

# Hyperinsulinism and polycystic ovary syndrome (PCOS): role of insulin clearance

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Received: 5 May 2015 / Accepted: 24 July 2015 / Published online: 21 August 2015  
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## Abstract

**Purpose** Insulin resistance and compensatory hyperinsulinism are the predominant metabolic defects in polycystic ovary syndrome (PCOS). However, hyperinsulinism, as well as being compensatory, can also express a condition of reduced insulin clearance. Our aim was to evaluate the differences in insulin action and metabolism between women with PCOS (with normal glucose tolerance) and age- and BMI-matched women with prediabetes (without hyperandrogenism and ovulatory disorders).

**Methods** 22 women with PCOS and 21 age/BMI-matched women with prediabetes were subjected to a Hyperinsulinemic-euglycemic clamp and an Oral Glucose tolerance Test (OGTT). Insulin sensitivity was assessed by the glucose infusion rate during clamp (*M* value); insulin secretion by Insulinogenic index, Oral Disposition Index (DIo) and  $AUC_{2h-insulin}$  during OGTT; and insulin clearance by the metabolic clearance rate of insulin (MCRI) during clamp.

**Results** Women with PCOS showed significantly higher levels of  $AUC_{2h-insulin}$  ( $p < 0.011$ ), Insulinogenic Index ( $p < 0.001$ ), DIo ( $p = 0.002$ ) and significantly lower levels of  $AUC_{2h-glucose}$  ( $p = 0.001$ ). No difference was found between the two groups regarding insulin sensitivity (*M* value). Lower levels of MCRI were found in women with PCOS [420 (IQR 227–588) vs. 743 (IQR 597–888) ml m<sup>-2</sup> min<sup>-1</sup>;  $p < 0.001$ ]. Furthermore, in the PCOS group, a strong independent inverse correlation was

only observed between MCRI and  $AUC_{2h-insulin}$  (PCOS:  $\beta: -0.878$ ;  $p < 0.001$ ; Prediabetes:  $\beta: -0.501$ ;  $p = 0.019$ ).

**Conclusions** Our study suggests that in normoglycemic women with PCOS there is peripheral insulin sensitivity similar to that of women with prediabetes. What sets PCOS apart is the hyperinsulinism, today still simplistically defined “compensatory”; actually this is mainly related to decreased insulin clearance whose specific causes and dynamics have yet to be clarified.

**Keywords** PCOS · Insulin clearance · Clamp · Hyperinsulinism · Insulin resistance

## Introduction

Polycystic ovary syndrome (PCOS) represents an independent risk for the development of glucose intolerance states [1] and insulin resistance and compensatory hyperinsulinemia appear to be a central etiological characteristic in most women with PCOS [2–4], although it is not yet clear which of the two aspects plays a prominent role in the genesis of hyperandrogenism and ovulatory disorder. Furthermore, insulin resistance and hyperinsulinemia may represent two distinct features of the insulin disorder in PCOS [5]. In women with PCOS, as in the general population, the onset of impaired glucose tolerance or type 2 diabetes marks a failure of the pancreas to maintain this state of compensatory hyperinsulinemia [6, 7]. For this reason, in PCOS insulin secretion should always be examined in the context of peripheral insulin sensitivity rather than in isolation [8]. In the general population, the relationship between insulin secretion and sensitivity is a constant hyperbolic function [8, 9] that can be quantitated by the Disposition Index (DI) [9]. DI is highly heritable [10], associated with

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specific genetic loci [11, 12] and the most powerful predictor of diabetes risk [13]. Unfortunately, there is no “gold standard” method for the evaluation of this aspect, although DI with insulin secretion assessed by the acute insulin response to glucose (AIRg) after an iv glucose bolus and insulin sensitivity assessed by Hyperinsulinemic-euglycemic clamp [14] or frequently sampled iv glucose tolerance test [FSIGT] [13] have been shown to predict the future development of T2D. Therefore, detailed study of these aspects can only be made using sophisticated and/or expensive diagnostic methods.

Another important issue that should be considered is the meaning of insulin serum levels, both fasting and after glucose stimulation. The measurements of peripheral levels of insulin do not always reflect the prehepatic concentration of hormones and do not elucidate potential actions of the liver on glucose metabolism; in addition, there is also a known large inter- and intra-individual variability in hepatic insulin extraction [15–18]. Therefore, in insulin-resistant patients (such as women with PCOS), it is difficult to quantify not only “compensatory” pancreatic ability, but also the impact of hepatic, renal and tissue insulin clearance on peripheral serum insulin levels.

In this context, we evaluated the existence in women with PCOS of intrinsic decreased insulin clearance and/or intrinsic compensatory pancreatic capacity, at the base of the condition of hyperinsulinism that characterizes the syndrome. For this purpose, we evaluated the differences in insulin action and metabolism between women with PCOS (with normal glucose tolerance) and age- and BMI-matched women with prediabetes (without hyperandrogenism and ovulatory disorders).

## Materials and methods

Fifty premenopausal Caucasian women [25 with PCOS according to the Rotterdam criteria [19] (but with normal glucose tolerance) and 25 with prediabetes matched for age and BMI (with Impaired Glucose Tolerance and/or Impaired fasting Glucose, without hyperandrogenism and ovulatory disorders)] followed up in our Day Hospital of Endocrinology (2010–2014) were recruited and subjected to both a Hyperinsulinemic-euglycemic clamp and an Glucose tolerance Test (OGTT). Three women with PCOS were excluded from the study as well as four women with prediabetes, who at the OGTT showed overt Diabetes (glycaemia to 120 min of the test  $>11.1$  mmol/L). Twenty-two women with PCOS and 21 women with prediabetes were selected for the study. At screening (1 month prior to testing), standard diet and lifestyle advice were delivered and any treatment with metformin, insulin sensitizers, anti-androgens and hormonal contraceptives was suspended.

The following relevant data were obtained: family history of diabetes, oligo-amenorrhea, hirsutism, acne and age of menarche; weight, BMI, WC, blood pressure and Ferriman–Gallwey (FG) score (11 domains). Hirsutism was defined as FG score  $>8$  [20]. In women with suspected PCOS (irregular menses and/or hirsutism), hyperandrogenemia was diagnosed by serum total testosterone, Sex Hormone Binding Globulin (SHBG) during the follicular phase (Day 7 from the beginning of the last period) and Free Androgen Index (FAI) was calculated as the ratio of total testosterone levels in nmol/l to SHBG levels in nmol/l  $\times 100$  (%) [21]. Biochemical hyperandrogenism was diagnosed when total testosterone  $>2.84$  nmol/l, calculated on the basis of the 95th percentile upper limits of basal serum androgen normality in 144 healthy Sicilian eumenorrheal women without hirsutism and family history of PCOS (used as a control group in our previous study [22]). In women with PCOS transvaginal ovarian ultrasound scanning was performed between Days 5 and 10 after the beginning of the last period using a 7.5-MHz vaginal probe transducer (General Electric LOGIQ 400MD, Milwaukee, WI, USA). Both ovaries were measured in the sagittal, transverse and coronal planes. Ovaries were classified as polycystic if 12 or more follicles measuring 2–8 mm in diameter were present in each ovary, and/or there was an increase in ovarian volume ( $>10$  ml) [23].

On the same day, we also tested for total cholesterol, HDL cholesterol, LDL cholesterol and Triglycerides. Metabolic Syndrome (MetS) was diagnosed according to the National Cholesterol Education Program Adult Treatment Panel (NCEP-ATP) III definition [24] and diagnosis of diabetes mellitus according to the recommendations of the American Diabetes Association [25].

An OGTT was performed after a 12-h fast and blood samples were collected basally and after ingestion of 75 g glucose in 150 ml water within 30, 60, 90 and 120 min.

The Hyperinsulinemic-euglycemic clamp was performed under standard conditions [26], i.e. infusion of an insulin primer (Actrapid; Novo Nordisk, Bagsvaerd, Denmark) of  $160$  mU/m<sup>2</sup> body surface for the first 4 min of the test, followed by  $40$  mU/m<sup>2</sup> for the remaining 116 min; venous fasting blood samples were collected, analyzed and stored as appropriate after arterialization. The rate of peripheral glucose utilization (*M* value) (mg Kg<sup>-1</sup> min<sup>-1</sup>) was calculated by dividing the glucose amount infused during the last 40 min by body weight measured in kilograms (milligrams per kilogram per minute). Under the steady-state conditions of euglycemia, the glucose infusion rate equals glucose uptake by all the tissues in the body and is therefore a measure of tissue sensitivity to exogenous insulin. The metabolic clearance rate of insulin (MCRI) (ml m<sup>-2</sup> min<sup>-1</sup>) was calculated as the insulin infusion

rate divided by the steady-state plasma insulin level of the clamp [26].

$\beta$ -cell function was determined by fasting insulin, Insulinogenic Index (the ratio of the incremental insulin concentration to the incremental glucose concentration at the 30 min sample) [27] and the  $AUC_{2h\ Insulin}$  during OGTT. A composite indirect measure of  $\beta$ -cell function relative to insulin sensitivity, assessed by oral disposition index (DIo) [28], was calculated as  $(\Delta\text{Insulin}_{0-30}/\Delta\text{Glucose}_{0-30}) \times (1/\text{fasting insulin})$ . The trapezoidal method was used for the calculation of the areas under the curves for insulin ( $AUC_{2h\ Insulin}$ ), and glucose ( $AUC_{2h\ Glucose}$ ).

In all patients, we also calculated the visceral adiposity index (VAI), a sex-specific mathematical index based on anthropometric and metabolic parameters. VAI has been shown to be a useful tool for early detection of a condition of cardiometabolic risk before it develops into an overt metabolic syndrome [29], independently correlated with insulin sensitivity in various endocrine diseases characterized by a metabolic risk, in addition to PCOS [22], such as acromegaly [30] and prolactinoma [31]. The VAI was calculated as described [32] using the following sex-specific equations, where TG is triglyceride levels expressed in mmol/l and HDL is HDL-Cholesterol levels expressed in mmol/l:

$$\text{Females : VAI} = \left( \frac{\text{WC}}{36.58 + (1.89 \times \text{BMI})} \right) \times \left( \frac{\text{TG}}{0.81} \right) \times \left( \frac{1.52}{\text{HDL}} \right)$$

This study was approved by the Institutional Review Board at the Faculty of Medicine of the University of Palermo and the identity of the participants remained anonymous during data analysis. At the time of the first observation in our Day Hospital of Endocrinology, all women regularly signed an informed consent for the scientific use of their data.

## Assays

All hormones were measured in our laboratory using commercial kits. These included ELISA (DRG Diagnostics, DRG Instruments GmbH, Germany) for total testosterone (ng/ml; analytical sensitivity: 0.083 ng/ml; the intra- and interassay CVs were 3.28 and 4.73 %, respectively), and insulin (mUI/l; the intra- and interassay CVs were  $\leq 4$  and  $\leq 3.6$  %, respectively). Chemiluminescence assays were used for serum SHBG (nmol/l; Immulite, Diagnostic Products, Genoa, Italy; analytical sensitivity: 0.015 nmol/l; the intra- and interassay CVs were 5.50 and 6.20 %, respectively). Blood glucose levels (mg/dl) were measured using an electrochemical system (Glucocard, Menarini Diagnostics, Italy). Total cholesterol, HDL and triglycerides were measured in our laboratory using standard assays.

LDL cholesterol levels were calculated with Friedewald's formula. The conversion factors for the International System (SI) were the following: glucose (mg/dl vs. mmol/l: 0.0555), insulin (mUI/l vs. pmol/l: 6.945), total cholesterol (mg/dl vs. mmol/l: 0.0259), and total testosterone (ng/ml vs. nmol/l: 3.467).

## Statistical methods

The Statistical Packages for Social Sciences SPSS version 17 were used for the explorative data analysis. Normality of distribution for quantitative variables was assessed by the Shapiro–Wilk test. The quantitative variables not having a normal distribution were presented as median and interquartile Range (IQR); rates and proportions were calculated for categorical data. Differences between women with PCOS and women with Prediabetes in univariate analysis were detected by the Mann–Whitney *U* test for the continuous variables and by the  $\chi^2$ -test and Fisher's exact test (when appropriate) for categorical variables. Two linear regression models were performed to investigate the variables that independently correlate with  $AUC_{2h\ Insulin}$ . These variables were natural logarithmic transformed before being included in the regression models.

A *p* value of  $<0.05$  was considered statistically significant.

## Results

The prevalence of women with insulin resistance (according to the recently proposed *M* value cutoff of  $4.9\ \text{mg}\ \text{Kg}^{-1}\ \text{min}^{-1}$ ) [30] in the two groups was comparable: 19/21 (90.5 %) for women with prediabetes and 22/22 (100 %) for women with PCOS ( $p = 0.223$ ); indeed, women with PCOS showed a *M* value comparable to the women with prediabetes [median (IQR); 1.28 (0.85–1.62) vs. 1.42 (0.63–3.18)  $\text{mg}\ \text{Kg}^{-1}\ \text{min}^{-1}$ ;  $p = 0.789$ ]. Also, no significant differences were found between the two groups for all anthropometric parameters studied (BMI, WC, Body Surface Area and VAI) (Table 1).

Regarding the metabolic profile women with PCOS had significantly lower levels of fasting glucose [4.72 (4.38–5.18) vs. 6.16 (5.83–6.38) mmol/l;  $p < 0.001$ ],  $AUC_{2h\ Glucose}$  [700 (659–893) vs. 913 (813–1029)  $\text{mmol}\ \text{l}^{-1}\ 120\ \text{min}$ ;  $p = 0.001$ ], MCRI [420 (227–588) vs. 743 (597–888)  $\text{mmol}\ \text{l}^{-1}\ 120\ \text{min}$ ;  $p = 0.001$ ] and significantly higher levels of fasting insulin [132 (112.2–234) vs. 72.60 (54–108) pmol/l;  $p < 0.001$ ],  $AUC_{2h\ Insulin}$  [54594 (36828–118188) vs. 26730 (21060–35142)  $\text{pmol}\ \text{l}^{-1}\ 120\ \text{min}$ ;  $p < 0.001$ ], Insulinogenic index [218 (93.93–529) vs. 40.15 (19.42–71.42) pmol/mmol;  $p < 0.001$ ] and DIo [1.59 (0.70–2.40) vs. 0.53 (0.28–0.94)  $\text{mmol}^{-1}$ ;  $p = 0.002$ ] (Fig. 1).

**Table 1** Clinical and biochemical characteristics of women with PCOS (without glucose tolerance alterations) and women with prediabetes (women with impaired fasting glucose and/or impaired glucose tolerance and without PCOS)

	Women with PCOS <i>N</i> = 22 Median (IQR)	Women with prediabetes <i>N</i> = 21 Median (IQR)	<i>p</i>
Age	30 (17.75–37)	28 (15.50–39)	0.855
BMI (Kg/m <sup>2</sup> )	30.47 (26.01–42.51)	32.52 (24.05–40.65)	0.697
Waist circumference (cm)	99 (83.25–117)	103 (82–111)	0.679
Body surface area (m <sup>2</sup> )	1.90 (1.83–2.13)	1.96 (1.75–2.11)	0.942
Phenotype PCOS <sup>a</sup>	Subjects (%)	Subjects (%)	<i>p</i>
Complete phenotype	6 (27.3)		
Hyperandrogenism + polycystic ovary	7 (31.8)		
Oligo-anovulation + polycystic ovary	1 (4.5)		
Hyperandrogenism + oligo-anovulation	8 (36.4)		
	Subjects (%)	Subjects (%)	
Metabolic syndrome <sup>b</sup> (according to NCEP-ATP III criteria)	1 (4.5)	5 (23.8)	0.095
Diabetes or fasting glucose $\geq 5.6$ mmol/l	–	21 (100)	
High blood pressure	2 (9.1)	5 (23.8)	
High triglycerides	2 (9.1)	4 (19)	
Low HDL cholesterol	4 (18.2)	4 (19)	
Increased WC	14 (63.6)	16 (76.2)	
Family history for diabetes	15 (68.2)	13 (61.9)	0.666
Smoker or former smoker	7 (31.8)	6 (28.6)	0.817
	Median (IQR)	Median (IQR)	
Total testosterone (nmol/l)	2.63 (1.89–3.02)	1.89 (1.35–2.39)	0.004
SHBG (nmol/l)	46.50 (33.75–59)	54 (39–79.50)	0.215
FAI [100 × (total testosterone/SHBG)] (%)	5.49 (3.60–8.39)	3.52 (2.04–4.62)	0.004
	Metabolic profiles		
Fasting glucose (mmol/l)	4.72 (4.38–5.18)	6.16 (5.83–6.38)	<0.001
Fasting insulin (pmol/l)	132 (112.5–234)	72.60 (54–108)	<0.001
AUC <sub>2hglucose</sub> (mmol l <sup>-1</sup> 120 min)	700 (659–893)	913 (813–1029)	0.001
AUC <sub>2h Insulin</sub> (pmol l <sup>-1</sup> 120 min)	54,594 (36,828–118,188)	26,730 (21,060–35,142)	<0.001
Insulinogenic index ( $\Delta I_{30min}/(\Delta G_{30min})$ pmol/mmol)	218 (93.93–529)	40.15 (19.42–71.42)	<0.001
Oral disposition index (Dio) (mmol <sup>-1</sup> )	1.59 (0.70–2.40)	0.53 (0.28–0.94)	0.002
Glucose disposal rate during clamp ( <i>M</i> value) (mg Kg <sup>-1</sup> min <sup>-1</sup> )	1.28 (0.85–1.62)	1.42 (0.63–3.18)	0.789
Metabolic clearance rate of insulin (MCRI) (ml m <sup>-2</sup> min <sup>-1</sup> )	420 (227–588)	743 (597–888)	<0.001
Visceral adiposity index (VAI)	1.39 (0.93–1.67)	1.59 (1.14–2.64)	0.166
Total cholesterol (mmol/l)	4.33 (4.15–5.06)	4.70 (4.21–5.60)	0.172
HDL cholesterol (mmol/l)	1.42 (1.32–1.61)	1.52 (1.43–1.55)	0.348
Calculated LDL cholesterol (mmol/l)	2.18 (2.03–3.14)	2.80 (2.64–3.38)	0.077
Triglycerides (mmol/l)	1.02 (0.70–1.19)	1.40 (0.94–1.60)	0.016

Univariate analysis: qualitative variables were analyzed through  $\chi^2$  test or Fisher exact test; quantitative variables were analyzed through the Mann–Whitney *U* test

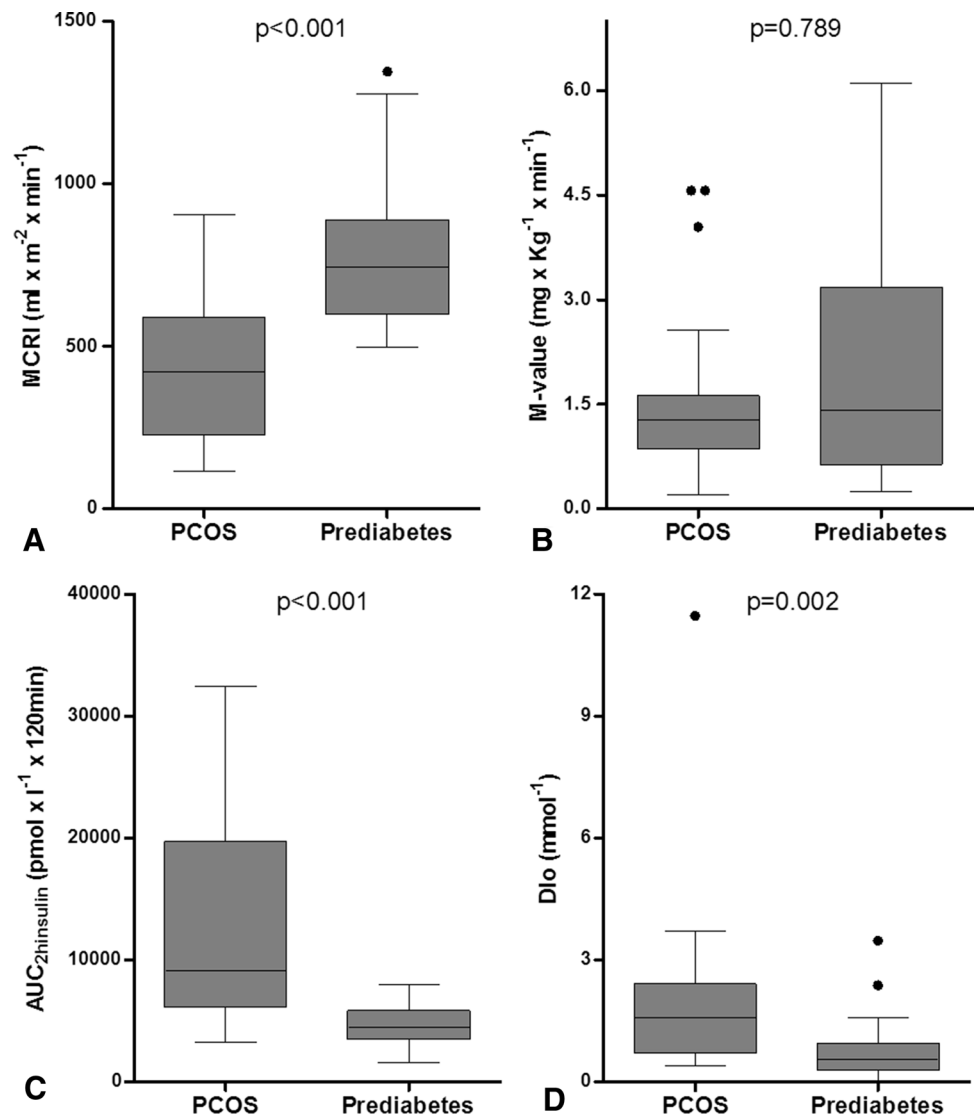
<sup>a</sup> Rotterdam criteria

<sup>b</sup> According to adult treatment panel (ATP) III criteria

At a multivariate analysis, we evaluated the variables that may potentially independently influence the AUC<sub>2hInsulin</sub>: age, BMI, WC, AUC<sub>2hGlucose</sub>, *M* value and MCRI. Only an

independent correlation was observed with the MCRI, more marked in women with PCOS (PCOS:  $\beta$ : -0.878; *p* < 0.001; Prediabetes:  $\beta$ : -0.501; *p* = 0.019) (Table 2; Fig. 2).

**Fig. 1** Differences in metabolic clearance rate of insulin (MCRI) **a** *M* value, **b** AUC<sub>2h</sub>-insulin, **c** and oral disposition index (Dio), **d** between women with PCOS and women with prediabetes



**Discussion**

In our study, using the gold standard for insulin resistance evaluation (Hyperinsulinemic-euglycemic clamp), we confirm that women with PCOS, despite being known to be more insulin-resistant than age-matched healthy women (regardless of BMI) [2, 31, 32], exhibit an insulin sensitivity comparable to that of the women with prediabetes and without hyperandrogenic and ovulatory disorder. This finding is also confirmed by the evidence of no difference in VAI, marker independently correlated with insulin sensitivity, between the two groups of patients. These data lead us to make some remarks on the real usefulness of the clamp in the assessment of insulin resistance of women with PCOS and on its real ability to provide information regarding all biological actions of insulin.

The clamp developed by DeFronzo et al. [26] in 1979 is a test based on the assumption that at high doses of insulin infused, the hyperinsulinemic state is sufficient to completely suppress hepatic glucose production and that there is no net change in glucose levels under steady-state conditions. Under such conditions, the rate of glucose infused is equal to the rate of whole-body glucose disposal (*M* value) and reflects the amount of exogenous glucose necessary to fully compensate for the hyperinsulinemia. Hence, the *M* value reflects only the aspect of the peripheral insulin-induced glucose disposal, without giving us any information about other biological actions of the hormone.

In women with PCOS the link between insulin resistance, hyperandrogenism, and ovulatory disorder is very complex: indeed, on the one hand, there is intrinsic insulin resistance characterized by insulin signaling abnormalities related to both the typical clinical features of PCOS and to common BMI-related abnormalities [33–35]. On the

**Table 2** Multivariate analysis of the hormonal, insulin-related and anthropometric parameters that independently correlate with  $AUC_{2h}$ 

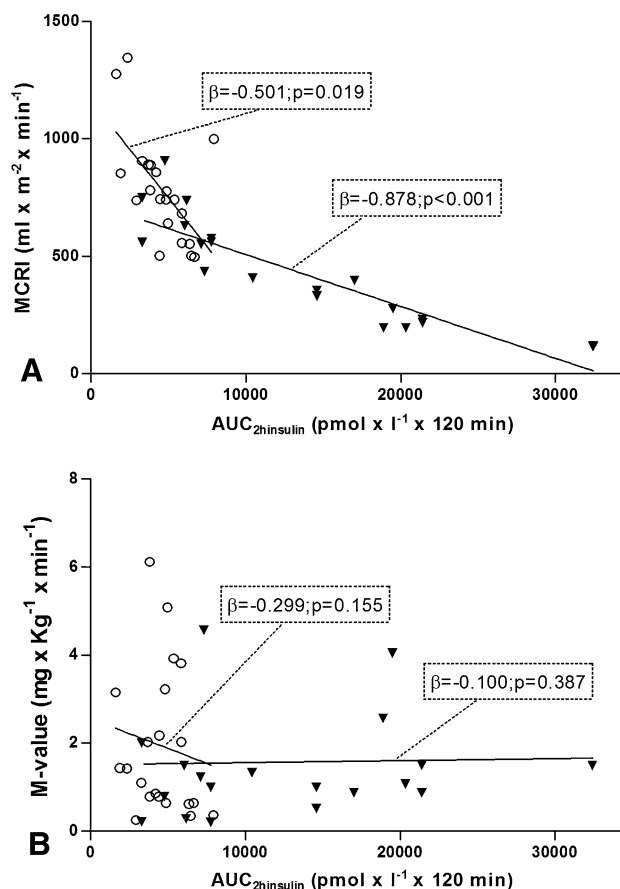
<i>Insulin</i>			
	Standardized coefficient $\beta$	Standard error	<i>p</i>
<b>Women with PCOS</b>			
Age	-0.014	95.31	0.904
BMI	0.493	260.35	0.070
Waist circumference	-0.337	127.10	0.224
$AUC_{2h\text{glucose}}$	-0.012	0.214	0.915
<i>M</i> value	-0.100	767.66	0.387
MCRI	-0.878	4.595	<0.001
<b>Women with prediabetes</b>			
Age	0.369	29.39	0.081
BMI	-0.524	60.32	0.170
Waist circumference	0.698	31.74	0.080
$AUC_{2h\text{glucose}}$	0.263	0.10	0.194
<i>M</i> value	-0.299	194.81	0.155
MCRI	-0.501	1.36	0.019

The natural logarithmic transformed values of the dependent variables included in the multiple linear regression model were used

other hand, the distinct molecular mechanisms of insulin in PCOS result in substantial hyperinsulinism causing hyperandrogenemia and anovulation [3, 4].

In our study, we showed that quantification of the glucose disposal rate during clamp is a technique that is not only complex, but also not very useful for understanding the pathophysiological mechanisms of the syndrome, we cannot define this evaluation useful to define the diabetes risk of a woman with PCOS, given that there are simple diagnostic alternatives derived from the OGTT that provide more specific information, such as  $AUC_{2h\text{insulin}}$  and the DIO. Indeed, women with prediabetes, while presenting a similar *M* value compared to women with PCOS, had a significantly lower DIO. This is because DIO values well reflect the known hyperbolic relationship existing between insulin sensitivity and pancreatic insulin response [28]. Our data agree with other evidence about the limited usefulness of *M* value evaluation and suggest that evaluation of insulin resistance alone does not fully characterize the PCOS population, since heterogeneity has been demonstrated in beta-cell activity, insulin clearance and peripheral insulin sensitivity in women with PCOS [5].

In our study, we observed that insulin levels (both fasting and after glucose stimulation), mainly in women with PCOS, were strongly influenced by the MCRI, although in women with PCOS we cannot exclude increased susceptibility to an exaggerated pancreatic response and/or increased susceptibility in women with prediabetes to a reduced pancreatic response: hyperinsulinemia can result from decreases in insulin clearance as well as from



**Fig. 2** Correlations between metabolic clearance rate of insulin (MCRI). **a** *M* value, **b** and  $AUC_{2h\text{insulin}}$  in women with PCOS (inverted triangle) and in women with prediabetes (open circle)

increases in insulin secretion [36, 37]. Also, our data partially contrast with the hypothesis that insulin clearance, being receptor-mediated, is usually decreased in insulin-resistant states [36]: women with PCOS had a lower MCRI despite a comparable *M* value to women with prediabetes.

Unfortunately, in our study we did not measure posthepatic insulin clearance; however, some evidence about normal posthepatic insulin clearance in PCOS would point to reduced hepatic clearance [38, 39].

Although there are studies in women with PCOS that have shown decreased hepatic extraction of insulin through an increase in circulating molar C-peptide/Insulin molar ratios [40, 41], we deemed it poorly useful to evaluate this molar ratio because it could be misleading; unfortunately, the assumption that the peripheral molar C-peptide/Insulin ratio can be used as a reflection of hepatic insulin extraction has not yet been experimentally validated [42, 43]. Anyway, other studies in hyperandrogenic women also lead us to hypothesize that decreased insulin clearance in PCOS is due mainly to increased insulin hepatic extraction [44, 45].

Considering the fact that most of the pathophysiological studies that have addressed these issues were carried out in the 1980–90s, it would be desirable to perform future new studies on larger series for also assessing these aspects according to PCOS phenotypes.

In conclusion, our study suggests that in women with PCOS there is a degree of insulin resistance similar to that found in other insulin resistance states, such as prediabetes. What sets PCOS apart is hyperinsulinism, today still simplistically defined “compensatory”; this is mainly related to decreased insulin clearance whose specific causes and dynamics have yet to be clarified. Further prospective studies could help to identify the most reliable indicator of diabetes risk and metabolic impairment in these patients.

**Acknowledgments** M. C. Amato—deceased.

#### Compliance with ethical standard

**Conflict of interest** The authors declare that they have no conflict of interest.

**Disclosure statement** The authors have nothing to disclose.

**Ethical approval** All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

**Informed consent** Informed consent was obtained from all individual participants included in the study.

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