD [132]. Furthermore, the same metabolic risk factors appear to be associated with its progression to NASH. In 109 NASH patients who underwent a second liver biopsy at least 3 years after the first, progression of liver fibrosis was found in approximately 30% of patients and was associated with increased waist circumference and BMI [133]. Severe liver damage was associated with decreased insulin sensitivity measured by the oral glucose tolerance test-derived oral glucose insulin sensitivity [134] and MetS [135]. Obesity and type 2 diabetes are also implicated in the development of HCC, even in the absence of severe fibrosis [136]. Of note, the relationship between NAFLD and MetS is bidirectional. Liver fat content, as detected by ^1H MRS, is about 4-fold higher in those with than in those without MetS [107]. The risk of CVD mortality and morbidity greatly exceeds the risk of liver-related mortality; thus, NAFLD is not merely the hepatic manifestation of MetS, but rather a systemic pathogenic component (or precursor) of this syndrome [137,138]. Accordingly, we propose a score combining visceral obesity with one or more features of MetS to stratify NAFLD individuals (Tables 2–4).

4.4.3. Thyroid dysfunction

A consistent line of research has associated thyroid dysfunction with NAFLD [139–141]. A German study reported that low free thyroxine levels were associated with ultrasound-detected steatosis in 3661 participants with no history of thyroid or liver diseases [142]. Another study of 4648 health-check subjects (2324 subjects with hypothyroidism vs age- and sex-matched controls) reported that subclinical hypothyroidism was related to NAFLD in a dose-dependent manner [143].

4.5. Cardiovascular system

In the last decade, it became evident that the clinical burden of NAFLD is not restricted to liver-related morbidity or mortality, and that the majority of deaths among these patients are related to malignancy, coronary heart disease (CHD) and other cardiovascular complications [6]. The spectrum of cardiovascular complications associated with NAFLD spans from premature atherosclerosis to aortic valve sclerosis and left ventricular dysfunction/hypertrophy leading to congestive heart failure and cardiac arrhythmias (mainly atrial fibrillation) [138,144–147]. Growing evidence indicates that NAFLD may play a role in the development and progression of cardiovascular complications, not only through MetS, but also through multiple pathophysiological derangements, including chronic inflammation, hypercoagulation, chronic kidney disease, hyperuricaemia, hypovitaminosis D, hypoadiponectinaemia, and increased fetuin-A levels [138,146,147]. NAFLD exacerbates IR, causes atherogenic dyslipidaemia, and releases pro-inflammatory,

Table 4
Cardiovascular risk.

Score	Degree of impairment of cardiovascular system	Patients	CVD risk factors and/or Framingham risk score ^a
0	Minimal	Asymptomatic ^b	Either <3 traditional CVD risk factors ^c or a low/intermediate Framingham risk score (i.e. a 10-year CHD risk <15%)
1	Intermediate	Asymptomatic ^b	Either a high Framingham risk score (i.e. a 10-year CHD risk >15%) or ≥3 traditional CVD risk factors ^c
2	Severe	In secondary prevention of CVD or at very high CVD risk (e.g., angina, myocardial infarction, coronary revascularization), stroke or other clinical CVD complications such as peripheral artery disease, abdominal aortic aneurysm or carotid artery stenosis >60% or established diabetes	

^a The 10-year risk of having a CVD can be estimated by the Framingham risk score equation inserting major CVD risk factors (i.e., age, sex, total cholesterol, HDL-cholesterol, smoking history, systolic blood pressure and current use of anti-hypertensive drugs) of the individual patient in the NIH web site: <http://cvdrisk.nhlbi.nih.gov/>.

^b All patients included in either score 0 or score 1 are in primary prevention of CVD.

^c Traditional CVD risk factors considered are as follows: family history of premature CVD, age, male sex, cigarette smoking, hypertension (i.e., blood pressure ≥140/90 mmHg or on treatment), LDL-C ≥130 mg/dl (or on treatment), atherogenic dyslipidemia (i.e., triglycerides ≥150 mg/dl and/or HDL-C <50 mg/dl in women <40 mg/dl in men or on treatment), obesity (BMI ≥30 kg/m²) and CKD (estimated glomerular filtration rate <60 ml/min/1.73 m²).

CVD, cardiovascular disease; LDL-C, low density lipoprotein cholesterol; HDL-C, high density lipoprotein cholesterol; CKD, chronic kidney disease.

pro-coagulant, pro-oxidant and pro-fibrogenic mediators of cardiovascular pathophysiology [138,144–147].

Based on this evidence, scientific societies have suggested an assessment of the overall CVD risk in patients with NAFLD [1,148]; however, how such an assessment should be conducted is still poorly defined [146] and in general it follows the guidelines for the CVD risk of the general adult population [146,117,149]. Preliminary evidence suggests that the Framingham risk score might accurately predict the higher 10-year CHD risk in patients in the USA with NAFLD and identify patients expected to benefit from early interventions to prevent CHD events [149,150]. However, the accuracy of the Framingham risk score needs to be further validated in European patients with NAFLD, where this equation may overestimate the CHD risk compared with populations of Anglo-Saxon ancestry. Furthermore, future studies in larger cohorts of NAFLD patients with different ethnicities are needed to validate the Framingham or other risk score systems for predicting the global CVD risk, given that subclinical inflammation, IR and hypertriglyceridaemia are not considered in any of the available risk score systems. In Table 4 we propose a grading scale for global CVD risk in NAFLD based on available evidence and guidelines.

4.6. Immune disorders, chronic viral infections, and gallstone disease

4.6.1. Immune disorders

Both innate and adaptive immune response play a pivotal role in the pathogenesis and progression of NAFLD [9,151,152]. Experimental observations match clinical evidence on the association between immune disorders and the presence/severity of NAFLD [153,154]. Circulating autoantibodies are frequent in NAFLD in the absence of autoimmune hepatitis and associated with more advanced liver disease at histology [155]. Serum IgA levels are significantly associated with NASH and more advanced liver fibrosis in NAFLD [153,154]. An intriguing link between immune disorders and NAFLD is suggested by the prevalence of fatty liver in rheumatological disorders [156], primary biliary cirrhosis [157–159], coeliac and inflammatory bowel diseases (IBD) [160–163]. NAFLD is the third most common IBD-associated liver disease independent of classical risk factors such as obesity, IR or drug toxicity [161,162].

4.6.2. Chronic viral infections

Fatty liver is common in patients with CHC, with prevalence ranging from 40 to 80%, higher than in chronic liver diseases of different aetiology [164,165]. The pathogenesis of steatosis in CHC is multifactorial, involving both host and viral factors. A direct steatogenic effect was shown for hepatitis C virus (HCV) genotype 3 [166–170], where steatosis is more frequent and severe according to viral load [167,168] and disappears after viral eradication

Table 5

Grading of the impact of chronic infections and/or immune disorders on fatty liver.

Grade	Infections and/or immune disorders
0	Absence of clinical and pathological signs of immune disorders, circulating autoantibodies and chronic HBV or HCV infections
1	One of the following features: clinic-pathological signs of immune disorders or circulating auto-antibodies and chronic HBV or HCV infection without chronic active hepatitis (inactive HBsAg carrier or anti-HCV positive but HCV-RNA negative)
2	Both clinical or pathological signs of immune disorders or circulating auto-antibodies and chronic HBV or HCV infection without chronic hepatitis (inactive HBsAg carrier or anti-HCV positive but HCV-RNA negative)
3	Chronic hepatitis B
4	Chronic hepatitis C

HBV, hepatitis B virus; HCV, hepatitis C virus; HBsAg, hepatitis B surface antigen.

[171]. Conversely, a combined viral and metabolic steatosis occurs in non-3 HCV genotype infections and correlates with age and IR [170]. Nevertheless, HCV genotype 1 can induce IR [171–175]. Interesting findings support the interplay between the host's genetics and fatty liver in HCV-infected individuals: the PNPLA3 I148M variant [176] is associated with steatosis, whereas the IL28B rs868 CC genotype is associated with a sustained response to peginterferon and ribavirin and lower prevalence of IR and steatosis [177]. Furthermore, steatosis has a relevant impact on the clinical history of HCV infection (liver disease progression and HCC) [170,178–185]. The association between chronic hepatitis B virus (HBV) infection and steatosis is less evident than in CHC; the prevalence of fatty liver is extremely variable (5–70%) [186–192]. Nevertheless, steatosis is more frequent in chronic hepatitis B (CHB) patients than in the general population in Western and Asian countries without the impact of ethnicity or viral genotypes [190–192]. In most studies of CHB patients steatosis was not associated with fibrosis; this raises the question whether NAFLD in HBsAg carriers might be linked to metabolic factors indirectly influenced by HBV. As in chronic HCV infection, the pathogenesis of steatosis is related to metabolic factors such as obesity and IR [187–189] and genetic background (PNPLA3 I148M variant) [70], but at variance with HCV a direct steatogenic impact of HBV has been reported in a few experimental studies [191–195]. Whatever the underlying mechanisms, steatosis in both CHB and CHC represents an important co-factor affecting the outcome of liver disease [192]. Thus, we proposed a scale for grading the combined impact of immune disorders and viral infections in NAFLD patients (Table 5).

4.6.3. Gallstones or cholecystectomy

Studies of the association between NAFLD and gallstones provided conflicting results. In a large study NAFLD was associated with cholecystectomy, but not with gallstones, suggesting that

cholecystectomy might be a potential risk factor [196]. Another study found a high prevalence of gallstones in NAFLD patients with glucose intolerance/diabetes and advanced liver disease [197].

5. Future perspectives

With this approach, it is possible to characterize different NAFLD/NASH phenotypes within the definition of the general metabolic syndrome. Based on this premise we aim to launch a large observational cohort study supported by AIFS and the Foundation for Research in Hepatology (FIRE). The goal is to study the clinical comorbidities and hepatic and extrahepatic outcomes associated with subsets of patients with NAFLD/NASH, to increase the understanding of the complex interplay among different physiological systems. A prospective cohort of consecutive asymptomatic individuals with fatty liver will be recruited in clinical centres distributed throughout Italy and followed up for at least five years. Novel statistical models such as non-linear and integrative epidemiological approaches [198], accounting for the patterns of interconnections between parameters affecting disease risk and their correlations, will be used to better understand the mechanisms involved. With this integrated approach we hope to identify new diagnostic and prognostic biomarkers and new targets for prevention and treatment.

Conflict of interest

None declared.

Appendix A. Collaborators

The following investigators contributed equally to the study as members of the writing committee:

Ele Ferrannini, National Research Council (CNR), Institute of Clinical Physiology, Pisa, Italy
Carmela Loguercio, Hepato-Gastroenterology, 2nd University of Naples, Italy
Amedeo Lonardo, Outpatient Liver Clinic and Internal Medicine NOCSAE, USL Modena, Italy
Fabio Marra, Department of Experimental and Clinical Medicine, University of Florence, Italy
Marcello Mancini, National Research Council (CNR), Institute of Biostructure and Bio-imaging, Naples, Italy
Luca Miele, Internal Medicine, Gastroenterology and Liver Diseases, Catholic University of Rome, Italy
Valerio Nobili, Metabolic Liver Diseases Research Unit, IRCCS Children Hospital “Bambino Gesù”, Rome, Italy
Gianluca Svegliati Baroni, Department of Gastroenterology 1 and Obesity Center 2, Polytechnic University of Marche, Ancona, Italy

In addition to the above listed authors of the writing committee the following investigators contributed equally to the study as external advisors:

Federico Alessandro, Hepato-Gastroenterology, 2nd University of Naples, Italy
Stefano Ballestri, Medicine, Pavullo Hospital, Modena, Italy
Maurizia Rossana Brunetto and Barbara Coco, Hepatology Unit, University Hospital of Pisa, Italy
Antonio Grieco, Internal Medicine, Gastroenterology and Liver Diseases, Catholic University of Rome, Italy
Silvia Fargion, Internal Medicine, Foundation IRCCS Ca' Granda Ospedale Maggiore Policlinico, Department of Pathophysiology and Transplantation, University of Milan, Italy
Loreta Kondili, Istituto Superiore di Sanità, Rome, Italy
Fabio Nascimbeni, Metabolic Medicine, NOCSAE, Baggiovara, Modena, Italy

Anna Prinster, National Research Council (CNR), Institute of Biostructure and Bioimaging, Naples, Italy
Dante Romagnoli, Outpatient Liver Clinic, Medicine NOCSAE, Baggiovara, Modena, Italy
Stefano Taddei, General Medicine 1, Department of Clinical and Experimental Medicine, University Hospital of Pisa, Italy
Ester Vanni, Gastroenterology and Hepatology, Department of Medical Sciences, Città della Salute e della Scienza di Torino Hospital, University of Turin, Italy
Stefano Vella, Istituto Superiore di Sanità, Rome, Italy

References

- [1] Loria P, Adinolfi LE, Bellentani S, et al. NAFLD Expert Committee of the Associazione Italiana per lo studio del Fegato. Practice guidelines for the diagnosis and management of nonalcoholic fatty liver disease. A decalogue from the Italian Association for the Study of the Liver (AISF) Expert Committee. *Digestive and Liver Disease* 2010;42:272–82.
- [2] Bellentani S, Scaglioni F, Marino M, et al. Epidemiology of non-alcoholic fatty liver disease. *Digestive Diseases* 2010;28:155–61.
- [3] Bedogni G, Miglioli L, Masutti F, et al. Incidence and natural course of fatty liver in the general population: the Dionysos study. *Hepatology* 2007;46:1387–91.
- [4] Lazo M, Hernaez R, Eberhardt MS, et al. Prevalence of nonalcoholic fatty liver disease in the United States: the Third National Health and Nutrition Examination Survey, 1988–1994. *American Journal of Epidemiology* 2013;178:38–45.
- [5] Li Z, Xue J, Chen P, et al. Prevalence of nonalcoholic fatty liver disease in mainland of China: a meta-analysis of published studies. *Journal of Gastroenterology and Hepatology* 2014;29:42–51.
- [6] Baumeister SE, Völzke H, Marschall P, et al. Impact of fatty liver disease on health care utilization and costs in a general population: a 5-year observation. *Gastroenterology* 2008;134:85–94.
- [7] Day CP, James OFW. Steatohepatitis: a tale of two hits. *Gastroenterology* 1998;114:842–5.
- [8] Neuschwander-Tetri BA. Hepatic lipotoxicity and the pathogenesis of non-alcoholic steatohepatitis: the central role of nontriglyceride fatty acid metabolites. *Hepatology* 2010;52:774–88.
- [9] Marra F, Lotersztajn S. Pathophysiology of NASH: perspectives for a targeted treatment. *Current Pharmaceutical Design* 2013;19:5250–69.
- [10] Ricchi M, Odoardi MR, Carulli L, et al. Differential effect of oleic and palmitic acid on lipid accumulation and apoptosis in cultured hepatocytes. *Journal of Gastroenterology and Hepatology* 2009;24:830–40.
- [11] Tilg H, Moschen AR. Evolution of inflammation in nonalcoholic fatty liver disease: the multiple parallel hits hypothesis. *Hepatology* 2010;52:1836–46.
- [12] Donnelly KL, Smith CI, Schwarzenberg SJ, et al. Sources of fatty acids stored in liver and secreted via lipoproteins in patients with nonalcoholic fatty liver disease. *Journal of Clinical Investigation* 2005;115:1343–51.
- [13] Fabbri E, Mohammed BS, Magkos F, et al. Alterations in adipose tissue and hepatic lipid kinetics in obese men and women with nonalcoholic fatty liver disease. *Gastroenterology* 2008;134:424–31.
- [14] Yamaguchi K, Yang L, McCall S, et al. Inhibiting triglyceride synthesis improves hepatic steatosis but exacerbates liver damage and fibrosis in obese mice with nonalcoholic steatohepatitis. *Hepatology* 2007;45:1366–74.
- [15] Cusi K. Role of obesity and lipotoxicity in the development of nonalcoholic steatohepatitis: pathophysiology and clinical implications. *Gastroenterology* 2012;142:711–25.
- [16] Henaoui-Mejia J, Elinav E, Jin C, et al. Inflammasome-mediated dysfunction regulates progression of NAFLD and obesity. *Nature* 2012;482:179–85.
- [17] Csak T, Ganz M, Pespisa J, et al. Fatty acid and endotoxin activate inflammasomes in mouse hepatocytes that release danger signals to stimulate immune cells. *Hepatology* 2011;54:133–44.
- [18] Vandanmagsar B, Youm YH, Ravussin A, et al. The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nature Medicine* 2011;17:179–88.
- [19] Guicciardi ME, Gores GJ. Apoptosis as a mechanism for liver disease progression. *Seminars in Liver Disease* 2010;30:402–10.
- [20] Gambino R, Musso G, Cassader M. Redox balance in the pathogenesis of nonalcoholic fatty liver disease: mechanisms and therapeutic opportunities. *Antioxidants and Redox Signaling* 2011;15:1325–65.
- [21] Hernandez-Gea V, Ghiassi-Nejad Z, Rozenfeld R, et al. Autophagy releases lipid that promotes fibrogenesis by activated hepatic stellate cells in mice and in human tissues. *Gastroenterology* 2012;142:938–46.
- [22] Marra F, Bertolani C. Adipokines in liver diseases. *Hepatology* 2009;50:957–69.
- [23] Wang J, Leclercq I, Brymora JM, et al. Kupffer cells mediate leptin-induced liver fibrosis. *Gastroenterology* 2009;137:713–23.
- [24] Tilg H, Moschen AR. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. *Nature Reviews Immunology* 2006;6:772–83.
- [25] Matthew Morris E, Fletcher JA, Thyfault JP, et al. The role of angiotensin II in nonalcoholic steatohepatitis. *Molecular and Cellular Endocrinology* 2013;378:29–40.
- [26] Machado MV, Cortez-Pinto H. Gut microbiota and nonalcoholic fatty liver disease. *Annals of Hepatology* 2012;11:440–9.

- [27] Yilmaz Y. Review article: fructose in non-alcoholic fatty liver disease. *Alimentary Pharmacology and Therapeutics* 2012;35:1135–44.
- [28] Molloy JW, Calcagno CJ, Williams CD, et al. Association of coffee and caffeine consumption with fatty liver disease, nonalcoholic steatohepatitis, and degree of hepatic fibrosis. *Hepatology* 2012;55:429–36.
- [29] Mofrad P, Contos MJ, Haque M, et al. Clinical and histologic spectrum of non-alcoholic fatty liver disease associated with normal ALT values. *Hepatology* 2003;37:1286–92.
- [30] Brunt EM, Janney CG, Di Bisceglie AM, et al. Nonalcoholic steatohepatitis: a proposal for grading and staging the histological lesions. *American Journal of Gastroenterology* 1999;94:2467–74.
- [31] Ratzliff V, Charlotte F, Heurtier A, et al. Sampling variability of liver biopsy in nonalcoholic fatty liver disease. *Gastroenterology* 2005;128:1898–906.
- [32] Arun J, Jhala N, Lazenby AJ, et al. Influence of liver biopsy heterogeneity and diagnosis of nonalcoholic steatohepatitis in subjects undergoing gastric bypass. *Obesity Surgery* 2007;17:155–61.
- [33] Charatcharoenwithaya P, Lindor KD. Role of radiologic modalities in the management of nonalcoholic steatohepatitis. *Clinics in Liver Disease* 2007;11:37–54.
- [34] Mishra P, Younossi ZM. Abdominal ultrasound for diagnosis of nonalcoholic fatty liver disease (NAFLD). *American Journal of Gastroenterology* 2007;102:2716–7.
- [35] Strauss S, Gavish E, Gottlieb P, et al. Interobserver and intraobserver variability in the sonographic assessment of fatty liver. *American Journal of Roentgenology* 2007;189:W320–3.
- [36] Dasarathy S, Dasarathy J, Khyami A, et al. Validity of real time ultrasound in the diagnosis of hepatic steatosis: a prospective study. *Journal of Hepatology* 2009;51:1061–7.
- [37] Edens MA, van Ooijen PM, Post WJ, et al. Ultrasonography to quantify hepatic fat content: validation by ¹H magnetic resonance spectroscopy. *Obesity (Silver Spring)* 2009;17:2239–44.
- [38] Mancini M, Prinster A, Annuzzi G, et al. Sonographic hepatic-renal ratio as indicator of hepatic steatosis: comparison with (1)H magnetic resonance spectroscopy. *Metabolism: Clinical and Experimental* 2009;58:1724–30.
- [39] Fischbach F, Bruhn H. Assessment of in vivo ¹H magnetic resonance spectroscopy in the liver: a review. *Liver International* 2008;28:297–307.
- [40] Reeder SB, Cruite I, Hamilton G, et al. Quantitative assessment of liver fat with magnetic resonance imaging and spectroscopy. *Journal of Magnetic Resonance Imaging* 2011;34:729–49.
- [41] Schwenzler NF, Springer F, Schraml C, et al. Non-invasive assessment and quantification of liver steatosis by ultrasound, computed tomography and magnetic resonance. *Journal of Hepatology* 2009;51:433–45.
- [42] Fishbein M, Castro F, Cheruku S, et al. Hepatic MRI for fat quantification. Its relationship to fat morphology, diagnosis and ultrasound. *Journal of Clinical Gastroenterology* 2005;39:619–25.
- [43] Noworolski SM, Lam MM, Merriman RB, et al. Liver steatosis: concordance of MR imaging and MR spectroscopic data with histologic grade. *Radiology* 2012;264:88–96.
- [44] Koelblinger C, Krassak M, Maresch J, et al. Hepatic steatosis assessment with ¹H-spectroscopy and chemical shift imaging at 3.0 T before hepatic surgery: reliable enough for making clinical decisions? *European Journal of Radiology* 2012;81:2990–5.
- [45] Schwimmer JB, Celedon MA, Lavine JE, et al. Heritability of nonalcoholic fatty liver disease. *Gastroenterology* 2009;136:1585–92.
- [46] Guerrero R, Vega GL, Grundy SM, et al. Ethnic differences in hepatic steatosis: an insulin resistance paradox. *Hepatology* 2009;49:791–801.
- [47] Makkonen J, Pietilainen KH, Rissanen A, et al. Genetic factors contribute to variation in serum alanine aminotransferase activity independent of obesity and alcohol: a study in monozygotic and dizygotic twins. *Journal of Hepatology* 2009;50:1035–42.
- [48] Dongiovanni P, Anstee QM, Valenti L. Genetic predisposition in NAFLD and NASH: impact on severity of liver disease and response to treatment. *Current Pharmaceutical Design* 2013;19:5219–38.
- [49] Speliotes EK, Yerges-Armstrong LM, Wu J, et al. Genome-wide association analysis identifies variants associated with nonalcoholic fatty liver disease that have distinct effects on metabolic traits. *PLoS Genetics* 2011;7:e1001324.
- [50] Anstee QM, Darlay R, Leathart J, et al. A candidate gene approach to validation of genetic modifier associations using a large cohort with histologically characterized non-alcoholic fatty liver disease. *Journal of Hepatology* 2013;58:S46.
- [51] Romeo S, Kozlitina J, Xing C, et al. Genetic variation in *pnpla3* confers susceptibility to nonalcoholic fatty liver disease. *Nature Genetics* 2008;40:1461–5.
- [52] Kitamoto T, Kitamoto A, Yoneda M, et al. Genome-wide scan revealed that polymorphisms in the *pnpla3*, *samm50*, and *parvb* genes are associated with development and progression of nonalcoholic fatty liver disease in Japan. *Human Genetics* 2013;132:783–92.
- [53] Kozlitina J, Smagris E, Stender S, et al. Exome-wide association study identifies a TM6SF2 variant that confers susceptibility to nonalcoholic fatty liver disease. *Nature Genetics* 2014;46:352–6.
- [54] Tian C, Stokowski RP, Kershenovich D, et al. Variant in *pnpla3* is associated with alcoholic liver disease. *Nature Genetics* 2009;42:21–3.
- [55] Sookoian S, Castano GO, Burgueno AL, et al. A nonsynonymous gene variant in the *adiponutrin* gene is associated with nonalcoholic fatty liver disease severity. *Journal of Lipid Research* 2009;50:2111–6.
- [56] Valenti L, Al-Serri A, Daly AK, et al. Homozygosity for the *pnpla3/adiponutrin* i148m polymorphism influences liver fibrosis in patients with nonalcoholic fatty liver disease. *Hepatology* 2010;51:1209–17.
- [57] Sookoian S, Pirola CJ. Meta-analysis of the influence of i148m variant of patatin-like phospholipase domain containing 3 gene (*pnpla3*) on the susceptibility and histological severity of nonalcoholic fatty liver disease. *Hepatology* 2011;53:1883–94.
- [58] Dongiovanni P, Donati B, Fares R, et al. PNPLA3 I148M polymorphism and progressive liver disease. *World Journal of Gastroenterology* 2013;19:6969–78.
- [59] Dongiovanni P, Petta S, Maglio C, et al. Transmembrane 6 superfamily member 2 gene variant disentangles nonalcoholic steatohepatitis from cardiovascular disease. *Hepatology* 2015;61:506–14.
- [60] He S, McPhaul C, Li JZ, et al. A sequence variation (I148M) in PNPLA3 associated with nonalcoholic fatty liver disease disrupts triglyceride hydrolysis. *Journal of Biological Chemistry* 2010;285:6706–15.
- [61] Davis JN, Le KA, Walker RW, et al. Increased hepatic fat in overweight hispanic youth influenced by interaction between genetic variation in *pnpla3* and high dietary carbohydrate and sugar consumption. *American Journal of Clinical Nutrition* 2011;92:1522–7.
- [62] Santoro N, Savoye M, Kim G, et al. Hepatic fat accumulation is modulated by the interaction between the rs738409 variant in the *pnpla3* gene and the dietary omega6/omega3 pufa intake. *PLoS ONE* 2012;7:e37827.
- [63] Ruhanen H, Perttälä J, Hölttä-Vuori M, et al. PNPLA3 mediates hepatocyte triacylglycerol remodeling. *Journal of Lipid Research* 2014;55:739–46.
- [64] Valenti L, Alisi A, Galmozzi E, et al. I148M patatin-like phospholipase domain containing 3 gene variant and severity of pediatric nonalcoholic fatty liver disease. *Hepatology* 2010;52:1274–80.
- [65] Santoro N, Zhang CK, Zhao H, et al. Variant in the glucokinase regulatory protein (*gckr*) gene is associated with fatty liver in obese children and adolescents. *Hepatology* 2011;55:781–9.
- [66] Miraglia Del Giudice E, Grandone A, Cirillo G, et al. The association of *pnpla3* variants with liver enzymes in childhood obesity is driven by the interaction with abdominal fat. *PLoS ONE* 2011;6:e27933.
- [67] Valenti L, Motta BM, Alisi A, et al. LPIN1 rs13412852 polymorphism in pediatric non-alcoholic fatty liver disease. *Journal of Pediatric Gastroenterology and Nutrition* 2012;54:588–93.
- [68] Nobili V, Bedogni G, Donati B, et al. The i148m variant of *pnpla3* reduces the response to docosahexaenoic acid in children with non-alcoholic fatty liver disease. *Journal of Medicinal Food* 2013;16:957–60.
- [69] Valenti L, Rumi M, Galmozzi E, et al. Patatin-like phospholipase domain-containing 3 i148m polymorphism, steatosis, and liver damage in chronic hepatitis C. *Hepatology* 2011;53:791–9.
- [70] Vigano M, Valenti L, Lampertico P, et al. *Pnpla3* i148m affects liver steatosis in patients with chronic hepatitis B. *Hepatology* 2013;58:1245–52.
- [71] Valenti L, Maggioni P, Piperno A, et al. *Pnpla3* i148m polymorphism, steatosis, and liver damage in hereditary hemochromatosis. *World Journal of Gastroenterology* 2012;18:2813–20.
- [72] Trepo E, Nahon P, Bontempi G, et al. Association between the *pnpla3* (rs738409 c>g) variant and hepatocellular carcinoma: evidence from a meta-analysis of individual participant data. *Hepatology* 2014;59:2170–7.
- [73] Valenti L, Motta BM, Soardo G, et al. *Pnpla3* i148m polymorphism, clinical presentation, and survival in patients with hepatocellular carcinoma. *PLoS ONE* 2013;8:e75982.
- [74] Liu YL, Patman G, Leathart J, et al. Carriage of *pnpla3* i148m is associated with increased risk of nonalcoholic fatty liver disease-associated hepatocellular carcinoma. *Journal of Hepatology* 2013;58:S516.
- [75] Hassan MM, Kaseb A, Etzel CJ, et al. Genetic variation in the *pnpla3* gene and hepatocellular carcinoma in USA: risk and prognosis prediction. *Molecular Carcinogenesis* 2013;52:E139–47.
- [76] Valenti L, Dongiovanni P, Ginanni Corradini S, et al. *Pnpla3* i148m variant and hepatocellular carcinoma: a common genetic variant for a rare disease. *Digestive and Liver Disease* 2013;45:619–24.
- [77] Liu YL, Reeves HL, Burt AD, et al. TM6SF2 rs58542926 influences hepatic fibrosis progression in patients with non-alcoholic fatty liver disease. *Nature Communications* 2014;5:4309.
- [78] Sevastianova K, Santos A, Kotronen A, et al. Effect of short-term carbohydrate overfeeding and long-term weight loss on liver fat in overweight humans. *American Journal of Clinical Nutrition* 2010;96:727–34.
- [79] Petta S, Miele L, Bugianesi E, et al. Glucokinase regulatory protein gene polymorphism affects liver fibrosis in non-alcoholic fatty liver disease. *PLoS ONE* 2014;9:e87523.
- [80] Fares R, Petta S, Lombardi R, et al. The UCP2-866 G>A promoter region polymorphism is associated with nonalcoholic steatohepatitis. *Liver International* 2014. <http://dx.doi.org/10.1111/liv.12707>.
- [81] Valenti L, Canavesi E, Galmozzi E, et al. Beta-globin mutations are associated with parenchymal siderosis and fibrosis in patients with non-alcoholic fatty liver disease. *Journal of Hepatology* 2010;53:927–33.
- [82] Cefalu AB, Pirruccello JP, Noto D, et al. A novel *apob* mutation identified by exome sequencing cosegregates with steatosis, liver cancer, and hypocholesterolemia. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2013.
- [83] Nobili V, Svegliati-Baroni G, Alisi A, et al. A 360-degree overview of paediatric NAFLD: recent insights. *Journal of Hepatology* 2013;58:1218–29.
- [84] Fowden AL, Giussani DA, Forhead AJ. Intrauterine programming of physiological systems: causes and consequences. *Physiology (Bethesda)* 2006.

- [85] Nobili V, Marcellini M, Marchesini G, et al. Intrauterine growth retardation, insulin resistance, and nonalcoholic fatty liver disease in children. *Diabetes Care* 2007;30:2638–40.
- [86] Alisi A, Panera N, Agostoni C, et al. Intrauterine growth retardation and nonalcoholic fatty liver disease in children. *International Journal of Endocrinology* 2011;2011:269853.
- [87] Nobili V, Bedogni G, Alisi A, et al. A protective effect of breastfeeding on the progression of nonalcoholic fatty liver disease. *Archives of Disease Childhood* 2009;94:801–5.
- [88] Alisi A, Cianfarani S, Manco M, et al. Nonalcoholic fatty liver disease and metabolic syndrome in adolescents: pathogenetic role of genetic background and intrauterine environment. *Annals of Medicine* 2012;44:29–40.
- [89] Papandreou D, Karabouta Z, Rousso I. Are dietary cholesterol intake and serum cholesterol levels related to nonalcoholic fatty liver disease in obese children? *Cholesterol* 2012;2012:572820.
- [90] Yasutake K, Nakamura M, Shima Y, et al. Nutritional investigation of nonobese patients with nonalcoholic fatty liver disease: the significance of dietary cholesterol. *Scandinavian Journal Gastroenterology* 2009;44:471–7.
- [91] Jin R, Le NA, Liu S, et al. Children with NAFLD are more sensitive to the adverse metabolic effects of fructose beverages than children without NAFLD. *Journal of Clinical Endocrinology and Metabolism* 2012;97:E1088–98.
- [92] Nomura K, Yamanouchi T. The role of fructose-enriched diets in mechanism of nonalcoholic fatty liver disease. *Journal of Nutritional Biochemistry* 2012;23:203–8.
- [93] Bertolotti M, Lonardo A, Mussi C, et al. Nonalcoholic fatty liver disease and aging: epidemiology to management. *World Journal of Gastroenterology* 2014;20:14185–204.
- [94] Lovejoy JC, Champagne CM, de Jonge L, et al. Increased visceral fat and decreased energy expenditure during the menopausal transition. *International Journal of Obesity* 2008;32:949–58.
- [95] Volzke H, Schwarz S, Baumeister SE, et al. Menopausal status and hepatic steatosis in a general female population. *Gut* 2007;56:594–5.
- [96] Park SH, Jeon WK, Kim SH, et al. Prevalence and risk factors of non-alcoholic fatty liver disease among Korean adults. *Journal of Gastroenterology and Hepatology* 2006;21:138–43.
- [97] Hamaguchi M, Kojima T, Ohhara A, et al. Aging is a risk factor of nonalcoholic fatty liver disease in premenopausal women. *World Journal of Gastroenterology* 2012;18:237–43.
- [98] Yang JD, Abdelmalek MF, Pang H, et al. Gender and menopause impact severity of fibrosis among patients with nonalcoholic steatohepatitis. *Hepatology* 2014;59:1406–14.
- [99] Erkan G, Yilmaz G, Konca Degertekin C, et al. Presence and extent of estrogen receptor- α expression in patients with simple steatosis and NASH. *Pathology, Research and Practice* 2013;209:429–32.
- [100] Itagaki T, Shimizu I, Cheng X, et al. Opposing effects of oestradiol and progesterone on intracellular pathways and activation processes in the oxidative stress induced activation of cultured rat hepatic stellate cells. *Gut* 2005;54:1782–9.
- [101] Yasuda M, Shimizu I, Shiba M, et al. Suppressing effects of estradiol on dimethylnitrosamine-induced fibrosis of the liver in rats. *Hepatology* 1999;29:719–27.
- [102] Kireev RA, Tresguerres AC, Garcia C, et al. Hormonal regulation of pro-inflammatory and lipid peroxidation processes in liver of old ovariectomized female rats. *Biogerontology* 2010;11:229–43.
- [103] Rogers A, Eastell R. The effect of 17 β -estradiol on production of cytokines in cultures of peripheral blood. *Bone* 2001;29:30–4.
- [104] Rettberg JR, Yao J, Brinton RD. Estrogen: a master regulator of bioenergetic systems in the brain and body. *Frontiers in Neuroendocrinology* 2014;35:8–30.
- [105] McKenzie J, Fisher BM, Jaap AJ, et al. Effects of HRT on liver enzyme levels in women with type 2 diabetes: a randomized placebo-controlled trial. *Clinical Endocrinology* 2006;65:40–4.
- [106] Kneeman JM, JMisdraji J, Corey KE. Secondary causes of nonalcoholic fatty liver disease. *Therapeutic Advances in Gastroenterology* 2012;5:199–207.
- [107] Cohen JC, Horton JD, Hobbs HH. Human fatty liver disease: old questions and new insights. *Science* 2011;332:1519–23.
- [108] de Wit NJ, Afman LA, Mensink M, et al. Phenotyping the effect of diet on non-alcoholic fatty liver disease. *Journal of Hepatology* 2012;57:1370–3.
- [109] Sofi F, Cesari F, Abbate R, et al. Adherence to Mediterranean diet and health status: meta-analysis. *BMJ* 2008;337:a1344.
- [110] Sofi F, Macchi C, Abbate R, et al. Mediterranean diet and health status: an updated meta-analysis and a proposal for a literature-based adherence score. *Public Health Nutrition* 2013;12:1–14.
- [111] Musso G, Gambino R, De Micheli F, et al. Dietary habits and their relations to insulin resistance and postprandial lipemia in nonalcoholic steatohepatitis. *Hepatology* 2003;37:909–16.
- [112] Johnson RK, Appel LJ, Brands M, et al. Dietary sugars intake and cardiovascular health: a scientific statement from the American Heart Association. *Circulation* 2009;120:1011–20.
- [113] Welsh JA, Sharma AJ, Grellinger L, et al. Consumption of added sugars is decreasing in the United States. *American Journal of Clinical Nutrition* 2011;94:726–34.
- [114] Vos MB, Lavine JE. Dietary fructose in nonalcoholic fatty liver disease. *Hepatology* 2013;57:2525–31.
- [115] Abdelmalek MF, Suzuki A, Guy C, et al. Increased fructose consumption is associated with fibrosis severity in patients with nonalcoholic fatty liver disease. *Hepatology* 2010;51:1961–71.
- [116] Petta S, Marchesini G, Caracausi L, et al. Industrial, not fruit fructose intake is associated with the severity of liver fibrosis in genotype 1 chronic hepatitis C patients. *Journal of Hepatology* 2013;59:1169–76.
- [117] Nascimbeni F, Pais R, Bellentani S, et al. From NAFLD in clinical practice to answers from guidelines. *Journal of Hepatology* 2013;59:859–71.
- [118] Dunn W, Sanyal AJ, Brunt EM, et al. Modest alcohol consumption is associated with decreased prevalence of steatohepatitis in patients with non-alcoholic fatty liver disease (NAFLD). *Journal of Hepatology* 2012;57:384–91.
- [119] Reaven GM. The insulin resistance syndrome: definition and dietary approaches to treatment. *Annual Review of Nutrition* 2005;25:391–406.
- [120] Bugianesi E, Gastaldelli A, Vanni E, et al. Insulin resistance in non-diabetic patients with non-alcoholic fatty liver disease: sites and mechanisms. *Diabetologia* 2005;48:634–42.
- [121] Matthews DR, Hosker JP, Rudenski AS, et al. Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985;28:412–9.
- [122] Wallace TM, Levy JC, Matthews DR. Use and abuse of HOMA modeling. *Diabetes Care* 2004;27:1487–95.
- [123] Katz A, Nambi SS, Mather K, et al. Quantitative insulin sensitivity check index: a simple, accurate method for assessing insulin sensitivity in humans. *Journal of Clinical Endocrinology and Metabolism* 2000;85:2402–10.
- [124] Vanni E, Bugianesi E, Kotronen A, et al. From the metabolic syndrome to NAFLD or vice versa? *Digestive and Liver Disease* 2010;42:320–30.
- [125] Leite NC, Salles GF, Araujo AL, et al. Prevalence and associated factors of non-alcoholic fatty liver disease in patients with type-2 diabetes mellitus. *Liver International* 2009;29:113–9.
- [126] Prashanth M, Ganesh HK, Vima MV, et al. Prevalence of nonalcoholic fatty liver disease in patients with type 2 diabetes mellitus. *Journal of the Association of Physicians of India* 2009;57:205–10.
- [127] Assy N, Kaita K, Mymin D, et al. Fatty infiltration of liver in hyperlipidemic patients. *Digestive Diseases and Sciences* 2000;45:1929–34.
- [128] Müller MJ, Lagerpusch M, Enderle J, et al. Beyond the body mass index: tracking body composition in the pathogenesis of obesity and the metabolic syndrome. *Obesity Reviews* 2012;SE:6–13.
- [129] Xu C, Yu C, Ma H, et al. Prevalence and risk factors for the development of nonalcoholic fatty liver disease in a nonobese Chinese population: the Zhejiang Zhenhai Study. *American Journal of Gastroenterology* 2013;108:1299–304.
- [130] Younossi ZM, Stepanova M, Negro F, et al. Nonalcoholic fatty liver disease in lean individuals in the United States. *Medicine* 2012;91:319–27.
- [131] Kwon YM, Oh SW, Hwang SS, et al. Association of nonalcoholic fatty liver disease with components of metabolic syndrome according to body mass index in Korean adults. *American Journal of Gastroenterology* 2012;107:1852–8.
- [132] Sung KC, Kim BS, Cho YK, et al. Predicting incident fatty liver using simple cardio-metabolic risk factors at baseline. *BMC Gastroenterology* 2012;6:84.
- [133] Fassio E, Alvarez E, Dominguez N, et al. Natural history of non-alcoholic steatohepatitis: a longitudinal study of repeat liver biopsies. *Hepatology* 2004;40:820–6.
- [134] Bugianesi E, Manzini P, D'Antico S, et al. Relative contribution of iron burden, HFE mutations, and insulin resistance to fibrosis in nonalcoholic fatty liver. *Hepatology* 2004;39:179–87.
- [135] Marchesini G, Bugianesi E, Forlani G, et al. Nonalcoholic fatty liver, steatohepatitis, and the metabolic syndrome. *Hepatology* 2003;37:917–23.
- [136] Vanni E, Bugianesi E. Obesity and liver cancer. *Clinics in Liver Disease* 2014;18:191–203.
- [137] Lonardo A, Ballestri S, Marchesini G, et al. Non-alcoholic fatty liver disease: a precursor of the metabolic syndrome. *Digestive and Liver Disease* 2014. <http://dx.doi.org/10.1016/j.dld.2014.09.020>.
- [138] Anstee QM, Targher G, Day CP. Progression of NAFLD to diabetes mellitus, cardiovascular disease or cirrhosis. *Nature Reviews Gastroenterology and Hepatology* 2013;10:330–44.
- [139] Carulli L, Ballestri S, Lonardo A, et al. Is nonalcoholic steatohepatitis associated with a high-though-normal thyroid stimulating hormone level and lower cholesterol levels. *Internal and Emergency Medicine* 2013;8:297–305.
- [140] Pagadala MR, Zein CO, Dasarthy S, et al. Prevalence of hypothyroidism in non-alcoholic fatty liver disease. *Digestive Diseases and Sciences* 2012;57:528–34.
- [141] Liangpunsakul S, Chalasan N. Is hypothyroidism a risk factor for non-alcoholic steatohepatitis. *Journal of Clinical Gastroenterology* 2003;37:340–3.
- [142] Ittermann T, Haring R, Wallaschofski H, et al. Inverse association between serum free thyroxine levels and hepatic steatosis: results from the Study of Health in Pomerania. *Thyroid* 2012;22:568–74.
- [143] Chung GE, Kim D, Kim W, et al. Non-alcoholic fatty liver disease across the spectrum of hypothyroidism. *Journal of Hepatology* 2012;57:150–6.
- [144] Targher G, Day CP, Bonora E. Risk of cardiovascular disease in patients with nonalcoholic fatty liver disease. *New England Journal of Medicine* 2010;363:1341–50.
- [145] Lonardo A, Sookoian S, Chonchol M, et al. Cardiovascular and systemic risk in nonalcoholic fatty liver disease – atherosclerosis as a major player in the natural course of NAFLD. *Current Pharmaceutical Design* 2013;19:5177–92.
- [146] Lonardo A, Ballestri S, Targher G, et al. Diagnosis and management of cardiovascular risk in nonalcoholic fatty liver disease. *Expert Review of Gastroenterology and Hepatology* 2014;1–22.

- [147] Ballestri S, Lonardo A, Bonapace S, et al. Risk of cardiovascular, cardiac and arrhythmic complications in patients with non-alcoholic fatty liver disease. *World Journal of Gastroenterology* 2014;20:1724–45.
- [148] Byrne CD, Targher G. Ectopic fat, insulin resistance, and nonalcoholic fatty liver disease: implications for cardiovascular disease. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2014;34:1155–61.
- [149] Treppasertuk S, Leverage S, Adams LA, et al. The Framingham risk score and heart disease in nonalcoholic fatty liver disease. *Liver International* 2012;32:945–50.
- [150] Eichler K, Puhani MA, Steurer J, et al. Prediction of first coronary events with the Framingham score: a systematic review. *American Heart Journal* 2007;153:722–31.
- [151] Vuppalanchi R, Gould RJ, Wilson LA, et al. Nonalcoholic Steatohepatitis Clinical Research Network (NASH CRN). Clinical significance of serum autoantibodies in patients with NAFLD: results from the nonalcoholic steatohepatitis clinical research network. *Hepatology International* 2011.
- [152] McPherson S, Henderson E, Burt AD, et al. Serum immunoglobulin levels predict fibrosis in patients with non-alcoholic fatty liver disease. *Journal of Hepatology* 2014, <http://dx.doi.org/10.1016/j.jhep.2014.01.010>.
- [153] Petta S, Muratore C, Craxi A. Non-alcoholic fatty liver disease pathogenesis: the present and the future. *Digestive and Liver Disease* 2009;41:615–25.
- [154] Wree A, Broderick L, Canbay A, et al. From NAFLD to NASH to cirrhosis – new insights into disease mechanisms. *Nature Reviews Gastroenterology and Hepatology* 2013;10:627–36.
- [155] Gadd VL, Skoien R, Powell EE, et al. The portal inflammatory infiltrate and ductular reaction in human non-alcoholic fatty liver disease. *Hepatology* 2014;59:1393–405.
- [156] Miele L, Vallone S, Cefalo C, et al. Prevalence, characteristics and severity of non-alcoholic fatty liver disease in patients with chronic plaque psoriasis. *Journal of Hepatology* 2009;51:778–86.
- [157] Híndi M, Levy C, Couto CA, et al. Primary biliary cirrhosis is more severe in overweight patients. *Journal of Clinical Gastroenterology* 2013;47:e28–32.
- [158] Sorrentino P, Terracciano L, D'Angelo S, et al. Oxidative stress and steatosis are cofactors of liver injury in primary biliary cirrhosis. *Journal of Gastroenterology* 2010;45:1053–62.
- [159] Drebber U, Mueller JJ, Klein E, et al. Liver biopsy in primary biliary cirrhosis: clinicopathological data and stage. *Pathology International* 2009;59:546–54.
- [160] Abenavoli L, Luigiano C, Larussa T, et al. Liver steatosis in celiac disease: the open door. *Minerva Gastroenterologica e Dietologica* 2013;59:89–95.
- [161] Rojas-Feria M, Castro M, Suárez E, et al. Hepatobiliary manifestations in inflammatory bowel disease: the gut, the drugs and the liver. *World Journal of Gastroenterology* 2013;19:7327–40.
- [162] Kummen M, Schrupf E, Boberg KM. Liver abnormalities in bowel diseases. *Best Practice and Research. Clinical Gastroenterology* 2013;27:531–42.
- [163] Wieser V, Gerner R, Moschen AR, et al. Liver complications in inflammatory bowel diseases. *Digestive Diseases* 2013;31:233–8.
- [164] Bugianesi E, Salamone F, Negro F. The interaction of metabolic factors with HCV infection: does it matter? *Journal of Hepatology* 2012;56(S1):S56–65.
- [165] Lonardo A, Adinolfi LE, Loria P, et al. Steatosis and hepatitis C virus: mechanisms and significance for hepatic and extrahepatic disease. *Gastroenterology* 2004;126:586–97.
- [166] Rubbia-Brandt L, Quadri R, Abid K, et al. Hepatocyte steatosis is a cytopathic effect of hepatitis C virus genotype 3. *Journal of Hepatology* 2000;33:106–15.
- [167] Abid K, Paziienza V, De Gottardi A, et al. An in vitro model of hepatitis C virus genotype 3a-associated triglycerides accumulation. *Journal of Hepatology* 2005;42:744–51.
- [168] Piodi A, Chouteau P, Lerat H, et al. Morphological changes in intracellular lipid droplets induced by different hepatitis C virus genotype core sequences and relationship with steatosis. *Hepatology* 2008;48:16–27.
- [169] Mihm S, Fayyazi A, Hartmann H, et al. Analysis of histopathological manifestations of chronic hepatitis C virus infection with respect to virus genotype. *Hepatology* 1997;25:735–9.
- [170] Adinolfi LE, Gambardella M, Andreana A, et al. Steatosis accelerates the progression of liver damage of chronic hepatitis C patients and correlates with specific HCV genotype and visceral obesity. *Hepatology* 2001;33:1358–64.
- [171] Poynard T, Ratziu V, McHutchison J, et al. Effect of treatment with peginterferon or interferon alfa-2b and ribavirin on steatosis in patients infected with hepatitis C. *Hepatology* 2003;38:75–85.
- [172] Leandro G, Mangia A, Hui J, et al. HCV Meta-Analysis (on) Individual Patients' Data Study Group. Relationship between steatosis, inflammation, and fibrosis in chronic hepatitis C: a meta-analysis of individual patient data. *Gastroenterology* 2006;130:1636–42.
- [173] Shintani Y, Fujie H, Miyoshi H, et al. Hepatitis C virus infection and diabetes: direct involvement of the virus in the development of insulin resistance. *Gastroenterology* 2004;126:840–8.
- [174] Cua IH, Hui JM, Kench JG, et al. Genotype-specific interactions of insulin resistance, steatosis, and fibrosis in chronic hepatitis C. *Hepatology* 2008;48:723–31.
- [175] Romero-Gómez M, Del Mar Vilorio M, Andrade RJ, et al. Insulin resistance impairs sustained response rate to peginterferon plus ribavirin in chronic hepatitis C patients. *Gastroenterology* 2005;128:636–41.
- [176] Trepo E, Pradat P, Potthoff A, et al. Impact of pnp1a3 (rs738409 c>g) polymorphism on fibrosis progression and steatosis in chronic hepatitis C. *Hepatology* 2011;54:60–9.
- [177] Vanni E, Abate ML, Gentilcore E, et al. Sites and mechanisms of insulin resistance in nonobese, nondiabetic patients with chronic hepatitis C. *Hepatology* 2009;50:697–706.
- [178] Milner KL, van der Poorten D, Trenell M, et al. Chronic hepatitis C is associated with peripheral rather than hepatic insulin resistance. *Gastroenterology* 2010;138:932–41.e1–3.
- [179] Petta S, Rosso C, Leung R, et al. Effects of IL28B rs12979860 CC genotype on metabolic profile and sustained virologic response in patients with genotype 1 chronic hepatitis C. *Clinical Gastroenterology and Hepatology* 2013;11:311–7.e1.
- [180] Petta S, Amato M, Cabibi D, et al. Visceral adiposity index is associated with histological findings and high viral load in patients with chronic hepatitis C due to genotype 1. *Hepatology* 2010;52:1543–52.
- [181] Patton HM, Patel K, Behling C, et al. The impact of steatosis on disease progression and early and sustained treatment response in chronic hepatitis C patients. *Journal of Hepatology* 2004;40:484–90.
- [182] Fartoux L, Chazouilleres O, Wendum D, et al. Impact of steatosis on progression of fibrosis in patients with mild hepatitis C. *Hepatology* 2005;41:82–7.
- [183] Castera L, Hezode C, Roudot-Thoraval F, et al. Worsening of steatosis is an independent factor of fibrosis progression in untreated patients with chronic hepatitis C and paired liver biopsies. *Gut* 2003;52:288–92.
- [184] Pekow JR, Bhan AK, Zheng H, Chung RT. Hepatic steatosis is associated with increased frequency of hepatocellular carcinoma in patients with hepatitis C-related cirrhosis. *Cancer* 2007;109:2490–6.
- [185] Lerat H, Honda M, Beard MR, et al. Steatosis and liver cancer in transgenic mice expressing the structural and nonstructural proteins of hepatitis C virus. *Gastroenterology* 2001;122:352–65.
- [186] Rosario R, Ramakrishna B. Histopathological study of chronic hepatitis B and C: a comparison of two scoring systems. *Journal of Hepatology* 2003;38:223–9.
- [187] Gordon A, McLean CA, Pedersen JS, et al. Hepatic steatosis in chronic hepatitis B and C: predictors, distribution and effect on fibrosis. *Journal of Hepatology* 2005;43:38–44.
- [188] Peng D, Han Y, Ding H, Wei L. Hepatic steatosis in chronic hepatitis B patients is associated with metabolic factors more than viral factors. *Journal of Gastroenterology and Hepatology* 2008;23:1082–8.
- [189] Rastogi A, Sakhuja P, Kumar A, et al. Steatosis in chronic hepatitis B: prevalence and correlation with biochemical, histologic, viral, and metabolic parameters. *Indian Journal of Pathology and Microbiology* 2011;54:454–9.
- [190] Kumar M, Choudhury A, Manglik N, et al. Insulin resistance in chronic hepatitis B virus infection. *American Journal of Gastroenterology* 2009;104:76–82.
- [191] Park SH, Kim DJ, Lee HY. Insulin resistance is not associated with histologic severity in nondiabetic, noncirrhotic patients with chronic hepatitis B virus infection. *American Journal of Gastroenterology* 2009;104:1135–9.
- [192] Petta S, Cammà C, Di Marco V, et al. Hepatic steatosis and insulin resistance are associated with severe fibrosis in patients with chronic hepatitis caused by HBV or HCV infection. *Liver International* 2011;31:507–15.
- [193] Kim K, Kim KH, Kim HH, et al. Hepatitis B virus X protein induces lipogenic transcription factor SREBP1 and fatty acid synthase through the activation of nuclear receptor LXR alpha. *Biochemical Journal* 2008;416:219–30.
- [194] Kim K, Shin H, Kim K, et al. Hepatitis B virus X protein induces hepatic steatosis via transcriptional activation of SREBP1 and PPARγ. *Gastroenterology* 2007;132:1955–67.
- [195] Na T, Shin Y, Roh K, et al. Liver X receptor mediates hepatitis B virus X protein-induced lipogenesis in hepatitis B virus-associated hepatocellular carcinoma. *Hepatology* 2009.
- [196] Ruhl CE, Everhart JE. Relationship of non-alcoholic fatty liver disease with cholecystectomy in the US population. *American Journal of Gastroenterology* 2013;108:952–8.
- [197] Fracanzani AL, Valenti L, Russello M, et al. Gallstone disease is associated with more severe liver damage in patients with non-alcoholic fatty liver disease. *PLoS ONE* 2012;7:e41183.
- [198] Drenos F, Grossi E, Buscema M, et al. Networks in coronary heart disease genetics as a step towards systems epidemiology. *PLOS ONE* 2015;10:e0125876.